PREPARATION PROCESS OF CURCUMINOID POWDER FROM TURMERIC RHIZOME (*Curcuma longa domestica, Vahl*) AND ITS CHARACTERISTIC AS FOOD INGREDIENTS

PROSES PEMBUATAN BUBUK CURCUMINOID DARI RIMPANG KUNYIT (Curcuma longa domestica, Vahl) DAN KARAKTERISTIKNYA SEBAGAI BAHAN INGREDIEN PANGAN

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ABSTRAK

Studi pada proses pembuatan bubuk curcuminoid dari akar kunyit (Curcuma longa domestica Vahl dan karakteristiknya sebagai bahan pangan telah dilakukan. Hasil percobaan menunjukkan bahwa jenis pelarut, perbandingan antara padatan-larutan dan suhu ekstrasi yang digunakan berpengaruh terhadap rendemen kandungan curcuminoid. Kandungan curcumnoid berkisar antara 11,96% hingga 14,01% (untuk ekstrasi dengan pelarut ethanol 60%), berkisar antara 12,91% hingga 15,20% (ekstrasi dengan pelarut ethanol 80%), dan berkisar antara 3,85 hingga 7,19% (ekstrasi dengan pelarut ethanol 50%). Hasil penelitian menunjukkan bahwa nilai tertinggi dan terbaik kandungan total senyawa fenol tercatat pada ekstrasi dengan ethanol (perbandingan S/L 1 : 50 dan suhu ekstrasi 60°C) dengan nilai 148,99 mg GAE/100 g sedang nilai terendah tercatat untuk ekstrasi menggunakan pelarut 50% (perbandingan S/L 1 : 30) dan suhu ekstrasi 30°C dengan nilai 101,43 mg GAE/100 g. Hasil penelitian juga menunjukkan bahwa nilai tertinggi aktifitas antioksidan tercatat pada ekstraksi menggunakan ethanol 80% dan menggunakan bahan kunyit kering sebanyak 15,0 g dengan nilai 47,65% ; sedang nilai terendah sebagai aktifitas antioksidan pada ekstrasi menggunakan ethanol 50% dan menggunakan bahan kunyit kering sebanyak 1,50 g dengan nilai 20,04%.

Kata kunci : rimpang kunyit (Curcuma longa domestica Vahl), kurkuminoid, proses ekstrasi, sifat karakteristik, ingredien pangan

ABSTRACT

Study on the preparation process of curcuminoid from turmeric root (*Curcuma longa domestica* Vahl) and its characteristics as food ingredient was conducted. The results revealed that the type of solvent, the solid-liquid ratio and extraction temperature were affected to the yield of curcuminoid content. The curcuminoid content ranged from 11.96% to 14.01% (for ethanolic 60% solvent extraction), ranged from 12.91% to 15.20% (ethanolic 80% solvent extraction), and ranged from 3.85% to 7.19% (ethanolic 50% solvent extraction). Results showed that the highest and the best value of total phenolic content was recorded in ethanolic extract (S/L ratio 1 : 50) and extraction temperature 60°C) with value 148.99 mg GAE/100 g; while the lowest for a ethanolic 50% extract (S/L ratio 1 : 30) and extraction temperature 30°C with value 101.43 mg GAE/100 g. Results also showed that the highest value antioxidant activities were recorded in ethanolic 80% extract and 15.00 g ground dry turmeric used with value 47.65%; while the lowest value of ethanolic 50% extract and 1.50 g ground dry turmeric used with value 20.04%..

Keywords: turmeric rhizome (*Curcuma longa domestica Vahl*), curcuminoid, extraction, characteristics, food ingredient

INTRODUCTION

urmeric root (Curcuma longa domestica, Vahl) is a kind of spices that belongs to family Zingebiraceae and its usually used in house hold as food spices (Revathy et al., 2011). Turmeric as an edible and herbaceous plant has been used in traditional medicines for a long time due to its many useful relating with human health. The dried and powdered rhizomes of Curcuma longa are used worldwide as a food-colouring agent because they are having a yellow color (Panlucci et al., 2012).

Curcuminoid is the main natural coloring agent that recognized as a rich source of phenolic compound, consisting of different compounds: curcumin, three demethoxycurcumin, and bisdemethoxycurcumin (Montor et al., 2016). The total of curcuminoids which is about 4 – 6% in wet turmeric rhizome, and turmeric also contains 2 - 4% essential oil and 2 – 3% of fixed oil and various volatile oils; including turmerone, atlantone, and zingiberone. Other constituents include sugars, proteins and resins (Revathy et al., 2016).

