

## Evaluation of Somaclones Peanut Plants Regenerated from Repeat Cycles of *In Vitro* Selection Against Drought Stress

A Farid Hemon<sup>1\*</sup> and Sudarsono<sup>2</sup>

<sup>1</sup>Agroecotechnology Study Program, Agriculture Faculty, Mataram University, NTB, Indonesia 83125

<sup>2</sup>Agronomy and Horticulture Department, Agriculture Faculty, Bogor Agricultural University (IPB), Jl. Meranti Kampus IPB Darmaga 16680, Indonesia

Received 22 December 2009/Accepted 22 March 2010

### ABSTRACT

The objective of this research was to evaluate the response of somaclonal peanut plants regenerated from repeated cycles of *in vitro* selection on medium containing 15% polyethylene glycol (PEG; w/v, corresponding to -0.41 Mpa osmotic potential) against drought stress. The R2 generation of peanut plants were used in this experiment with cv "Kelinci" and "Singa" as control cultivars. Drought treatment was the plants irrigated with water to field capacity (optimum condition) while other plants were grown under water deficit. Drought treatment was given at 16 to 85 days old peanut plants; after 85 days old, the plants were treated under optimum condition until plants were ready to harvest. Drought stress was measured using drought sensitivity index value (S) on scored parameters. Results of the experiment showed that peanut lines produced from repeated cycles of *in vitro* selection in medium containing 15% PEG were more tolerant to water deficit, had a better vegetative growth, a higher dry pod yield, and a lower dry pod yield reduction. This research demonstrated that repeated cycles of *in vitro* selection method was effective to produce drought tolerant peanut genotypes with a higher proline content than genotypes without *in vitro* selection.

Keywords : somaclonal variation, polyethylene glycol, drought tolerance

### INTRODUCTION

Water scarcity represents the most severe constraint to agriculture, account for most of potential yield losses. Drought has been one of the major causes of low yields in peanut production as well as in other crop production in the dry areas in Indonesia. The development of crop cultivars with increased tolerance to drought, both by conventional breeding methods and by *in vitro*, is an important strategy for agricultural production in the dryland. The use of drought-tolerant cultivars is a more practical and a cost-efficient approach in comparison to improving the cultural techniques and facilities for dryland farming.

*In vitro* selection is one of the methods that may be used to select drought tolerant plants. This method is based on the induction of genetic variation among cells, tissues and or organs in cultured and regenerated plants (Mohamed *et al.*, 2000). Genetic changes occurred during *in vitro* selection is called somaclonal variation.

Plants regenerated from somaclonal variation were superior than the control plants without *in vitro* selection, such as in wheat. R3 wheat regenerated from somaclonal variation had height of 60.9 cm, spike length of 10.2 cm and number of pods per spike 54.5, whereas control plants had height of 67.9 cm, spike length of 9.2 cm and number of pods per spike of 43.5 (Ivanov *et al.*, 1998).

Plant tissue culture leading to somaclonal variation has been considered as a rapid and reliable technique for improving crop production characteristics (Maralappanavar *et al.*, 2000). Induction of somaclonal variation followed by *in vitro* selection can identify plant variance with superior characteristics, such as drought tolerance in soybean (Widoretno and Sudarsono, 2004), and in rice (Adkins *et al.*, 1995).

The indicators of *in vitro* selection success are the efficiency to proliferate the desirable variant cells or tissues as well as to inhibit growth of unwanted cells or tissues using selection agent. PEG or Polyethylene glycol, non penetrating osmotic agent that lowers the water potential of the medium and has been used to simulate drought stress in plants, might be used in an *in vitro* selection to identify drought-tolerant individuals or mutant cells/tissues. PEG-induced osmotic potential in growth media has a major effect on the formation of embryo somatic in an *in vitro* culture. The decrease in water potential in the media due to PEG resulted in a decrease in tissue explant proliferation, shoot growth and regeneration (Tewary *et al.*, 2000). This decrease in proliferation was hypothesized to be a result of decreased endogenous polyamines in plant tissues. Polyamine has also been reported to affect morphogenesis, particularly in the formation of somatic embryo in *Picea glauca* (Kong *et al.*, 1998).

