

The Effect of Visible Light Cure (VLC) Exposure to Gingival Tissue's *Sprague dawley* Rats

Kwartarini Murdiastuti
Suryono
Aini Moeljono
Mefi Priba Sari
Rani Gamawati

*Department of Periodontology, Faculty of Dentistry,
Universitas Gadjah Mada, Yogyakarta, Indonesia*

E-mail: kmurdiastuti@yahoo.com
Received August 15, 2010; Accepted December 10, 2010

Abstract

Visible Light Cure (VLC) is a blue light used in dentistry as an activator for restorative material and fixed orthodontic bonding. The wavelength of VLC is between 400-500 nm and considered non-ionizing radiation that can produce free radicals. According to previous research, the light at wavelength < 500 nm could inhibit cells mitosis, cause cells damage, and reduce cells growth and inflammation. The purpose of this study was to investigate the effect of VLC exposure on gingival epithelial thickness, total neutrophil and macrophage count of gingival connective tissue of *Sprague dawley* rats. The subjects of this study consisted of 20 *Sprague dawley* rats, in 2-3 months of age and divided into 4 groups. Each group was 5 rats. The rats in each group were sacrificed before (0 day, as group A) and after 1st (group B), 3rd (group C), 5th (group D) day of VLC exposure, respectively. The exposure of VLC was done in labial aspect of cervical anterior teeth of mandible. The distance of exposure was as thick as 2 layers of celluloid strip and the histological specimens were stained by Hematoxylin Eosin. Each specimen was measured for its gingival epithelial thickness by using a micrometer and the number of neutrophil and macrophage were counted. The data of gingival epithelial thickness from 4 groups were analyzed by Kruskal Wallis. The number of neutrophil and macrophage were analyzed by using one way ANOVA. The results of this study showed that there were significant differences among groups on the thickness of gingival epithelial, the number of neutrophil and macrophage in the gingival connective tissue of *Sprague dawley* rats. The result of this study indicated that VLC exposure might decrease the thickness of gingival epithelial but increase the number of neutrophil and macrophage of gingival connective tissue of *Sprague dawley* rats.

Keywords: Visible Light Cure, radiation, epithelial thickness, neutrophil, macrophage.

Introduction

In dentistry curing unit is commonly used when restoring teeth using composite resin materials. The device produces Visible Light Cure (VLC) to initiate the polymerization of composite resin. Visible Light

Cure is a light with 400-500 nm wavelength in the blue region of the visible light spectrum¹.

Based on its wave length, VLC belongs to nonionizing radiation which is classified into optical radiation. Radiation exposure can cause damage at the level of molecules, cells, tissues, or organs.

Radiation induces cell damage and even death due to the mechanism of cell mitosis disturbance².

Nowadays the use of composite resin polymerized by visible light is getting more popular, as well as replacing composite resin polymerized by chemical agent or ultra violet. Along with that condition, the current dental practices have been using VLC³. Meanwhile, Alatas and Lusiyananti stated that visible blue light exposure (400-550 nm) can induce retinal damage, known as blue-light retinal injury².

Gingiva is one of periodontal tissues that surrounding the teeth and alveolar bone and expanding to mucogingival. This structure attaches to the teeth and alveolar bone. The function of gingiva is to protect the subgingival tissue from oral cavity environment. Histologically, gingiva consists of gingival epithelial layers and gingival connective tissue⁴. Blood vessel supplies of gingival connective tissue are formed by arterioles plexus, capillaries, and small veins that extend from the sulcular epithelial toward the outer surface of the gingiva⁵.

The uses of VLC in dental practice may influence the gingival tissue. VLC exposure can cause cell damage and reduce cell growth. In this case, the gingival epithelial cells exposed by VLC possibly undergo structural alteration.

Radiation of VLC exposure causes acute inflammation⁶. Inflammation is an essential process that is critical to host defense. Meanwhile, acute inflammation implies an inflammatory reaction that develops over a rapid time scale (hours to days). Acute inflammation can provide a clean-up function that segues to tissue repair⁷. Acute inflammation response includes the process of vascular changes and blood flow, capillary permeability increases, and formation of cellular components through the migration of leukocytes into the extravascular tissues⁸. Only two types of leukocytes involve in acute inflammation those are neutrophil and macrophage⁹.

