

## EXOGENOUS APPLICATION OF TRYPTOPHAN AND INDOLE ACETIC ACID (IAA) TO INDUCE ROOT NODULE FORMATION AND INCREASE SOYBEAN YIELD IN ACID, NEUTRAL AND ALKALINE SOIL

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### ABSTRACT

The research aimed to study whether soil pH affects exogenous application of amino acid tryptophan and IAA in increasing root nodules and soybean yield. The experiment was conducted in greenhouse using 20 cm diameter plastic pot filled with fine soil, arranged in completely randomized design (CRD) with four treatments. Each treatment combination was repeated three times. Three seeds of local soybean variety were planted into each pot then to remain one plant a week after planting. Variables observed were root nodules number, shoot dry weight and seed yield. Plant nutrients were supplied in the form of solution to meet crop needs. The data obtained were analyzed statistically by F test at 5% of level confidence, followed by Duncan's multiple range test when there was a significant effect. The results showed that both exogenous amino acid tryptophan and IAA increased number of root nodules, shoot and root dry weight and soybean yield. Higher root nodules number was taken from the treatment combination of 0.001 ppm IAA applied at V<sub>3</sub> in Alfisol and Vertisols, and 1.0 ppm on Entisols. However, the highest soybean yields were taken from the treatment combination of 1.0 ppm tryptophan applied at V<sub>0</sub> in Alfisols (6.51 g plant<sup>-1</sup>).

Keywords: alfisol, entisol, extra cellular, phyto-hormones, vertisol

### INTRODUCTION

Biological nitrogen fixation is the second most important biological processes after photosynthesis. Even in natural ecosystems where there is no factory-made fertilizers and

animal waste, plant and microbial N needs are fully and dependent on the conversion of dinitrogen (N<sub>2</sub>) air by various types of prokaryotic microbes. For soybean, the existence and effectiveness of root nodules is critical (Zuberer, 1999; Elmerich and Newton., 2007; Graham, 2008; Howieson *et al.*, 2008). Effective root nodules capable of meeting the needs of up to 75% nitrogen (Jutono, 1985; Anonymous, 1986; Ohyama *et al.*, 2009).

According to Jutono (1985) the existence of effective root nodules should be considered in an effort to increase soybean production and reducing the use of fertilizer N. Ohyama *et al.* (2009) said that one of the reasons for the low yield of soybean root nodules compared to its potency is less effective but it is very difficult to create the optimum conditions for biological N<sub>2</sub> fixation. For instance, the formation of root nodules and N<sub>2</sub> is very sensitive to the effects of a poor environment such as drought, poor aeration, nutrient deficiencies or nutrient imbalance, soil pH (Richardson *et al.*, 1988), as well as compatible rhizobia populations and the competitiveness to native ineffective soil rhizobia. Root nodule formation is influenced by internal factors both from plants and bacteria, such as the ability of crops to excrete tryptophan (Gray and William, 1971) and the ability of bacteria to oxidize it to IAA. In the normal process, rhizobium oxidized the amino acid tryptophan excreted by soybean plant to IAA.

But, due to certain environmental factors, such as low or high soil pH, plants may not be able to produce the amino acid tryptophan so the IAA can not be synthesized, and root nodule was not formed. This problem may be overcome if both IAA or tryptophan could be given exogenously. According to Mishra *et al.* (1999) the use of extragenous fitohormon shows positive

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response to the formation of root nodule. The indole acetic acid (IAA) is a natural auxin and one of the main growth regulators in higher plants (Prieto C *et al.*, 2011). Plants synthesizing higher IAA showed a significant increase in shoot fresh weight and upregulation of nitrogenase gene (Bianco *et al.*, 2014).

