

## SOMATIC EMBRYOGENESIS FROM MERISTEM EXPLANTS OF GINGER

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### ABSTRACT

The use of planting materials from *in vitro* culture, especially derived from somatic embryos has some advantages such as genetically stable and pathogen-free. Meristem culture of ginger through somatic embryogenesis could be a potential method for producing pathogen-free planting materials. Somatic embryogenesis on ginger was performed to obtain vigorous plantlets having the same rhizome size as the mother plant. Callus was induced from meristem tissue of inner bud of Indonesian ginger rhizome Var. Cimanggu-1 and consecutively subcultured into certain media at each steps of experiments. The vigorous embryogenic calli were observed on MS medium containing 100 mg<sup>l</sup><sup>-1</sup> glutamine and 2% sucrose with addition of 1.0 mg<sup>l</sup><sup>-1</sup> 2,4-D + 3.0 mg<sup>l</sup><sup>-1</sup> BA. The highest number of somatic embryos (about 82.0.g<sup>-1</sup> friable calli) was achieved on that medium, 4 weeks after culturing. Furthermore, the optimum growth of embryogenic calli containing somatic embryo was obtained on MS medium enriched with 6% sucrose. The highest number of mature somatic embryos (57.2 embryos) was achieved on MS medium, 18 days after incubation. The regeneration potency of somatic embryos obtained from ginger meristem was 51.20%.g<sup>-1</sup> friable callus. The valuable result of this study was the achievement of normal rhizome size of regenerated plantlets, instead of micro rhizome.

**Key words:** *Zingiber officinale* Rosc., meristem culture, somatic embryogenesis.

### INTRODUCTION

Ginger (*Zingiber officinale* Rosc.) is one of the most important export commodities of Indonesia. This crop is also one of the important components of Indonesian herbal medicine, locally called *jamu*, phytopharmaca and is contributing to the Indonesian foreign exchange and work opportunity for labors. The lack of market stock and serious damages due to pest and disease, have caused high fluctuations to Indonesian ginger exports and prices within the last decades. To overcome the limiting factors, the use of healthy planting material is necessary to be developed.

Plant tissue culture has been adopted for the purpose of *in vitro* mass propagation on various crop species. In case of the Big-White Ginger Cultivar of Indonesian Variety Cimanggu-1, the use of shoot bud-derived plantlets, either by direct organogenesis or through callusing, has been developed to obtain disease-free planting materials (Mariska

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and Syahid 1994). In further field experiments, the healthy plantlets and vigorous plants were not able to produce normal rhizomes (the same-big rhizome as their mother plants), even though they were repeatedly planted in three consecutive generations (Syahid and Hobir 1996). It is suggested that genetic alteration or epigenetic change during the *in vitro* culture and regeneration have been performed. In order to eliminate the genetic change during the *in vitro* culture, other regeneration pathway should be considered.

Somatic embryogenesis has been accomplished in some species for producing millions of seeds. The form of somatic seeds through somatic embryogenesis, is more efficient than organogenesis. Besides, somatic embryogenesis is more preferable in plant genetic improvement through *in vitro* culture and genetic transformation as well, because single cell derived-plant is easier to be controlled as somatic embryo derived-plant. In general, somatic embryogenesis and meristem derived plants are true-to-type and genetically identical to the mother plants (Evans and Sharp 1986; Jimenez 2001), however, certain differences might appear according to the plant species characteristics. The true-to-type of somatic embryo derived plantlet has been found in *in vitro* culture of *Picea abies* (Heinze and Schmidt 1995). On the other hand, among recalcitrant species such as serealia crops and conifers, good results have been performed by the induction of somatic embryogenesis.

Achievements in inducing somatic embryogenesis in monocots are mostly derived from generative explants. Meanwhile, in dicotyl vegetative explant, such as leaves, are commonly applied for inducing somatic embryo derived-plantlet. Leaf explant of orchard grass completely induces somatic embryo through the formation of embryogenic calli with addition of a strong auxin like dicamba (Bhojwani and Razdan 1996). On the other hand, Kackar *et al.* (1993) performed the somatic embryo culture of Indian ginger Var. Eruttupetta by using an aseptic leaf-explant with the addition of 2,4-D and dicamba. The same explant source was also performed in inducing somatic embryogenesis of fingerroot (*Boesenbergia rotunda* L.) and galangal (*Kaempferia galanga* L.) (Tan *et al.* 2005; Rahman *et al.* 2004).