The traditional uses of turmeric or natural curcuminoids are many in folk and some of these including medicine. antioxidant (Priyadarsini, 2009), antiinflammatory properties (Jurenka, 2009), anti-carcinogenic effects or antitumor (Wilken et al., 2011), antidiabetic (Wickenberg et al., 2010) and imunomodullatory effects in humans (Rogers *et al.*, 2010). Turmeric has a healthy influence on digestive system and it also enhances the mucin secretion in the digestive tract (Nisar et al., 2015). Although the chemical structure curcumin was determined in the 19th century, the immense value of this molecule is being realized now with several extensive studies on its pharmaceutical and nutraceutical potential (Johnson Mukthar, 2007).

Several *in vitro* and *in vivo* studies confirmed that turmeric extracts have

powerful biological activities, such as antiinflammatory (Jurenka, 2009), bacterial (De et al., 2009), antidepressant et al., 2009), antidiabetic (Kulkarni ((Wickenberg et al., 2010), antitumor (Wilken et al., 2011), immunomodulatory (Rogers et al., 2010) and gastroprotective (Kim et al., 2005) properties. Curcuminoid has been found to be an excellent scavenger of most reactive oxygen species (ROS), a propperty that bestows curcumin with antioxidant activity in normal cells (Priyadarsini, 2014).

The antioxidant activity of turmeric justifies its use in broad range of applications, includina cosmetics (Thornfeldt, 2005), nutraceuticals 2010) and phytomedicines (Anggarwal, Harikumar, (Anggarwal and Priyadarsini, 2014). These medical and functional attributes can be related to turmeric's high content of curcuminoids, especially curcumin, while is considered a chemical marker of this species (Gupta et al., 2012).

Developing a functional ingredient or a phytomedicine requires the use of specific processing technologies, including preparation and extraction of spice active components or chemical markers (Rocha et al., 2008). Extraction is the first step in the isolation of phenolic compounds from agro-industrial residues and plant materials, like curcumin or curcuminoids (Mussatto et al., 2011). Different techniques have been applied to recover antioxidant phenolic compounds from natural sources including solid-liquid extraction with organic solvents, ultrasound-assisted extraction, microwaveassisted extraction, supercritical fluids extraction, and high pressure processes. Among these techniques, solid-liquid extraction is widely employed for phenolics extraction (like curcuminoid) from vegetable/spice sources (Markom et al., 2007).

A number of studies are undertaken to separate curcuminoid pigments by thin layer chromatography (TLC), high-performance thin layer chromatography (HPTLC) and column chromatography (CC) (Revanthy et al., 2011; Pusphakumari et al., 2014). Solvent column extraction followed by chromatography has been most the commonly employ method reported for curcumin/curcuminoid separating from turmeric, and several polar and non-polar organic solvents have been used, including hexane, acetone, ethylacetate, methanol, ethanol, etc. (Priyadarsini, 2014). Of the organic solvent employed, ethanol has been found to be the most preffered solvent for extracting curcumin/curcuminoid.

stationery phase The most commonly used is silica gel with different solvent system including benzene, ethyl acetate, ethanol, chloroform, hexane, and methanol for chromatographic separation (Waghmare et al., 2015). Extraction time, temperature, liquid-to-solid ratio and the extraction-assisted methods are important improvement concerns in the polyphenols (including curcuminoid) yield rate (Hadiyanto et al., 2014). Therefore, with the same technique of even extraction, for different marker compounds in different plant materials; different operating conditions may be required (Noriega et al., 2012; Gokhul et al., 2015). Thereby, the method and processing conditions employed in the extraction of chemical markers from plant raw materials play important roles.

Carboxymethyl cellulose (CMC), a long chain polysaccharide, is a well-known polymer, which is used as a versatile, functional ingredient in a wide variety of processed foods, as a thickener, binding agent, stabilizer, protective colloid and suspending agent (Bar *et al.*, 1995). CMC can dissolve well in water and produce a translucent solution. Its safety has been cofirmed (JECFA, 1990). Thus, it was selected to be used in curcuminoid powder preparation.

Based on these considerations, it is interesting to undertake studies in order to

investigate the relationship between extraction parameters and extract properties on the development of turmeric's functional food. The aim of this extract the was to curcuminoid from turmeric by varying parameters such as extraction solvent, solid/liquid (SL) ratio and extraction temperature. The curcuminoid content, as well as the antioxidant capacity of the extract was also investigated. The watersoluble curcuminoid powder was also study by entrapment a curcuminoid with CMC, as a complex formation. The properties of the powder product have been extensively examined.