In the previous research, Sudadi (2011) showed that the use of extra cellular (exogenous) amino acid tryptophan and IAA increased the number of root nodules and yield of soybeans grown in sterile sand media (neutral pH). Srinivasan and Gopalkrishna cit. Mishra *et al.* (1999) stated that the addition of extra-cellular auxin or Naphthilic acetic acid (NAA) encourage the formation and the growth of *Medicago sativa* root nodules and *Arachis hypogaea* (peanut) and 1-naphthoxy and acetic acid (p-chlorophenoxy)-isobutyric acid improve root nodulation of nuts (Cartwright cit. Mishra *et al.*, 1999). Kamal (2000) stated that epibrasinoloid added on soybean growth media increased the number of root nodules per plant. Formation of root nodules can also be induced by other extra cellular fitohormon as cytokines (Nandwal and Bharti cit. Mishra *et al.*, 1999) and Giberilic Acid (GA) (Bishnoi and Krishnamoorty cit. Mishra *et al.* 1999). This research aimed to investigate if soil pH affects the the ability of extra cellular amino acid tryptophan and IAA to enhance root nodules formation and increase soybean yield.

## MATERIALS AND METHODS

The experiments were conducted from March - October 2012 in greenhouse, used 2-gallon plastic pots with Alfisol, Entisol or Vertisol soil planting medium. Soybean of local variety was used and the Rhizobium inoculant was taken from The Laboratory of Soil Microbiology and Environmental, Department of Microbiology, Faculty of Agriculture, Gadjah Mada University, Yogyakarta. The experiment was arranged in completely randomized design (CRD) factorial with four treatment factors, namely the soils ordo ( $T_1$  = Alfisol,  $T_2$  = Entisol and  $T_3$  = Vertisol), extra-cellular phytohormones ( $Z_1$  = amino acid tryptophan and  $Z_2$  = IAA), time of phytohormone application ( $V_0$  = at planting day and  $V_3$  = vegetative phase 3) and phytohormone concentrations ( $K_0$  = 0 ppm ;  $K_1$  = 0.001 ppm;  $K_2$  = 0.1 ppm and  $K_3$  = 1.0 ppm). Each treatment

combination was repeated three times. The variables observed were the number of root nodules, shoot dry weight and plant yield. Sampling for observation of root nodules and plant dry weight was performed at  $R_1$  growth stage (maximum vegetative phase) and at  $R_8$  growth stage (fully matured pods) for plant yield (Hidayat, 1985, Kamal, 2000). Fertilizer N was applied at a dose of 20  $\text{kg ha}^{-1}$  N before planting as a starter (Afza *et al.* 1987), while P and K fertilizer were given prior to planting at a dose equivalent to 75  $\text{kg ha}^{-1}$   $\text{P}_2\text{O}_5$  and 100  $\text{kg ha}^{-1}$   $\text{K}_2\text{O}$  (Pasaribu and Suprpto, 1985). Calcium (Ca) and magnesium (Mg) were given in the form of dolomite with the dose of 200  $\text{kg ha}^{-1}$  (Jutono, 1985). Micro nutrients were added in the following doses (Ismunadji and Mahmud, 1985): 10  $\text{kg ha}^{-1}$  Fe, 10  $\text{kg ha}^{-1}$  Mn, 4  $\text{kg ha}^{-1}$  Zn, 10  $\text{kg ha}^{-1}$  Cu, 200  $\text{g ha}^{-1}$  Mo, and 1.2  $\text{kg ha}^{-1}$  B. The data obtained were analyzed statistically with F test at the level confidence of 5%, followed by Duncan's multiple range test if significant.

## RESULTS AND DISCUSSION

In addition to the internal factors, soil properties is one of external factors influencing soybean growth and yield as well as root nodulation. Soil nutrient availability (Pasaribu and Suprpto S., 1985; Ismunadji and Mahmud, 1985; Ohyama *et al.*, 2009), aluminium /manganese toxicity and soil pH (Widjaja-Adhi, 1985; Richardson, 1988) are some of soil chemical properties which have been known to influence them. These soil chemical properties were also believed to influence the population growth of Rhizobium and its ability to nodulate soybean root (Yutono, 1985; Coyne, 1999). For rhizobia, the most problematic environments are marginal land with low rain fall, extreme temperature, acid soil with low nutrients status, and poor water holding capacity (Bottomly *cit.* Keyser *et al.*, 1993). According to Ohyama *et al.* (2009) there are three factors influencing soybean yield: soybean is sensitive to soil physical, chemical and biological factors stress, including climatic factor. Secondly, soybean yield will decrease significantly due to pests and diseases, and the third, soybean yield was strongly influenced by biological nitrogen fixation of its root nodule although it is very difficult to make soil condition favorable for optimum

biological nitrogen fixation. Some chemical properties and texture of the soil used for the study are presented in the following table.