This study was aimed at obtaining the normal-size of rhizome (the same size as their mother plant) through induction of somatic embryos from meristem explant of Indonesian Var. Cimanggu-1. Most meristematic cells of meristem tissue derived-somatic embryo are genetically stable and not easy to be mutated (Bach and Pawlowska 2003). Therefore, to eliminate somaclonal variation and other genetic alteration during the *in vitro* culture, meristem tissue will be applied for the induction of somatic embryogenesis on ginger.

## MATERIALS AND METHODS

### Explant preparation

The inner shoot bud (meristem) of Indonesian Var. Cimanggu-1, was excised and simultaneously sterilised by using sterilizing agents such as 70% EtOH, 5% sodium hypochlorite and 0.2% mercury chloride, for about 5-10 minutes, followed by rinsing with sterile-aquadest.

### Callus induction

Sterilized meristems were placed on MS basal medium (Murashige and Skoog 1962) consisting of 8% agar, 2% sucrose, 100 mg l<sup>-1</sup> L-glutamine, 1.0 to 3.0 mg l<sup>-1</sup> 2,4-dichlorophenoxy acetic acid (2,4-D) and 0 to 5.0 mg l<sup>-1</sup> N<sup>6</sup>-benzyl adenin (BA). A single factor experiments were arranged in completely randomized design, replicated three times.

### Subculturing.

Embryogenic calli were formed 8 weeks after culturing on callus induction medium, and then transferred on to hormone-free MS or N<sub>6</sub> basal media with the addition of 3% mannitol for calli proliferation. A single factor experiments were arranged in completely randomized design, replicated four times for each treatments. Pro-embryos were then subcultured into the best basal medium, either MS or N<sub>6</sub> basal media, according to the best result of the previous stage of experiment, with the addition of 6% sucrose (6S) for obtaining mature embryo. Single-factor experiments were arranged in completely randomized design, replicated four times.

### Regeneration

Mature embryos (torpedo-like structure-embryos) were subcultured into regeneration medium for further development. MS basal medium with addition of 3% sucrose and 200 mg l<sup>-1</sup> L-proline were applied in combination with PGRs such as BA at the concentration of 0, 0.1, 0.5 and 1.0 mg l<sup>-1</sup>, GA<sub>3</sub> at the concentration of 0, 0.1, 0.3, 0.5 and 1.0 mg l<sup>-1</sup>. Single-factor experiments were arranged in completely randomized design, replicate three times.

### Histology

The callus at each stage of development was subjected to histological analysis according to Sass (1951) by using paraffin-embedded callus dissection following Formaldehyde-glacial Acetic acid-Alcohol (FAA) fixation series.

### Statistical analysis

All collected data were analyzed by using ANOVA (P<sub>0.05</sub>) followed by Duncan's Multiple Range Test (P. 0.05) (Windows Computers, 2000). Observation results on embryo proliferation were analyzed by using t-test according to Furlong *et al.* (2000).

## RESULTS AND DISCUSSION

### Callus induction

Callus initiation was on tract when the basal-edge of the meristems enlarged, followed by the change of shoot-dome colour from white to yellowish, at two weeks after culturing. Eight weeks after culturing, friable callus (embryogenic callus) formation of about 93.33%/explant was formed on MS basal medium enriched with 1 mg l<sup>-1</sup> 2,4-D in combination with 3 mg l<sup>-1</sup> BA. Though, that combination was statistically the same as the treatment without BA (Table 1).

Table 1. Percentage of embryogenic calli from ginger meristem cultured on MS medium enriched with various concentration of 2,4-D and BA, 8 weeks after culturing

Treatment		Percentage of embryogenic calli (%)
2,4-D (mg.L <sup>-1</sup> )	BA (mg.L <sup>-1</sup> )	
1	0	86.67 a*)
	1.0	8.33 e
	3.0	93.33 a
	5.0	70.00 b
2	0	46.67 c
	1.0	25.00 d
	3.0	63.33 b
	5.0	61.67 b
3	0	28.33 d
	1.0	36.67 cd
	3.0	63.33 b
	5.0	61.67 b

Note:

Numbers followed by the same letters are not significantly different according to DMRT (5%).