The use of 15% PEG in the media decreased the number of somatic embryo (SE) to 98.5% after only one

\* Corresponding author. e-mail : faridhemon\_1963@yahoo.com

selection cycle of three months on SE (Widoretno *et al.*, 2003). Most of selected soybean plants were still drought sensitive. The use of repeated cycles of *in vitro* selection was expected to increase the number of drought-resistant individual plants and tissues.

Repeated cycles of *in vitro* selection on PEG containing media have been conducted previously to obtain drought tolerant peanut cultivars (Hemon *et al.*, 2006). The next step of the research is to determine the tolerance level of these cultivars *in vivo*. Jain (2001) reported that several generations of somaclonal plants have to be assessed for their genetic stability and multiplication rate before they are developed into new cultivars. The objectives of this research was to evaluate the responses of somaclonal plants regenerated from repeated cycles of *in vitro* selection on PEG containing media towards drought, and to determine their endogenous proline levels.

## MATERIALS AND METHODS

### *Repeated In Vitro Selection, Regeneration and Seed Production of R1 Generation*

Embryogenic callus was established from peanut leaf embryo in the MS media (Murashige and Skoog, 1962) containing 2% sucrose, 8 g L<sup>-1</sup> agar, 0.1 mg L<sup>-1</sup> vitamins and amino acids (glycine, thiamin, pyridoxin and niacin), and 16 µM picloram (MS-P16 medium) in 25 mL medium for each culture.

Callus were then subcultured on the same medium with an addition of 15% PEG. All cultures were grown on a filter paper support in a liquid medium. PEG at 15% concentration was a sub-lethal concentration (Rahayu *et al.*, 2005). PEG at this level inhibited SE proliferation up to ≥ 95% compared to proliferation in the media without PEG.

Five hundred embryogenic callus (4000 SE) were evaluated during the first selection cycle. Each culture consists of five embryogenic callus (8-10 SE). Cultures were incubated at 26 °C in the dark and sub-cultured twice into fresh media during three months period. Embryogenic callus and SE developing from the PEG media were hence called Pi-I, i.e. the results of the first selection cycle, were multiplied in MS-P-16 media without PEG.

Half of Pi-I somatic embryo were subcultured to regenerate Pi-I plantlets, and another half were used as explants for the second selection. SE Pi-I were then reselected in media with 15% PEG before subcultured to a fresh media without PEG for three months. Surviving SE

and embryogenic callus from the second cycle were then proliferated in MS-P16 media without PEG. The surviving SE from the second cycle are called Pi-II. SE Pi-II were then regenerated to produce Pi-II plantlets.

Pi-I and Pi-II plantlets were then grown into R0 plants and were planted in plastic pots with mix of (pasteurised) soil and sand and maintained in the green house until plants were ready to harvest. Seeds from these plants were then replanted to produce R1 plants, which were then grown under the green house condition (as above).

### *Drought Tolerance Evaluation of Somaclonal Plants*

R2 plants were used in this evaluation (Table 1). Pi-0 population from cv “Kelinci” and “Singa” without *in vitro* selection were used as control plants. Seeds from R2 generation were planted and thinned to one plant per polybag.

Before applying the drought treatment, plants were watered daily to ± 90% field capacity until they were 15 days old. Field capacity was estimated by watering to media saturation. When plants were 16 days old the other half were exposed to drought treatment where as the other half were maintained under optimal condition (watering daily to field capacity). Drought treatment was applied for 69 days (i.e. terminated when plants were 85 days old) before transferred to optimal condition until the plants were ready to harvest. Watering of the drought-treated plants was conducted when leaves started wilting in 70% of plants.

Drought tolerance was measured using Drought Sensitivity Index (S) as described by Fischer and Maurer (1978). In summary,  $S = (1 - Y/Y_p) / (1 - X/X_p)$ , where (Y) = average of pod number, pod dry weight, root length, root dry weight of a drought-treated genotype; (Y<sub>p</sub>) = average of variables of optimally grown genotype; (X) = variable value of all drought-treated genotypes; and (X<sub>p</sub>) = variable values of all optimally grown genotype. Plants were classified as tolerant if having sensitivity Index of < 0.5, mildly tolerant if 0.5 ≤ S ≤ 1, and sensitive if S > 1.