The purpose of this study was to investigate the effect of VLC exposure on gingival epithelial thickness, total neutrophil and macrophage count of gingival connective tissue of *Sprague dawley* rats.

Materials and Methods

The subjects of this study were 20 *Sprague dawley* rats, 2-3 months of age, which were divided into 4 groups. Rats in group A were sacrificed before VLC exposure (day 0), while in group B, C and D, rats were sacrificed on 1st, 3rd and 5th day after VLC exposure respectively. The exposure of VLC was done for 30 seconds and the distance was as thick as 2 layers of celluloid strip from labial cervical of mandible anterior teeth.

Histological specimens were stained with Hematoxylin Eosin and observed using 400x magnification of light microscope. The thickness of gingival epithelial was measured using a micrometer from the basal to the cornified layer in 10 fields of view for each specimen. For the number of neutrophil and macrophage, specimens were observed in 10 fields of view as well. The average thickness of gingival epithelial was analyzed by Kruskal Wallis and Mann-Whitney test. However the average number of neutrophil and macrophage were analyzed by one way ANOVA.

Results

The highest average of gingival epithelial thickness (80.5 μ m) and the average of neutrophil and macrophage numbers (2.8 and 1.8, respectively) were found before VLC exposure as shown in Figure 1.

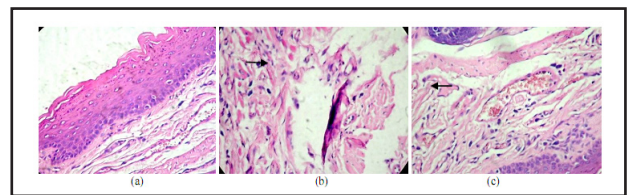


Figure 1. Hematoxylin Eosin staining of the gingival tissue of *Sprague dawley* rats in group A, before VLC exposure (0 day), (a) The thickness of gingival epithelial; (b) the number of neutrophil (arrows); (c) the number of macrophage (arrows) (400x Magnification).

Figure 2 shows histological specimens at 1st day after VLC exposure. A decrease on average of gingival epithelial thickness to be 77.7 μ m was found. Meanwhile, the average of neutrophil and macrophage number increased to 11.8 and 2.2 respectively. Decrease of gingival epithelial thickness average to 76.4 μ m was found 3rd day after VLC exposure as shown in Figure 3. However the average of neutrophil number decreased to 7.00, whereas the average of macrophage number increased to 6.2.

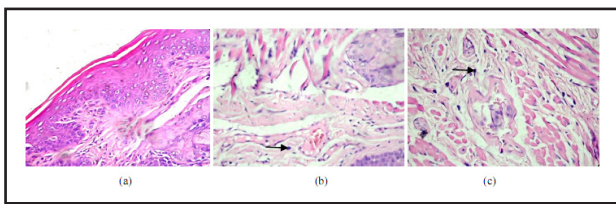


Figure 2. Hematoxylin Eosin staining of the gingival tissue of *Sprague dawley* rats in the group B, the 1st day after VLC exposure, (a) the thickness of gingival epithelial; (b) the number of neutrophil (arrows); (c) the number of macrophage (arrows) (400x Magnification).

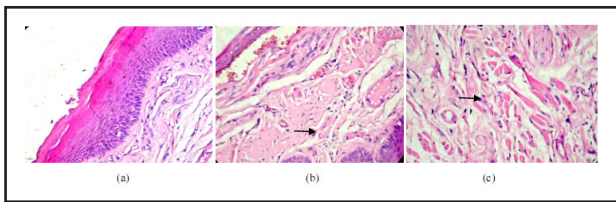


Figure 3. Hematoxylin Eosin staining of the gingival tissue's *Sprague dawley* rats in the group C, the 3rd day after VLC exposure, (a) the thickness of gingival epithelial; (b) the number of neutrophil (arrows); (c) the number of macrophage (arrows) (400x Magnification).