Table 1. Some chemical properties and texture of the soil used for research

Soil properties	Alfisol	Entisol	Vertisol
pH (H <sub>2</sub> O)	5.60	6.80	7.80
OM content, %	1.89	3.35	1.93
Total-N, %	0.09	0.18	0.12
Available-P, ppm	3.36	42.00	35.02
Exch.-K, cmol(+) kg <sup>-1</sup>	0.27	0.29	0.30
CEC, cmol(+) kg <sup>-1</sup>	25.60	24.20	48.80
Soil texture	clayed	Sandy clay loam	clayed
- sand	33	37	30
- silt	7	31	14
- clay	60	32	56

From the above table it can be said that Alfisol had fertility constraints that most of its chemical properties such as pH, organic matter content, total N, available P and K were low, whereas Alfisol had the highest fertility rate. In addition, the soil texture of Alfisol was probably better than the others.

The analysis of variance (Anova) showed that the influence of soil ordo (soil pH), and kind,

concentration and application time of phytohormones as well as their interaction on root nodules number of soybean at maximum vegetative phase were highly significant ( $P \leq 0.01$ ). The effect of those treatment factor interaction on nodule numbers was presented in Figure 1. The highest root nodules number obtained from treatment combination of K<sub>3</sub> Z<sub>2</sub>V<sub>3</sub>T<sub>2</sub> (1.0 ppm IAA at vegetative phase-3 in Entisol) with 20.67 nodules plant<sup>-1</sup>. According to Gosh and Basu (2002) in culture medium, Rhizobium synthesized IAA from L-tryptophan and the maximum production at 20 hr when the bacteria reached the stationary phase of growth. Nevertheless, this nodule number was neither different significantly with both nodule numbers obtained from treatment combination of K<sub>3</sub>Z<sub>2</sub> V<sub>0</sub> T<sub>2</sub> (1.0 ppm IAA at planting day in Entisol) nor of treatment combination K<sub>2</sub>Z<sub>2</sub> V<sub>0</sub> T<sub>2</sub> (0.1 ppm IAA at planting day in Entisol). It mean that IAA was more efficient when applied at planting day (early planting) than at vegetative-3 phase. In general, the number of root nodules in Vertisols was lower than that of Alfisols and Entisols presumably due to the physical properties of Vertisols which had hard soil structure so that plant roots were not able to penetrate it well.