### *Proline Content*

Proline content was measured on the second leaf from the tip when plants were 60 days after planting and had experienced six times of drought treatment. Proline content was analysed using methods developed by Bates *et al.* (1973).

Table 1. R2 generation of peanut genotypes following *in vitro* selection.

Cultivar	Plant population from SE selection	Number of genotypes	Genotype number
Singa	Pi-I	10	121-4, 121-1, 232-3, 124-3, 124-1, 123-4, 22-4, 232-1, 123-3, 22-2
	Pi-II	8	141-1, 32-4, 141-2, 82-1, 82-2, 12-2, 12-1, 32-3
Kelinci	Pi-I	12	13-3, 21-2, 12-3, 21-3, 11-2, 11-3, 14-4, 72-1, 12-2, 72-4, 14-2, 13-4
	Pi-II	10	32-4, 81-2, 32-2, 22-1, 11-2, 84-2, 84-4, 11-4, 81-4, 22-2

**RESULTS AND DISCUSSION**

*The Effect of Drought Stress on Plant Growth*

Drought tolerance was evaluated by comparing yields and yield components in optimal and drought stress conditions. Drought stress significantly reduced plant vegetative growth and pod yield (Table 2). Drought stress was reported to reduce number and dry weight of peanut

Pods, and this reduction was, at least partly, caused by a delay in gynophore initiation and elongation (Chapman *et al.*, 1993).

Plants under drought stress were significantly shorter than plants grown under optimal condition. Similarly, plants regenerated from SE selection under drought stress were shorter than control cultivars of “Singa” and “Kelinci”. The second cycle of SE selection, cv “Singa” had the shortest plants (Table 2).

Table 2. The effect of drought stress on plant growth Pi-0 (without in vitro selection) of R2 somaclone population generated from SE of cv “Singa” and “Kelinci” following the first selection cycle (Pi-I), or repeated selection cycles (Pi-II) of drought condition in vitro in PEG media

Plant population from SE selection	cv Singa		cv Kelinci	
	Optimum	Drought stress	Optimum	Drought stress
	Plant height (cm)			
Pi-0	70.61 aA	62.73 aB	68.97 aA	62.73 aB
Pi-I	70.50 aA	61.53 aB	65.21 aA	52.81 bB
Pi-II	65.28 aA	54.83 bB	67.28 aA	56.20 bA
	Pod dry weight (g)			
Pi-0	10.71 bA	7.30 cB	10.58 bA	7.21 cB
Pi-I	11.99 bA	9.57 bB	11.84 bA	9.16 bB
Pi-II	13.86 aA	11.75 aB	13.45 aA	10.96 aB
	Number of productive pods			
Pi-0	8.27 bA	5.73 cB	8.93 bA	6.47 cB
Pi-I	9.80 abA	7.97 bB	9.39 bA	7.86 bB
Pi-II	10.83 aA	9.50 aA	10.73 aA	9.50 aB
	Root length (cm)			
Pi-0	54.99 aA	46.00 aB	49.31 aA	35.93 aB
Pi-I	56.18 aA	45.73 aB	55.07 aA	38.22 aB
Pi-II	52.16 aA	49.79 aB	53.07 aA	39.33 aB
	Root dry weight (g)			
Pi-0	1.75 aA	0.96 cB	1.72 aA	0.81 bB
Pi-I	1.69 aA	1.24 bB	1.61 aA	0.98 bB
Pi-II	1.80 aA	1.55 aB	1.73 aA	1.21 aB

Notes: Values followed by same letters within a row are not significantly different by Duncan Multiple Range Test (P < 0.05)