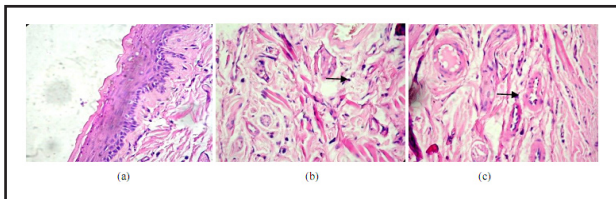


Figure 4. Hematoxylin Eosin staining of the gingival tissue's *Sprague dawley* rats in the group D, the 5th day after VLC exposure, (a) the thickness of gingival epithelial; (b) the number of neutrophil (arrows); (c) the number of macrophage (arrows) (400x Magnification).

The lowest average of gingival epithelial thickness of 72.95 μ m was found at 5th day after VLC exposure, shown in Figure 4. In the other hand, the average of neutrophil number was not different from 3rd to 5th day after VLC exposure. However the average of macrophage number declined by 2.4.

Table 1. Mean and standard deviation of gingival epithelial thickness, neutrophil and macrophage number in gingival tissue's *Sprague dawley* rats.

Group	Epithelial thickness (μ m)	Neutrophil number	Macro-phage number
0 day (A)	80.50 \pm 1.94	2.80 \pm 0.84	1.8 \pm 0.84
1 st day (B)	77.70 \pm 0.54	11.8 \pm 3.11	2.2 \pm 0.84
3 rd day (C)	76.40 \pm 0.60	7.00 \pm 1.58	6.2 \pm 0.84
5 th day (D)	72.95 \pm 1.08	2.80 \pm 1.30	2.4 \pm 1.14

Figure 5 showed increasing gingival epithelial thickness from 0 to 5th day after VLC exposure. The average differences between groups B and C was 1.3 μ m, and group C to D was 3.45 μ m. Data on the thickness of epithelial gingival was not homogeneously distributed, thus Kruskal Wallis test was conducted and the results showed p=0.001. It means VLC exposure effected epithelial thickness (p<0.05).

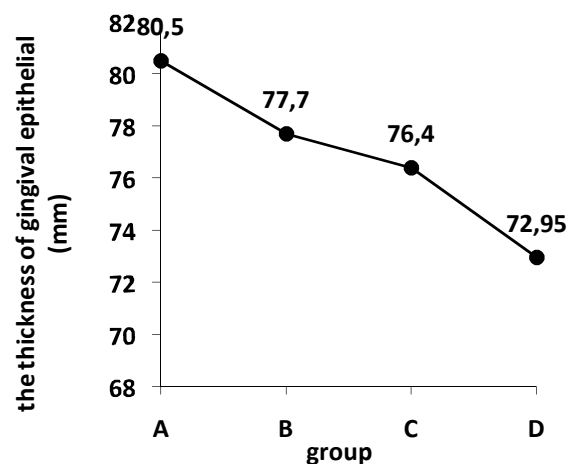


Figure 5. The thickness of gingival epithelial *Sprague dawley* rats in each group.

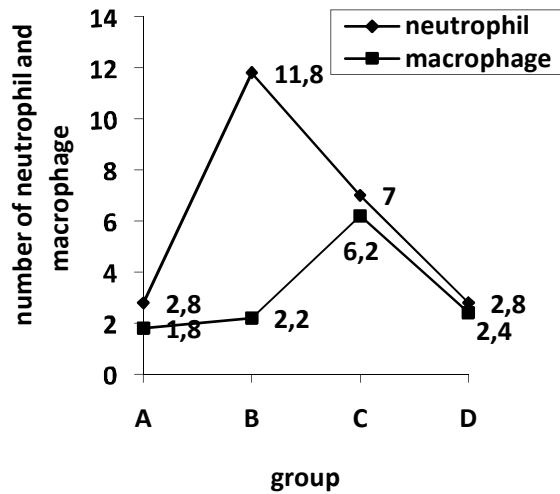


Figure 6. The number of neutrophil and macrophage of gingival connective tissue of *Sprague dawley* rats in each group.

Fluctuation on neutrophil and macrophage number from 0 to 5th day is shown in Figure 6. After increasing from 0 to 1st day, neutrophil number decreased up to 5th day. Observation on macrophage number showed increasing results from 0 to 3rd day and decreasing until 5th day. One way ANOVA test was applied on neutrophil and macrophage number of connective tissue of *Sprague dawley* rat. One way ANOVA test results in Table II and III showed $p < 0.05$, thus demonstrated significant differences among groups on the average of neutrophil and macrophage number upon VLC exposure on gingival connective tissue *Sprague dawley* rats, respectively.