The application of 2,4-D on ginger meristem culture showed that the higher the concentration applied, the more compact calli were observed. Furthermore, when 2,4-D was applied at 2-3 mg.l<sup>-1</sup> browning and necrotic calli were observed. An addition of 1.0 mg.l<sup>-1</sup> BA into medium containing 2,4-D neither vigorous embryogenic calli were obtained, low quantity and compact calli were observed (Table 1).

### Embryogenesis from the subcultured callus

For obtaining globular somatic embryos structure the formed embryogenic calli were cultured on either MS or N<sub>6</sub> basal media with the addition of 3% mannitol. Transparent globular embryos were developed on both media in one week after subculturing. The shape of ginger embryos at globular phase was generally round or oval (Fig. 1a).

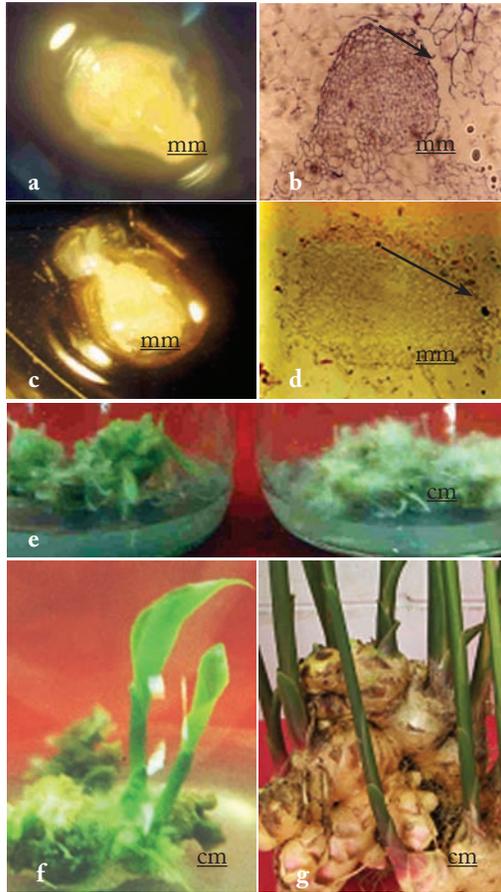


Figure 1. The development of ginger meristem into newly regenerated plant with normal rhizome size through somatic embryogenesis.

- The structure of globular somatic embryo of ginger, 4 weeks after subculturing into proliferation medium (30 times enlargement).
- Globular embryo, 2 weeks after proliferation (40 times enlargement). Protoderm layer started to differentiate (arrowed).
- The structure of torpedo somatic embryo of ginger, 18 days after subculturing into maturation medium (10 times enlargement).
- Torpedo embryo, 18 days after subculturing into maturation medium (40 times enlargement). Arrowed: differentiated procambium.
- Embryo somatic-derived seedlings on MS medium supplemented with  $1 \text{ mg l}^{-1}$  BA (Left) and embryo somatic-forming adventitious roots on hormone free-MS medium (Right), 30 days after subcultured (1 : 1.4 scaled).
- Embryo somatic-derived normal plantlet, 8 weeks after subcultured on to hormone-free MS medium (1: 1.3 scaled).
- Normal rhizome size of ginger-regenerated plants obtained from meristem culture through somatic embryogenesis.

Numbers of globular phase of somatic embryos increased due to the age of cultures. The highest number of somatic embryo was obtained at 4 weeks after subculturing,  $82.00 \pm 12.25$  embryo.g<sup>-1</sup> embryogenic callus on MS medium and  $70.00 \pm 19.26$  embryo.g<sup>-1</sup> embryogenic callus on N<sub>6</sub> medium. Statistical analysis showed that MS basal medium was more capable in inducing higher numbers of somatic embryo than that of N<sub>6</sub> basal medium (P<sub>0.05</sub>). The number of somatic embryo decreased by the increase of age of cultures (Table 2). Histological analysis conformed that the somatic embryo was induced from cortex tissue and that the globular phase developed at 2 weeks after culturing on proliferation medium (Fig. 1b).