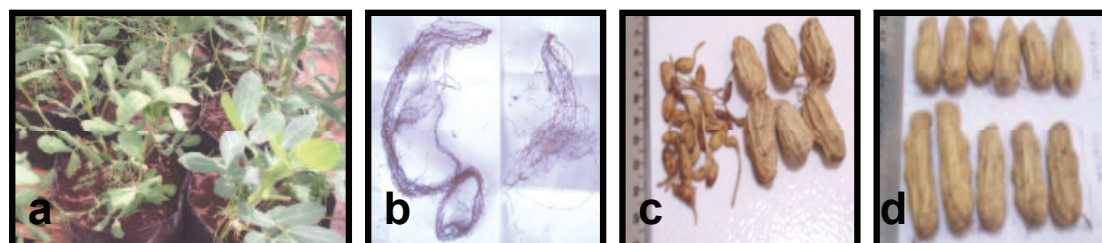


Figure 1. (a) wilting in a sensitive plant (left) and in a tolerant plant (right; has not started wilting) after exposed to the similar drought stress condition; (b) root growth of tolerant plants (left) and drought sensitive plans (right); (c) empty pods and non-productive pods of the sensitive plants; and (d) pod yields of drought tolerant plants

Plants of “Singa” and “Kelinci” regenerated from the second cycle of SE selection had longer roots and a greater root dry weight compared to those from other selection methods (Figure 1). Pod weight and number of dry pods in control plants were less than those in plants regenerated from *in vitro* selection for drought tolerance. Pod weight and number of dry pods obtained from the second cycle was greater than those from the first cycle. However, there was no significant difference between optimum and drought condition in term of number of pods in cv “Singa” (Table 2).

Plant growth was affected by the interaction between cultivars and selection cycles (data not shown). “Singa” and “Kelinci” plants from the second cycle had a greater pod dry weight, number of pods and root dry weight compared to the plants from the first cycle and from the control plants.

The greatest decrease in pod dry weight and number of empty pods following drought treatment were experienced by control plants of “Singa” and “Kelinci”. The percentage of decrease were 31.84% and 31.98% for pod dry weight, 30.71% and 27.55% for dry seed pods of “Singa” and “Kelinci” respectively. The plants from the second cycle had less significant decrease, i.e. 15.22% and 12.28% for pod dry weight, and 18.51% and 11.46% for dry seed pods, of “Singa” and “Kelinci” respectively (Table 3).

In general, plants from the second cycle had better vegetative growth, greater yield, and less decrease in dry pod weight and pod number following drought treatment compared to plants from the first cycle and control plants. The effects of drought treatment on plant’s physical condition, pod yields and root growth were presented in Figure 1.

Sensitive plants wilted earlier than tolerant plants following drought treatment. Tolerant plants regenerated

from SE selection might have a certain mechanism to withstand the drought condition, probably due to genetic changes occurred during *in vitro* selection. These changes resulted in an accumulation of mutant cells or tissues rendering them more tolerant to drought and had a better adaptation to media with 15% PEG. Previous studies demonstrated that plants from the second cycle had 15% more survived embryogenic callus than the plants from the first selection cycle (Hemon *et al.*, 2006).

*Plant Tolerance to Drought Stress*

Plant tolerance to drought was measured using Sensitivity Index (S) and the results are presented in Table 4 and 5. Based on calculation on S value on pod dry weight, number of pods, root length and root dry weight, S value for “Singa” ranged between 0.71–1.90, “Kelinci” 1.17–1.63. The plants regenerated from the first cycle had S value of between 0.72–1.05 for “Singa” and 1.00–1.33 for “Kelinci”. The plants regenerated from the second cycle had S value of between 0.20–0.74 for “Singa” and 0.75–1.12 for “Kelinci”.

Based on the S value on pod dry weight, “Singa” and “Kelinci” plants from the second selection cycle were classified as mildly tolerant with S value of 0.74 and 0.90, respectively, “Singa” plants from the first selection cycle were mildly tolerant ( S value of 0.97) and “Kelinci” were sensitive with S value of 1.10. These results indicated that it is possible to avoid yield loss and to maintain root growth under drought stress.