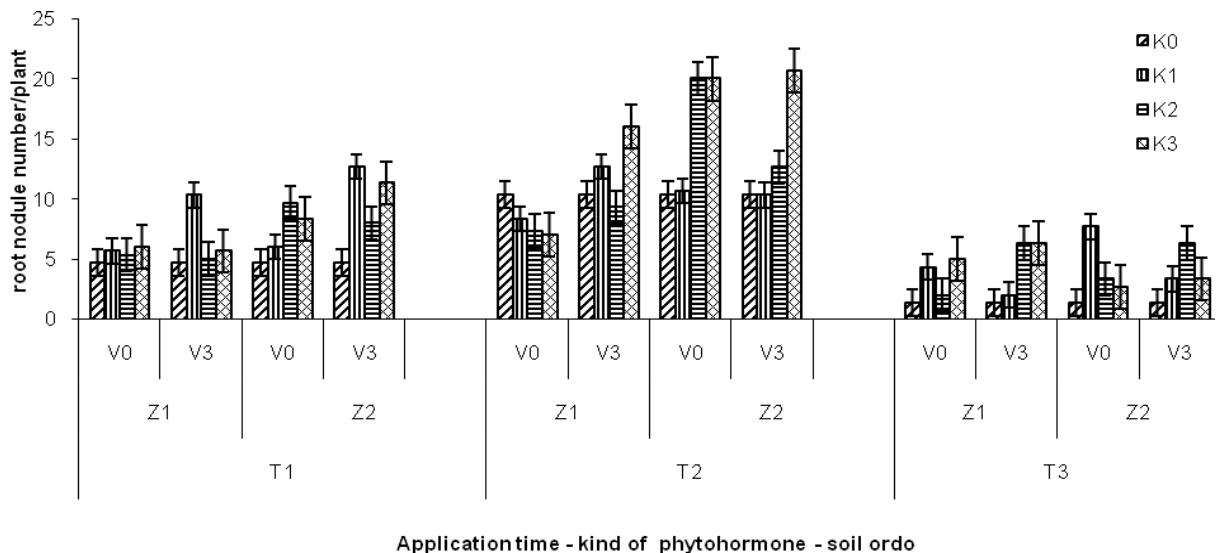


Figure 1. Effect of phytohormones concentration (K), application time (V), kind of phytohormones (Z) and soil ordo (T) on soybean root nodules at maximum vegetative phase. (Note: K<sub>0</sub> = 0 ppm; K<sub>1</sub> = 0.001 ppm; K<sub>2</sub> = 0.1 ppm and K<sub>3</sub> = 1.0 ppm; V<sub>0</sub> = at planting date; V<sub>3</sub> = at vegetative phase-3; Z<sub>1</sub> = amino acid tryptophan; Z<sub>2</sub> = IAA; T<sub>1</sub> = Alfisols; T<sub>2</sub> = Entisols; T<sub>3</sub> = Vertisols)

Phytohormone including auxin (IAA) plays an important role in various physiological processes during plant growth and development (Baca and Elmerich, 2007). At the cellular level, auxin affects the process of division, elongation and differentiation of cells, whereas the level of auxin affects plant tropism, the dominance of shoots, leaf senescence, sprouts morphology, leaf abscission, flowering, fruit ripening and formation and root growth (Nonhebel and Bandurski, 1984; Pandey *et al.*, 1996; Guilfoyle *et al.*, 1998).

In addition to plant height, plant growth can be measured by the increase of biomass dry weight, which can be expressed in terms of the dry weight of shoots and roots. From the analysis of varian, it was known that soil ordos,

as well as kinds, concentration and application time of phytohormones had very significant effect on shoot dry weight, but interaction among them had no significant effect.

According to the shoot dry weight (Figure 2), the growth of soybeans in Vertisol was the best followed by Entisols and Alfisols. This is because Vertisols had the best soil fertility as demonstrated by soil pH, cation exchange capacity and organic matter content. Generally, Vertisols is categorized in fertile soil if there is enough water and organic matter content. The influence of phytohormone kinds on soybean shoot dry weight is significant as shown in Figure 3.