Table 2. Number of somatic embryos derived from ginger meristem culture on MS and N<sub>6</sub> basal media enriched with 3% mannitol

Period (weeks)	Average number of somatic embryos/g of embryogenic callus	
	MS	N <sub>6</sub>
1	32.75 ± 11.97	39.75 ± 5.31
2	54.50 ± 20.16	51.50 ± 9.66
3	70.50 ± 31.97	48.75 ± 15.16
4	82.00 ± 12.25 <sup>1)</sup>	70.00 ± 19.26 <sup>1)</sup>
5	53.75 ± 16.32	36.25 ± 9.28
6	51.50 ± 10.92	22.75 ± 15.28
7	43.50 ± 19.55	11.00 ± 7.65

Note: Based on t-test at 5% level.

Mature embryo formation was performed on MS basal medium, enriched with 6% sucrose. Cylindrical form of mature embryo started to develop into 2 different domes, *i.e.* cotyledons and apical shoot-forming dome as well as root-forming dome. Two days after subculturing into MS medium enriched with 6% sucrose, about  $10.60 \pm 4.76$  torpedo-shape somatic embryos were initiated from 1 g of embryogenic calli (Table 3).

Table 3. Number of mature somatic embryos on MS basal medium supplemented with 6% sucrose

Age(day)	Average number of mature embryo/g of embriogenic calli <sup>1)</sup>
2	10.60 ± 4.76
6	33.40 ± 6.74
10	55.80 ± 12.73
14	38.60 ± 6.53
18	57.20 ± 15.99
22	45.80 ± 10.50
26	28.00 ± 8.65
30	16.20 ± 8.91

The highest number of torpedo-shape somatic embryo ( $57.20 \pm 15.99$  embryos) was obtained at 18 days after subculturing. Mature somatic embryo was capable to form two different domes, *i.e.* shoot and root (Fig. 1c-d). However, in some cases apical meristem structure was more dominant than shoot meristem.

Histological section showed that during the formation of torpedo-shape somatic embryo, root-apical, procambium and scutellum differentiations were performed (Fig. 1c-d).

### Regeneration

Germination of somatic embryo was characterized by the formation of the triangle-shape at meristem apical dome with yellow greenish in color, at 6 days after subcultured. At the second week of culturing, germinating seeds started to appear and developed to become normal seedlings/plantlets, especially when BA was added into MS basal medium.

Three weeks after culturing, various shapes of somatic embryos either a single normal or fused embryos, had been differentiated into plantlets. Meanwhile, the abnormal embryos underwent necrosis and then aborted. Embryo germination was indicated by the presence of shoot and root buds. Four weeks after culturing, the morphogenesis of somatic embryos were clearly visible, nevertheless the low ability of somatic embryos to germinate was still a limiting factor in meristem culture of ginger.

MS basal medium supplemented with  $1 \text{ mg l}^{-1}$  BA was found to be the best culture medium for obtaining normal plantlets at the age of 30 days, to which the highest number of seedlings was obtained (22.1 seedlings). On the other hand, instead of growing plantlet, vigorous adventitious roots were obtained on hormone-free MS medium (Fig. 1e). Furthermore, it was found that both BA and  $\text{GA}_3$  significantly affected the somatic embryo regeneration ( $P_{0.05}$ ). However, the higher the concentration of  $\text{GA}_3$ , the lower the number of seedlings was obtained (Table 4).

Table 4. Number of seedlings derived from somatic embryo of ginger meristem cultured on MS medium supplemented with BA and GA<sub>3</sub>

Treatment		Average number of seedlings
BA (mg l <sup>-1</sup> )	GA <sub>3</sub> (mg l <sup>-1</sup> )	
0	0	7.3 cd*
	0.1	16.4 ab
	0.3	16.4 ab
	0.5	9.3 bcd
	1.0	9.3 bcd
0.1	0	7.3 cd
	0.1	10.0 bcd
	0.3	3.7 de
	0.5	16.1 ab
	1.0	11.5 bc
0.5	0	0.5 f
	0.1	0.5 f
	0.3	0.5 f
	0.5	12.8 abc
	1.0	17.4 ab
1	0	22.1 a
	0.1	12.6 abc
	0.3	12.5 abc
	0.5	0.5 f
	1.0	1.2 ef

Notes : Numbers followed by the same letter are not significantly different according to DMRT (5%). For statistical analysis data were transformed into  $\sqrt{(y + 0.5)}$ .