Not all lines showed increased tolerance to drought following repeated cycles of *in vitro* selection. Some lines even had S value of greater than the control. Out of 10 lines of “Singa” tested using repeated cycles of *in vitro* selection, three lines were classified as sensitive (30%), four lines as mildly tolerant, and three lines as tolerant, whereas out of 12 lines of “Kelinci” tested, six lines were classified as sensitive (50%), three lines (25%) as mildly tolerant, and three lines (25%) as tolerant. These results might have been caused by the random and spontaneous characteristics of somaclonal variation (Karp, 1995). Results from the second cycle of selection is presented in Table 5.

In addition, this selection method has reduced the number of sensitive lines with S value of > 1 after the second selection (Table 5). These results demonstrated that tolerance level to drought can be improved using this technique. Similar results were reported by Adkins *et al.* (1995) in rice and by Widoretono and Sudarsono (2004) in soybean. The use of PEG media was effective to select drought tolerant callus cells, which subsequently regenerated to develop lines with a higher tolerance to drought stress compared to the parental lines (Adkin *et al.*, 1995). Widoretono and Sudarsono (2004) reported that repeated selection in 15% PEG media was effective to produce mutant cells or mutant tissues of soybeans. These cells and tissues can then regenerated to develop drought tolerant mutant somaclones.

Qualitative variability of the R0 and R1 plants were found more on plants regenerated from the second cycle, i.e.

Table 3. Decrease percentage of weight and pod numbers of Pi-0 population (without *in vitro* selection) and of R2 somaclone population generated from SE of cv “Singa” and “Kelinci” following selection of drought condition *in vitro*

Plant population from SE and genotypes selection	Decrease percentage = $(1 - Y/Y_p) \times 100$	
	Pod dry weight (g)	Pod number
cv. Singa :		
Pi-0	31.84	30.71
Pi-I	20.18	18.67
Pi-II	15.22	12.28
cv. Kelinci :		
Pi-0	31.98	27.55
Pi-I	22.37	16.29
Pi-II	18.51	11.46

Notes : decrease percentage, where Y = dry weight of pod numbers under drought stress, Yp = pod dry weight of number of productive pods under optimal condition

Table 4. Drought Sensitivity Index of various agronomical characters of Pi-0 population (without *in vitro* selection) and of R2 somaclone population generated from SE of cv “Singa” and “Kelinci” following selection of drought condition *in vitro*

Plant population from SE and genotypes selection	Drought sensitivity index (S)				Somaclones phenotypes
	Pod dry weight (g)	Pod number	Root length (cm)	Root dry weight (g)	
cv. Singa :					
Pi-0	1.55	1.90	0.71	1.22	Sensitive
Pi-I	0.97	1.05	0.81	0.72	Mildly Tolerant
Pi-II	0.74	0.73	0.20	0.38	Mildly tolerant
cv. Kelinci :					
Pi-0	1.57	1.63	1.17	1.42	Sensitive
Pi-I	1.10	1.00	1.33	1.05	Sensitive
Pi-II	0.90	0.75	1.12	0.81	Mildly tolerant

Notes : Drought sensitivity index (S) and somaclones phenotypes based on pod dry weight

Table 5. Drought Sensitivity Index (based on dry pod weight and pod number) of Pi-0 population (without *in vitro* selection) and in R2 somaclone population generated from SE of cv “Singa” and “Kelinci” following selection of drought condition *in vitro*

Cultivars or Genotypes	Drought sensitivity index (S)		Somaclones phenotypes
	Pod dry weight (g)	Pod number	
cv. Singa :			
Pi-0 :	1.55	1.90	Sensitive
Pi-I :			
121-4	2.04	2.94	Sensitive
121-1	2.00	2.15	Sensitive
232-3	0.87	0.79	Mildly tolerant
124-3	0.35	0.20	Tolerant
124-1	0.71	1.13	Mildly tolerant
123-4	0.51	0.16	Mildly tolerant
22-4	0.08	0.59	Tolerant
232-1	2.10	2.05	Sensitive
123-3	0.74	0.88	Mildly tolerant
22-4	0.25	-0.38	Tolerant
Pi-II :			
141-1	0.18	-0.17	Tolerant
32-4	0.76	0.94	Mildly tolerant
141-2	0.45	0.54	Tolerant
82-1	0.56	0.40	Mildly tolerant
82-2	1.33	1.77	Sensitive
12-2	0.70	0.59	Mildly tolerant
12-1	0.80	0.94	Mildly tolerant
32-3	1.17	0.86	Sensitive