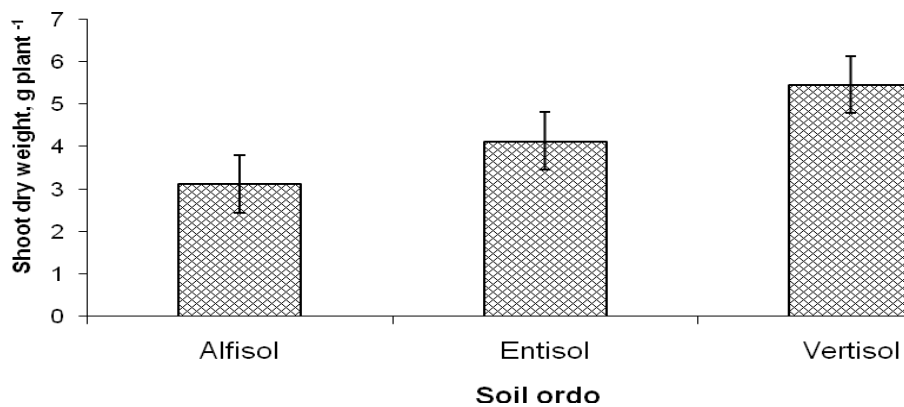


Figure 2. The influence of soil ordos on soybean shoot dry weight

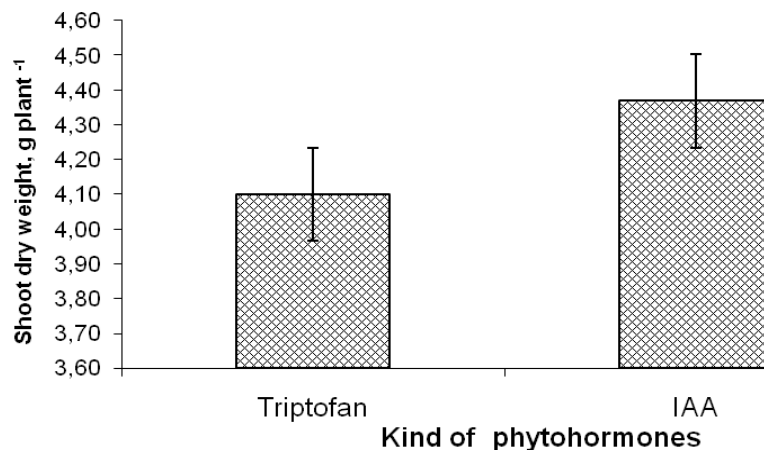


Figure 3. The influence of phytohormone kinds on soybean shoot dry weight

Treatment with IAA produced shoot dry weight higher than the amino acid tryptophan (Figure 3). This is because the IAA is a plant growth regulator capable of affecting the division, elongation and differentiation of cells, and it is ready to use, whereas amino acid tryptophan is needed first to oxidize IAA before it is ready to use. Bianco *et al.* (2014) said that plants synthesizing higher IAA will increase in higher shoot fresh weight, and Phytohormone auxin regulates plant developmental processes (Spaepen and Vanderleyden, 2010).

Existing research suggests that there are opportunities to use extra cellular phytohormones to increase root nodule formation and  $N_2$  fixation of soybean plants. For example, Srinivasan & Gopalkrisna stated that the addition of exogenous auxin or Naphthilic acetic acid (NAA) stimulate the formation and growth of *Medicago sativa* root nodules and *Arachis hypogaea* (peanut), while Cartwright stated that the addition of 1 - naphthoxy acetic acid and (p-Chlorophenoxy)-isobutyric acid improves the process of root nodulation in legumes (Mishra *et al.* 1999). Likewise, Bishnoi and Krishnamoorthy, and Nandwal an& Bharti stated that root nodule formation can also be induced by other phytohormones such as extra cellular cytokinin and GA (Mishra *et al.* 1999).

In the case of application time, phytohormones which was applied at  $V_3$  induced higher plant growth as indicated by shoot dry weight although it was not significantly different when applied at  $V_0$  (Figure 4). It was probable that

the applied amino acid tryptophan and IAA at planting day ( $V_0$ ) would damage more phytohormones than when applied at  $V_3$  so the increase in soybean shoots dry weight was less.

Soybean growth increased concomitantly with the increase of phytohormones concentration applied up to 0.1 ppm, then it decreased with increasing phytohormones concentration as showed Fig. 5. Bianco *et al.* (2014) stated that the knowledge of maximum level of IAA biosynthesis, resulting in the maximal increase of plant growth.

The influence of soils, as well as the kinds, time of application and concentration of the extra cellular amino acid tryptophan and IAA on soybean seed dry weight are presented in Figure 6.

The result showed that there were highly significant ( $P=0.0000$ ) soil ordo, application time of phytohormones, and their concentrations as well as the interaction of the fourth treatments on seed dry weight. The kind of phytohormone showed no significant effect. Highest soybean grain yield was obtained from combination of treatments of 1.0 ppm amino acid tryptophan in the early planting ( $V_0$ ) for Alfisol soil ( $6.51 \text{ g plant}^{-1}$ ) with an increase of 202% compared to no phytohormone ( $3.22 \text{ g plant}^{-1}$ ). Figure 2 above shows that the effect of phytohormone application on soybean seed dry weight (than without phytohormone) in Alfisols was higher than that in both Entisols and Vertisols. Presumably, this is because the Alfisols was less fertile than the others. This condition made phytohormone give more positive impact to nodule formation,  $N_2$  fixation, growth and yield of soybean.