Based on this study, it was found that regeneration potency of meristem culture of ginger through somatic embryogenesis was 51.20%.g<sup>-1</sup> of embryogenic calli when cultured on MS medium supplemented with 1 mg l<sup>-1</sup> BA. The addition of GA<sub>3</sub> into MS basal medium produced only 34.13% of regeneration potency, and 1.13 to 31.84% of regeneration potency when BA was combined with GA<sub>3</sub>. Therefore, to gain an optimal germination of ginger somatic embryo, higher concentration of BA ( $\geq 1$  mg.l<sup>-1</sup>) without addition of GA<sub>3</sub>, was necessary.

Germinating somatic embryos from MS medium supplemented with either 1 mg l<sup>-1</sup> BA, 0.5 mg l<sup>-1</sup> BA, 0.1 mg l<sup>-1</sup> BA + 0.3 mg l<sup>-1</sup> GA<sub>3</sub> or 0.1 mg l<sup>-1</sup> GA<sub>3</sub> were then transferred onto hormone-free MS medium with the addition of 3% sucrose. Among them, the most vigorous plantlets were obtained from MS + 0.1 mg l<sup>-1</sup> BA + 0.3 mg l<sup>-1</sup> GA<sub>3</sub> culture medium-derived seedlings (Fig. 1f), at 8 weeks after subculturing. However, the number of embryo somatic-generated plantlets remained low, of which only about 2-6 plantlets were observed.

Abundant embryogenic calli (93.33%/explant) from meristem culture of ginger were achieved on MS basal medium supplemented with 1 mg l<sup>-1</sup> 2,4-D in combination with 3 mg l<sup>-1</sup> BA, and 100 mg l<sup>-1</sup> L-glutamine, at 8 weeks after incubation. This result showed that 2,4-D was necessary for initiation of embryogenic callus of ginger, with the addition of BA at proper concentration. The same result was also found in embryogenic callus initiation of *Kaempferia galanga* L. (Vincent *et al.* 1992).

Callus proliferations were obtained on hormone-free MS medium consisting of 3% mannitol. A globular-shape pro-embryo started to develop at the first week after subculturing into proliferation medium. Quantities of globular embryo increased due to the age of culturing up to seven weeks, then gradually decreased.

Ammonium-rich basal media such as MS, which consists of high concentration of NH<sub>4</sub>NO<sub>3</sub> and myo-inositol, supported well the differentiation of ginger somatic embryo, though an adverse effect was observed on other Grammineae species (Talwar and Rashid 1989; Adkins *et al.* 2002). Salt nutrients in MS medium with addition of 3 to 12% sucrose are usually sufficient to support embryo differentiation (Raghavan 2003; Anbazhagan and Ganapathi 1999). An addition of 3% mannitol into MS basal medium, assumed that it acted as an osmotic regulator which in turn affected the rate of cell differentiation and stimulated embryogenic cells morphogenesis in meristem culture of ginger. Torres *et al.* (2001) reported that the development of synchronized embryo and the increase of its quantity were observed when mannitol was added into MS medium. The same results were also found on embryogenic culture of papaya, celery, alfalfa and maize (Ziv 1999; Bronsema *et al.* 1997).

Somatic embryo maturation in meristem culture of ginger was indicated by the change of embryo color and formation of the brown spots. These brown spots suggest the presence of an active substance *i.e.* amylum, protein and lipid, secreted by somatic embryo from the osmotic cell pressure. The increase of secretion of active substances suggests its correlation with the desiccation process when somatic embryo enters the germination phase. Bach and Pawlowska (2003) suggested that the presence of amylum, protein and lipid around the vascular cells was an indicator in somatic embryo development. Similarly, Oropeza *et al.* (2001) reported that the development of embryogenic calli was associated with the number and type of intracellular protein and callus cells. Hence, somatic embryo maturation from proliferated calli of ginger meristem is interrelated with the type and number of active substance among the embryogenic cells.