Tabel 5. Lanjutan

Cultivars or Genotypes	Drought sensitivity index (S)		Somaclones phenotypes
	Pod dry weight (g)	Pod number	
cv. Kelinci :			
Pi-0 :	1.57	1.63	Sensitive
Pi-I :			
13-3	2.12	2.17	Sensitive
21-2	1.83	1.77	Sensitive
12-3	1.40	1.93	Sensitive
21-3	0.92	1.03	Mildly tolerant
11-2	0.67	0.23	Mildly tolerant
11-3	0.88	-0.40	Mildly tolerant
14-4	-0.13	0.44	Tolerant
72-1	2.32	1.52	Sensitive
12-2	1.55	1.10	Sensitive
72-4	1.11	1.13	Sensitive
14-2	0.39	0.49	Tolerant
13-4	0.44	0.98	Tolerant
Pi-II :			
32-4	2.40	2.58	Sensitive
81-2	0.51	0.40	Mildly tolerant
32-2	0.43	0.00	Tolerant
22-1	0.26	0.00	Tolerant
11-2	1.02	1.27	Sensitive
84-2	0.92	0.71	Mildly tolerant
84-4	0.73	0.39	Mildly tolerant
81-4	0.03	-0.67	Tolerant
22-2	0.61	0.00	Mildly tolerant

Notes: Drought sensitivity index (S) and somaclones phenotypes based on pod dry weight

nine qualitative characters with frequency of 5–55%. These results provided a further proof that the chance of obtaining tolerant lines were greater in plants regenerated from the repeated selection (the second cycle) rather than the single selection only (Hemon *et al.*, 2006).

*Proline Content of Drought-treated Somaclonal Plants*

Proline has an important role in plant tolerance to drought (Watanabe *et al.*, 2000). Increased tolerance to drought resulted in accumulation of compounds protecting

Table 6. Proline and sugar content of Pi-0 population (without *in vitro* selection) and of R2 somaclone population generated from SE of cv “Singa” and “Kelinci” following selection of drought condition *in vitro*

Population of SE selection	cv. Singa			cv. Kelinci		
	Optimum	Drought	Increase (%)	Optimum	Drought	Increase (%)
Proline [ $\mu\text{g (g dry weight)}^{-1}$ ]						
Pi-0	3010.5 cA	4119.3 bA	36.8	2709.5 cA	3380.7 cA	24.8
Pi-I	4766.7 bB	7181.1 aA	50.7	4447.5 bA	5653.0 bA	27.1
Pi-II	7217.0 aA	8608.0 aA	19.3	6996.4 aA	7941.0 aA	13.5

Notes: Values followed by same letters within a row are not significantly different by Duncan Multiple Range Test ( P < 0.05)

cells from damages caused by low water potential (Jensen *et al.*, 1996). Increase in drought-tolerance also increased endogenous proline content (Table 6). SE selection using 15% PEG significantly increased proline content at both optimal and non-optimal/drought condition. Plants regenerated from the second cycle of cv “Singa” and “Kelinci” had a higher endogenous proline content than those from the first cycle. In addition, the increase in proline content of plants regenerated from the second cycle was lower than control plants.

### CONCLUSION

Somaclones ( $R_2$ ) regenerated from repeated cycles of *in vitro* culture on media supplemented with 15% PEG had a better vegetative growth with a higher pod yield and a less decrease of pod yield compared to  $R_1$  plants, or to control plants (without *in vitro* selection).

Somaclones ( $R_2$ ) regenerated from SE cv “Singa” and cv “Kelinci” had a better drought tolerance index with pod dry weight of 11.75 and 10.96 g plant<sup>-1</sup>, respectively. The second cycle of SE selection in cv “Kelinci” produced a greater number of drought-tolerant individuals than those produced in the first cycle. In addition, repeated cycles of *in vitro* selection resulted in plants with a higher proline content.