Table 2. One Way ANOVA results on neutrophil number of gingival connective tissue of *Sprague dawley* rats.

Source	Sum of Squares	df	Mean Square	F ratio	sig.
Between groups	275.400	3	91.800	25.151	.000
Within groups	58.400	16	3.650		
Total	333.800	19			

df: degree of freedom; sig.: significance.

Table 3. One Way ANOVA results on macrophage number of gingival connective tissue of *Sprague dawley* rats.

Source	Sum of Squares	df	Mean Square	F ratio	sig.
Between groups	62.950	3	20.983	24.686	.000
Within groups	13.600	16	.850		
Total	76.550	19			

df: degree of freedom; sig.: significance.

Discussion

Visible Light Cure (VLC), a blue light with a wavelength between 400-500 nm¹, can be classified as visible light radiation or non-ionizing radiation². It can cause thermal and photochemical reactions¹⁰. This study results showed that VLC not only decreases the average of epithelial gingival thickness but also increases the average of neutrophil and macrophage number. Those were presumably because VLC exposure interfere the epithelial cell itself and the process of turnover.

Disruption of epithelial cells by VLC exposure occurs in mitochondria, which is the center of the cell metabolism. Cell metabolism requires factor of electron carriers, such as flavins and cytochrom¹¹. Visible Light Cure absorbed by flavins causes decreasing SDH (succinic dehydrogenase) activity¹⁰. Succinic dehydrogenase is oxidation catalyze of fumarat in the Kreb's cycle. Succinic dehydrogenase disturbance decreases the production of ATP which is the energy for cell metabolism. Visible light cure absorbed by cytochrome forms excess of Reactive Oxygen Species (ROS)¹². However, ROS is a molecule with free valence electrons that can cause cell damage¹³.

In response to tissue damage, acute inflammation reaction will occur in early stages of inflammation such as capillary vasodilation and increasing blood flow and capillary permeability¹⁴. This process is stimulated by inflammatory mediators, such as histamin, serotonin, leukotrien, bradikinin, and prostaglandin⁴.

After blood flow decreases, leucocytes move closer to the vessel wall. Begins with neutrophil then monocytes move aside and adhere to the wall. This will be followed by an active amoeboid movement of the cells into perivascular tissue through gaps between endothelial cells. After neutrophil extravasation, monocytes differentiate into macrophages and move toward areas of inflammation in a chemotactic way, thus accumulation of neutrophil followed by macrophage in the tissue can be observed¹⁵.

This study showed decreasing average of gingival epithelial thickness from 1st to 5th day which apparently caused by mitosis disruption in basal lamina. Based on the previous study, the turnover of rat gingival epithelial is 6-8 days⁷. Cells of basal layer take 4-6 days to move towards spinous and granular layer. In next 2 days, the cells move to cornified layer⁷. Decrease on gingival epithelial thickness average between 1st to 3rd day was smaller than 3rd to 5th day. This possibly because at 3rd day, epithelial cells were still in basal layer, while at 5th day epithelial cells were in spinous layer. In this layer, epithelial cells is observed in its largest size. Exposure of VLC disrupts cells mitosis and leads to decrease on cell production. Reducing number of cells causes decrease on epithelial thickness. When cells enter movement phase towards spinous layer, decreasing on epithelial thickness become obviously observed.

This study showed on the 1st day after VLC exposure, average of neutrophil number was highly increased. The results is in accordance with Kumar *et al.* (1997) that neutrophil domination is observed at 6-24 hours after exposure¹⁶. However, the average of neutrophil number was then subsequently decreasing until 5th day. This finding is in agreement with Junquiera and Carneiro (2007) that the neutrophil has a lifespan of over one to four days in the connective tissue and finally died after its mission⁹. In this study, most of neutrophil are presumably completed the mission and died at 5th day. Consequently, average number of neutrophil at 5th day returned as before VLC exposure.

Small increase on average of macrophage number before and 1 day after VLC exposure observed in this study is presumably because not

all the monocytes from the blood vessel are mature and differentiating into macrophage in tissue. Study on literature reveals that monocytes are circulating in the blood 1-2 days and less motile^{15,17}.