The growth and development of ginger mature somatic embryo into complete plantlets (seedlings) sometimes go along with the abnormal germinated seeds, such unexpected growth of hairy roots, hereafter we named as an early germinating seeds. According to Bronsema *et al.* (1997), somatic embryo maturation is associated with the presence of scutellum, a coleoptile and root-bud like structures. Somatic embryo maturation was also affected by the composition of applied culture medium. The presence of ammonium ion and nitrate at a high concentration is a prerequisite in ginger somatic embryo development. High concentration of ammonium ion supported the process of

somatic embryo differentiation into normal plantlets (George 1993). These results were in agreement with the findings of Krikorian (1995), Adkins *et al.* (2002) and Ramage and Williams (2002). However, the needs of ammonium ion for the growth of somatic embryo and its morphogenesis depend on the explant sources and the initial plant growth regulators applied.

According to Percy *et al.* (2000), high osmotic pressure in culture medium would increase the somatic embryo formation. Sucrose, usually applied as a carbon source, is also a potent osmotic agent (van Creij *et al.* 1999). Another potential osmotic agent is Absciscic Acid (ABA), that at a proper concentration it would enable to control the medium osmolarity (Raghavan 2003; Mohan and Krishnamurthy 2002). To overcome the early seeds germination (the formation of unexpected hairy roots) in somatic embryogenesis of ginger, an experiment on addition of ABA, aside from sucrose, into culture medium of somatic embryo maturation is now in progress.

Somatic embryo germination was indicated by the simultaneous formation of shoots and roots. Goh *et al.* (1999) stated that embryo germinated when the plumulae started to emerge. In this research, adventitious roots formation was observed at first stage of regeneration and increased by the increase of age of culture up to the fourth week.

Addition of plant growth regulators into regeneration medium would accelerate the germination process and increase the quantity and quality of seedlings. In this research, a high percentage of somatic embryo germination was observed on the medium containing  $1 \text{ mg.l}^{-1}$  BA, at 30 days after culturing. Regeneration potency of ginger meristem somatic embryo reached  $51.20\%.\text{g}^{-1}$  of embryogenic calli cultured on MS basal medium enriched with  $1 \text{ mg.l}^{-1}$  BA, of which the number of seedlings was 8.98 times higher than that of the control medium. Meanwhile,  $\text{GA}_3$  alone or in combination with BA, remained ineffective in supporting somatic embryo germination of ginger. However, to obtain the optimum growth of plantlet, the presence of BA is a prerequisite in meristem culture derived somatic embryo of ginger. Similar result was also found in the development of ginger leaf derived embryogenic callus (Kackar *et al.* 1993), though simultaneous development of somatic embryo derived either root or shoot meristems remained unclear.

The development of somatic embryo into normal plantlet commonly occurs through four stages *i.e.* callus induction, callus proliferation, embryo maturation and germination. In case of somatic embryo derived from ginger meristem culture, its development into normal plantlet needs a transfer process of germinated seeds from the regeneration medium into new medium. Such new medium is called growth medium for plantlet development. In this experiment the medium used for the growth of plantlet was hormone-free MS medium with the addition of 3% sucrose.

Although the development of protocol in ginger somatic embryogenesis required more researches, the results of this experiment, somehow, have a definite advantage in *in vitro* studies of ginger, especially in generating healthy planting material with normal rhizome size, a bigger size than the usual micro rhizomes resulted from shoot

tip-derived *in vitro* plantlets (Fig. 1 g). Therefore, the encouraging results of this study gave great possibility for development of ginger resistant variety either through *in vitro* selection, somatic hybridization or genetic transformation.

## CONCLUSIONS

The addition of 1 mg.l<sup>-1</sup> 2,4-D and 3 mg.l<sup>-1</sup> BA into MS basal medium enriched with 100 mg.l<sup>-1</sup> of glutamine and 2% sucrose, was effective in inducing embryogenic calli from meristem explants of Indonesian ginger. In further embryo development, MS medium was found to be more capable to increase mature somatic embryo than N<sub>6</sub> medium.

The regeneration potency of somatic embryos obtained from Indonesian ginger meristem was 51.20%/g friable callus. Though, further researches are needed to be conducted, especially to eliminate early germinating seeds. The protocol for generating somatic embryo derived plantlet from meristem culture of ginger was developed. The most valuable result of this study was the achievement of normal rhizome size of regenerated plantlet, instead of micro rhizome.

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