MATERIALS AND METHODS Materials and Research Location

Turmeric roots were purchased from local market (Pasar Bogor and Pasar Anyar)-west Java province. Sodium carboxymethyl cellulose (CMC) 1500 cps was purchased from Setia Guna Chemicals grocery at Bogor.

The reference standard of purified curcuminoid was a product of Sigma Chemicals-USA (Sigma C1386) that obtained from PT Brataco Chemicals-Jakarta. Other chemicals used in this research were food grade except for the chemicals for analysis that were either analytical grade or HPLC grade.

The research was conducted in Center of Agro-Based Industry (CABI) – Bogor; Academy of Chemical Analysis Bogor (ACAB) and Institute for Research and Development of Post-Harvest of Agricultural – Bogor.

Methods Dried Turmeric Preparation

Each of fresh turmeric roots from Pasar Bogor and Pasar Anyar was cleaned under running tap water in order to remove adherent sand and clay particles; afterward; turmeric rhizomes were steamed for 10 minutes and sliced into small pieces before drying in a hot air oven at 50°C for about 8 hours.

Dry turmeric was collected and ground into fine powder using a high-speed blender. The dry, ground turmeric was packed in a plastic bag (polypropylene), sealed and kept in the refrigerator (10°C) until used.

Preparation of Turmeric Extracts

Turmeric extracts were prepared by using two solvents; ethanol (at two different concentrations 60% and 80%) and a mixture of ethanol and water 1 : 1 (etanol concentration 50%), according to the protocol as specified by Muhktar and Ghoni (2012) with some modifications.

Approximately 20 am of the sample from the selected, ground, dry turmeric of curcuminoid variety were taken into a thimble and placed in a Soxhlet apparatus. The extraction conditions were performed by factorial design as follows: solid: liquid (S/L) ratio: 1:30, 1:40, and 1:50 at extraction temperature 30°C; 45°C and 60°C with extraction time for 3 hours. Further, the extraction solution obtained from each set of conditions were filtered through Whatman No. 1 filter paper and the clear supernatant was collected. Then the clear supernatant was concentrated on a rotary evaporator. Subsequently, the curcuminoid and total phenolic content were examined. The results of the extraction conditions that provided the highest curcuminoid and total phenolic content were selected for the further experiment.

Preparation of Curcuminoid Powder

Water-soluble curcuminoid powders with different curcuminoid content were prepared. In this study, various quantities of ground turmeric of 1,50; 3,00; 6,00; 7.50 and 15.00 grams respectively were used; and each of it was extracted individually under selected conditions of solid liquid ratio.

After extraction, the mixture was filtered through nylon cloth (320 mesh). The clear solution was adjusted with ethanol to obtain a final volume of 200 ml,

while any excess solvent from the extraction that exceeds 200 ml was removed using a vacuum rotary evaporator (Buchii, Switzerland). The obtained solution was gradually added into 100 ml of 2.5% (w/v) carboxymethylcellulose (CMC) solution at pH 8 -10, stirred for 30 minutes; thereafter it was adjusted to pH 4.0 with 20% ascorbic acid solution.

Maltodextrin (DE = 11) 600 g was added and thoroughly mixed with the curcuminoid solution in a glass container. The viscous mixture was then subjected to drying using a double drum dryer. The drum dryer was operated at a surface temperature of 130° C , with clearances between the drum of 0.04 inch and a rotation speed of 2 rpm. Dried curcuminoid flakes from the drum dryer were then ground into fine powder and the yield was calculated.

Characteristics of Curcuminoid Powder Physical Properties

The physical properties analysis were conducted in focused at color as L*; a* and b* values of the curcuminoid powder. At this point 1 gram curcumonoid powder/100 ml water (dissolved solution) was analyzed using a color instrument (Cronometer, Data color, Spectrafrash SF 600 plus –USA).

Chemical Properties

Analysis for moisture content of turmeric powder was carried out according to its standard methods AOAC (2006); while aw was carried out using aw meter (Novasiana, Switzerland). All the tests were carried out in triplicates.

Determination of Curcuminoid Contents

Determination of curcuminoid content was modified from Thai Herbal Pharmacopoeia (Ministry of Public Health, 2011). Briefly, an amount of 200 mg of dry turmeric powder weighed accurately in a 15 ml screw-cap test and then was added

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with 5 ml ethanol. The mixture was vortexed every 15 minutes for 3 hours and the mixture was centrifuged at 4.000 rpm/min for 15 minutes at room temperature.