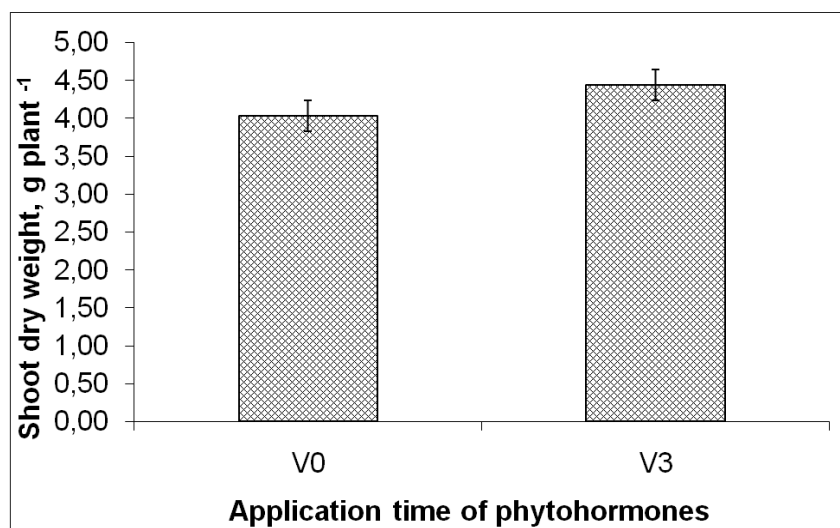


Figure 4. The influence of phytohormones application time on soybean shoot dry weight

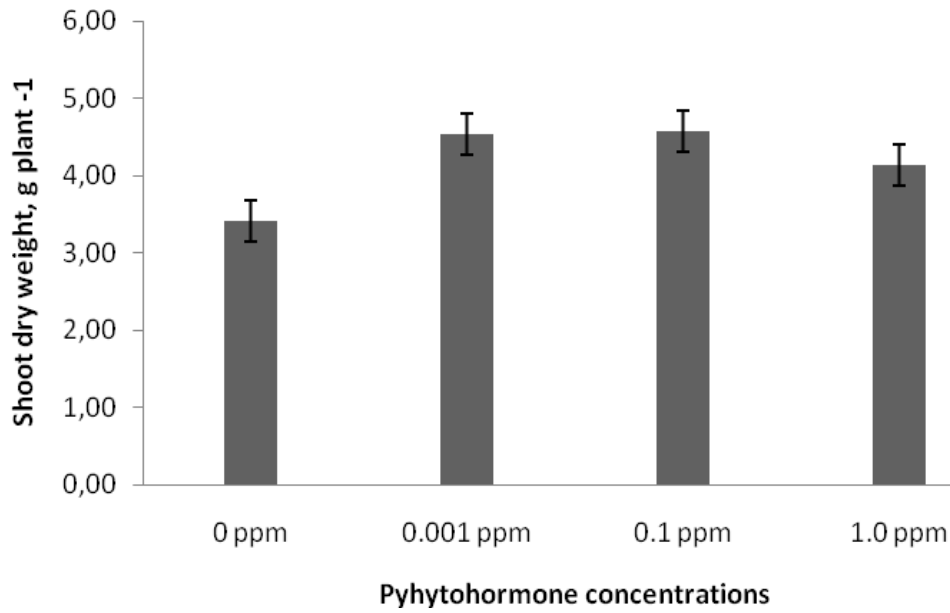


Figure 5. The influence of phytohormone concentrations on soybean shoot dry weight