### REFERENCES

- Adkins, S.W., R. Kunanuvatshidah, I.D. Godwin. 1995. Somaclonal variation in rice drought tolerance and other agronomic characters. *Aust. J. Bot.* 43:201-209.
- Bates, L.S., R.P. Waldren, I.D. Teare. 1973. Rapid determination of free proline for water stress studies. *Plant and Soil* 39:205-207.
- Chapman, S.C., M.M. Ludlow, F.P.C. Blamey, K.S. Fisher. 1993. Effect of drought at pod filling on utilization of water and growth of cultivars of groundnut. *Field Crop Res.* 32:243-255.
- Fischer, R.A., R. Maurer. 1978. Drought stress in spring wheat cultivars : I. Grain yield responses. *Aust. J. Agric. Res.* 29:897-912.
- Hemon, A.F., L. Ujianto, Sudarsono. 2006. Seleksi berulang dan identifikasi embrio somatik kacang tanah yang insensitif polietilena glikol (PEG) dan filtrat kultur *Sclerotium rolfsii*. *Agroteksos* 16:21-32.
- Ivanov, P., Z. Atanassov, V. Milkoca, L. Nikolova. 1998. Culture selected somaclonal in five *Triticum aestivum* L., genotypes. *Euphytica* 104:167-172.
- Jain, S.M. 2001. Tissue culture-derived variation in crop improvement. *Euphytica* 118 :153-156.
- Jensen, A.B., P.K. Busk, M. Figueras, M.M. Alba, G. Peraccia, R. Messeguer, A. Goday, A.M. Pages. 1996. Drought signal transduction in plant. *Plant Growth Reg.* 20:105-110.
- Karp, A. 1995. Somaclonal variation as a tool for crop improvement. Kluwer Acad. Publ. The Netherlands.
- Kong, L., S.M. Attree, L.C. Fowke. 1998. Effect of polyethylene glycol and methylglyoxal (guanyldihydrazone) on endogenous polyamine levels and somatic embryo maturation in white spruce (*Picea glauca*). *Plant Sci.* 133:211-220.
- Maralappanavar, M.S., M.S. Kuruvina shetti, C.C. Harti. 2000. Regeneration, establishment and evaluation of somaclones in *Sorghum bicolor* (L.) Moench. *Euphytica* 115:173-180.
- Mohamed, M.A.H., P.J.C. Harris, J. Henderson. 2000. *In-vitro* selection and characterisation of a drought tolerant clone of *Tagetes minuta*. *Plant Sci.* 159:213-222.
- Murashige, T., F. Skoog. 1962. A revised media for rapid growth and bioassay with tobacco tissue cultures. *Physiol. Plant* 15:473-493.
- Rahayu, E.S., E. Guhardja, S. Ilyas, Sudarsono. 2005. Seleksi *in-vitro* embrio somatik kacang tanah pada media dengan polietilena glikol untuk mensimulasikan cekaman kekeringan. *Majalah Ilmiah Biologi BIOSFERA*.
- Tewary, P.K., A. Sharma, M.K. Raghunath, A. Sarkar. 2000. *In-vitro* response of promising mulberry (*Morus sp.*) genotypes for tolerance to salt and osmotic stresses. *Plant Growth Reg.* 30:17-21.
- Watanabe, S., K. Kojima, Y. Ide, S. Sasaki. 2000. Effect of saline and osmotic stress on proline and sugar accumulation in *Populus euphratica in-vitro*. *Plant Cell Tissue Organ Cult.* 63:199-206.
- Widoretno, W., E. Guhardja, S. Ilyas, Sudarsono. 2003. Reaksi embrio somatik kedelai terhadap polietilena glikol dan penggunaannya untuk seleksi *in-vitro* terhadap cekaman kekeringan. *Hayati* 10:134-139.
- Widoretno, W., Sudarsono. 2004. Evaluasi galur kedelai varian somaklonal hasil seleksi *in-vitro* terhadap stres kekeringan. *Hayati* 11:11-20.