The clear supernatant was collected in a 25 ml volumetric flask. The extraction was repeated and the supernatant was deposited until a pale yellow solution was observed. The collected solution was then adjusted to volume 25 ml with ethanol.

The curcuminoid content in the extracted solution was tested and analysis using HPLC instrument (Agilent 1100 series) which is equipped with synergy 4 μ RP80A column. The column temperature was controlled at 35°C. The mobile phase consisted of 2% (v/v) acetic acid aqueous solution and acetonitrile in the ratio of 40: 60 (v/v) with flow rate of 1 ml/min. The injection volume was 5 μ l and the quantitation wavelength was detected at 425 nm.

Total Phenolic Contents

The amount of total phenolic content (TPC) in turmeric powder was estimated and determined using Folin-Ciocalteu method as describe by Wojdylo et al., (2007) with litle modification to each samples without comparing one another. The mechanism is based on the reduction of phosphotungstic acid to phosphotungstic blue and as a result absorbance increase due to rise in the number of aromatic phenolic groups. For this purpose, 50 µl of each prepared extract was separately added to test tubes, each containing 250 µl of Folin-Ciocalteu's reagent and 750 µl of 20% sodium carbonate solution and final volume was made up to 5 ml with distilled water. After 2 hours heated in water bath at 40°C, and absorbance was measured at UV/Visible 765 nm using Spectrophotometer (CECIL CE 7200). In this case, control having all reaction reagents except sample extract. Total polyphenols were estimated and values were expressed as gallic acid equivalent

(GAE; mg gallic acid/100 g) using the following formula : $C = c \times V/m$; where:

C = Total phenolic content (mg/g plant extraction, in GAE)

c =. Concentration of gallic acid (mg/ml)

V = Volume of extract (mL)

m = Weight of turmeric extract (g).

Antioxidant Capacities

The turmeric extracts were analyzed for DPPH (1,1-diphenyl-2-picryhydrazyl) free radical scavenging activity using the method as described by Maizura *et al.*, (2011) with some modifications. Sample solutions were prepared by dissolving 0.0025 mL of sample extract in 10 mL of ethanol. A fresh solution of DPPH mixed in ethanol (6 x 10^{-5} M) was prepared before measurements, 3 mL of this solutions was mixed with 77 µl extract in 1 cm path length microcuvettes.

The mixture samples were vortexed and then placed in the dark at room temperature for about 20 minutes. Decreasing in absorbance was measured afterward, at 515 nm on UV/Visible light Spectrophotometer. Absorbance for a blank sample having the same amount of ethanol and DPPH solution as that for sample extract was also estimated at 515 nm on Spectrophotometer. UV/Visible light Botylated Hidroxy toluene (BHT) and Botylated Hydroxy Anisole (BHA) were used as a standard. Free radical scavenging activity was measured as:

Reduction of absorbance (%) = $[(AA - AB)/AA] \times 100$; where :

AA = Absorbance of blank sample at t = 0 minute

AB = Absorbance of tested extract solution at t = 20 minutes.

Statistical Analysis

Data collected from the experiments were analyzed by complete randomized design (CRD) using SPSS Version 12.00. Analysis of variance (ANOVA) and Duncan's Multiple Range test (DMRT) at p=0.05 were used to determine the differences between treatments

RESULTS AND DISCUSSION Effect of Extraction Conditions on the curcuminoid content

Different solvents with varying polarity have ever used in the study for extraction of curcuminoids from turmeric rhizome by some researchers. However, at this study, only two types of solvents were used, i.e. ethanol (concentration 60% & 80%) and a mixture of ethanol and water 1:1 (ethanol concentration 50%). The results of extraction are presented in Table 1. Under the established conditions, the results showed that the curcuminoid content from Pasar Bogor and Pasar Anyar (local market) ranged from 11.96 to (ethanol solvent 14.01% extraction concentration 60%), ranged from 12.91 to 15.20% (ethanol solvent concentration 80%), and ranged from 3.85 to 7.19% (ethanol - water 1:1 or etanol extraction concentration 50%).

Results revealed that the type of solvent, the solid-liquid (S/L) ratio and the extraction temperature were affected the curcuminoid content. Ethanol extraction (60% and 80% concentration) produced

higher curcuminoid content than a mixture of ethanol and water. The highest curcuminoid content was obtained with the S/L ratio 1:50, an extraction temperature 60°C and extraction with ethanol solvent (80% concentration). Ethanol solvent (80% was resulted concentration) higher than other solvent curcuminoid concentration because it might be due to easier extract the curcumoid content and more polarity. In addition, the expressive extraction of curcuminoid using ethanol solvent 80% may be due to the higher dielectric constant of this solvent, when compared to other proportion used (60% and 50%) (Jouvban et al., 2004).