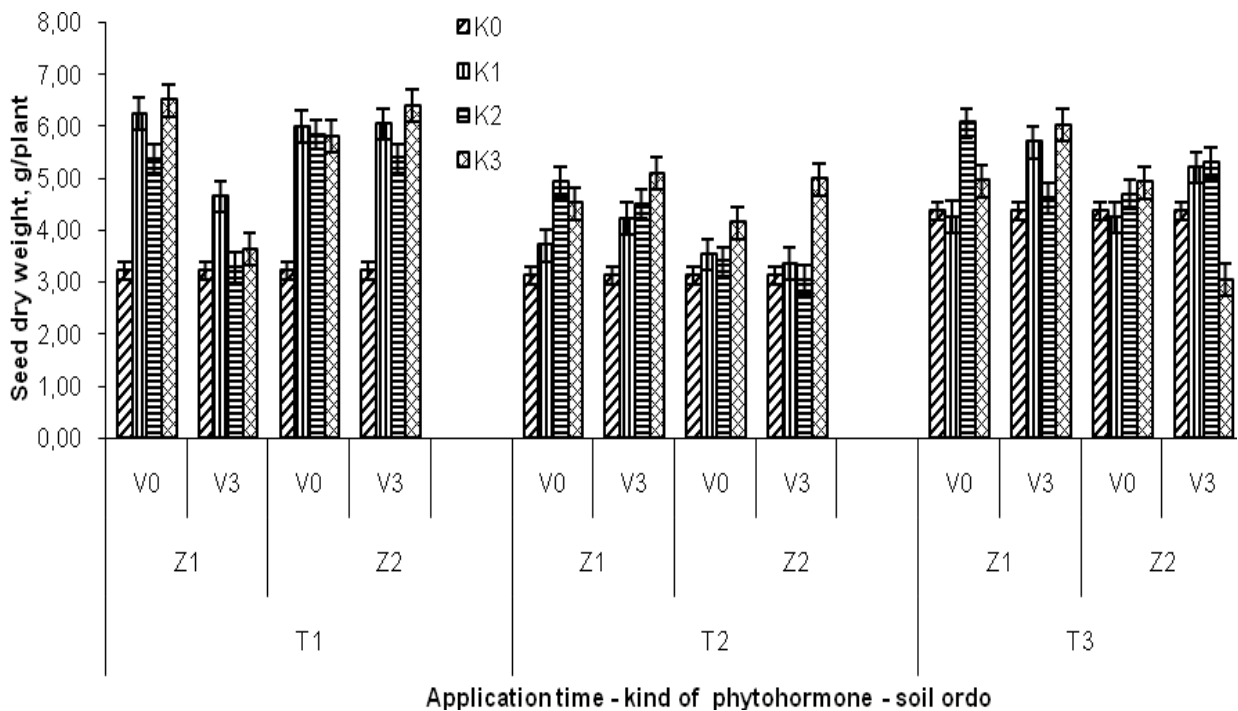


Figure 6. Effects of phytohormones concentration (K), application time (V), kind of phytohormones (Z) and soil ordo (T) on soybean seed dry weight. (Note: K<sub>0</sub> = 0 ppm; K<sub>1</sub> = 0.001 ppm; K<sub>2</sub> = 0.1 ppm and K<sub>3</sub> = 1.0 ppm; V<sub>0</sub> = at planting date; V<sub>3</sub> = at vegetative phase-3; Z<sub>1</sub> = amino acid tryptophan; Z<sub>2</sub> = IAA; T<sub>1</sub> = Alfisols; T<sub>2</sub> = Entisols; T<sub>3</sub> = Vertisols)

## CONCLUSIONS

The exogenous amino acid tryptophan and indole acetic acid (IAA) given as extra cellular growth regulators (PGR) increased the number of root nodules, shoot and seed dry weight of soybean in Alfisols, Entisols and Vertisols. Indole acetic acid (IAA) increased the number of root nodules higher than amino acid tryptophan, so did the application time of extra cellular phytohormone at planting day ( $V_0$ ) which gave higher number of root nodules than when applied at  $V_3$  stage (plant have three leaves).

The treatment combination giving the highest root nodule number was  $K_3Z_2V_3T_2$  (1.0 ppm of IAA given when the plants had three leaves in Entisols) that was 20.67 nodule plant<sup>-1</sup>. Meanwhile, the highest seed weight was obtained from the treatment combination of  $K_3Z_1V_0T_1$  (1.0 ppm of amino acid tryptophan given at planting date in Alfisols) that was 6.51 g plant<sup>-1</sup>, which increased 202% compared with that of no phytohormone treatment. This is consistent with the results of Sudadi (2011), emphasizing that the application of 1.0 ppm of amino acid tryptophan at planting day ( $V_0$ ) gave the highest soybeans seed dry weight in neutral sterile sand planting media.

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