In this study, average of macrophage number was notably increased between 1st to 3rd day. It was probably due to maturation and differentiation of all monocytes into macrophage to perform phagocytosis. Phagocytosis is the main function of macrophage, including process of particles adherence on the surface of phagocytes, digestion, and eradication of damaged cells¹⁶.

The highest average of macrophage number in this study was found at 3rd day. This is in accordance with Harrison and Jurosky (1991). They suggested that macrophage is predominantly found in inflammatory tissue on the 3rd day⁸.

However, decreasing average of macrophage number was observed between 3rd to 5th day in this study and indicated completion phase and beginning phase of proliferative inflammation. Kumar *et al.* (1997) suggested the proliferation phase occurs after 2-5 days after injury¹⁶. After phagocytosis, most of macrophage dead then gradually experience autolysis and absorbed into the surrounding tissue⁶. Midwood *et al.* (2004) stated that reduction on macrophage number possibly happens in order to avoid inflammatory processes that are too long, which in the other hand can induce tissue damage¹⁸.

The results of this study indicated that VLC exposure may decrease the thickness of gingival epithelial and increase the number of neutrophil and macrophage of gingival connective tissue of *Sprague dawley* rats.

References

1. Alatas Z. 2007. Efek Kesehatan Pajanan Radiasi Dosis Rendah. *Cermin Dunia Kedokteran* 154: 17-23.
2. Alatas Z, Lusiyaniti Y. 2003. Efek Kesehatan Radiasi Non Pengion pada Manusia. *Cermin Dunia Kedokteran* 138 : 34-40.
3. Anusavice KJ. 1996. *Ilmu Bahan Kedokteran Gigi*. 10th Ed. EGC, Jakarta: 232-236.

4. Fedi PF, Vernino AR, Gray JL. 2004. *Silabus Periodonti*. 4th Ed. EGC. Jakarta.
5. Newman MG, Takei HH, Klokkevold PR, Carranza FA. 2006. *Carranza's Clinical Periodontology*, 10th Ed. Saunders Co., Los Angeles.
6. Guyton A.C and Hall J.E. 1997. Buku Ajar Fisiologi Kedokteran. 9th Ed. EGC, Jakarta: 543.
7. Hamilton IA and Blackwood HJJ. 1974. Cell Renewal of Oral Mucosal Epithelial of Rat. *Journal Anatomi* 117 (2): 313-27.
8. Harrison JW and Jurosky KA. 1991. Healing of Surgical Wounds in Oral Mucoperiosteal Tissues. *Journal of Endodontics* 17 (9): 425-435.
9. Junquiera L.C and Carneiro J. 2007. Histologi Dasar: Text and Atlas. 10th Ed. EGC. Jakarta: 236-8.
10. Wataha JC, Lockwood PE, Lewis JB, Rueggeber FA, Messer RLW. 2004 . Biological effects of blue light from dental cure units, *Dental Materials Journal* 20: 150-7.
11. Willey JM, Sherwod LM, and Woollverton CJ. 2009. *Prescott's Principles of Microbiology*. Mc Graw Hill Higher Education, Boston: 174.
12. Voskanyan KS. 2009. UV and Visible Light-Induced Mutation in Eschericia Coli, <http://www.photobiology.com/photobiology99/contrib/karin/index.html> . Downloaded July 7th, 2010.
13. Wiseman H and Halliwell B. 1996. Damage to DNA by Reactive Oxygen and Nitrogen Species: Role in Inflammatory Disease and Progression to Cancer, *Biochem Journal* 313: 17-29.
14. Underwood JCE. 2000. *General and Systematic Pathology*. 3rd Ed. Churchill Livingstone. London: 202-14.
15. Lawler W, Ahmed A, Hume WJ. 1992. *Buku Pintar Patologi untuk Kedokteran Gigi*. EGC. Jakarta: 9-11.
16. Kumar V, Ramzi SC, Stanley LR. 1997. Basic Pathology. W.B. Saunders Company. Philadelphia: 26-40.
17. Roeslan BO. 2002. *Imunologi Oral*. Faculty of Dentistry-Universitas Indonesia. Jakarta: 5, 38.
18. Midwood KS, Williams LV, and Schwarzbauer JE. 2004. Tissue Repair and te Dynamics of the Extracellular Matrix. *The International Journal of Biochemistry and Cell Biology* 36 (6): 1031-7.