Mostly, the curcuminoid content was higher in the present study as compared to previous one (Sogi *et al.*, 2010), using ethanol solvent; in which were obtained curcuminoid yields ranging from 3.85 to 15.20%. This, it might be due to the different composition of Curcuma (different sources), extraction conditions and analytical technique employed in the curcuminoid quantification, since Sogi *et al.*, (2010) used spectrophotometry.

Table 1. Influence Of Extraction Conditions (Extraction Temperature And Solid-Liquid Ratio) On Total Curcuminoid Extraction Yield From Ground Dried Turmeric (*)

Solid/Liquid	Extraction	Curcuminoid content (% w/w)		
Ratio	temperature (°C)	Ethanol 60%	Ethanol 80%	Ethanol-water
(g/ml)				
1:30	30	11.96 ^a	12.91 ^a	3.85 ^a
	45	12.31 ^{cd}	13.70 ^b	5.24 ^b
	60	13.92 ^{cd}	14.37 bc	5.27 ^b
1:40	30	12.19 ^{ab}	13.18 ^{ab}	5.36 ^b
	45	13.63 ^c	14.54 ^c	5.81 ^c
	60	13.97 ^{cd}	14.92 ^{cd}	5.98 ^c
1:50	30	12.28 ^b	13.05 ^{ab}	7.03 ^d
	45	12.87 ^{cd}	14.76 ^{cd}	7.10 ^d
	60	14.01 ^d	15.20 ^d	7.13 ^d

^(*) repeated experiments (3x). Different alphabet in the same column indicate statistical differences (p<0.05) by DMRT

Many factors such as the extraction method, solvent composition, extraction time, extraction temperature, the solvent to material extract ratio and extraction pressure, among others, are assumed to significantly influence the efficiency of

curcuminoid extraction (Wakte *et al.*, 2011). The order of curcuminoid content in the extract obtained by various S/L ratio were : 1:50>1:40 and >1:30. As the extracted temperature was increased, the curcuminoid content also increased;

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although indicating that compound was heat stable. This fenomena can affected by with high temperature, so the molucule of curcuminoid was more extracted. According to Panlucci *et al* (2012), the optimum temperature for curcuminoid extraction was found to be at 70°C. The content of curcuminoids many vary in turmeric rhizome grown in different agroclimate zones (Revathy *et al.*, 2011).

The effect of extraction conditions on the phenolic contents can be seen in Table 2. The phenolic contents proved to be dependent on the type of solvent used at a significant level of 5% (p<0.05).

Table 2 shows that the ethanol extraction of ground dry turmeric produced a greater total phenolic content than from the ethanol-water extraction. The Folin-Ciocateu method was reported to react not only with phenols but also any reductive substance present. According to Yang et al., 2006, the high content of total phenols in the extracts might refer to the high properties. antioxidant The extract condition with the highest total phenolic content was found to be the some as the one that had the curcuminoid content.

Table 2. Total Phenolic Content Of Turmeric Extracted Under Various Condtions (*)

Table 21 Total Therione content of Tarmene Extracted order Various Conditions ()					
Solid/Liquid	Extraction	Total phenolic content as mg gallic acid			
Ratio	temperature (°C)	equivalent (GAE)/100 g sample			
(g/ml)		Ethanol 60%	Ethanol 80%	Ethanol-water	
1:30	30	115.30 a	117.67 ^a	101.43 a	
	45	116.75 ^b	119.05 ^a	110.34 ^b	
	60	120.14 ab	122.37 ^{ab}	120.12 ^b	
1:40	30	127.98 ^c	129.84 bc	101.87 ^b	
	45	129.40 ^c	130.04 bc	102.40 ^c	
	60	137.96 ^d	136.25 ^c	121.16 ^c	
1:50	30	124.57 bc	127.36 b	108.67 ^d	
	45	127.80 ^c	131.81 bc	110.19 ^d	
	60	146.25 ^e	148.99 ^d	122.08 ^d	

^(*) repeated experiments (3x). Different letters in the same column indicate statistical differences (p<0.05) by DMRT

Table 3. Properties of Curcuminoid powder

Table 5. Properties of Carcaminola powder							
Ground dry	Moisture		Yield	Curcumionoid	Color Value		e
turmeric used in the extract	(%)	Aw	(%)	(µg/g)	L*	b*	b*
1.50	3.01 ^a	0.24 b	67.51 ^c	164.95 a	97.20 ^a	-9.21 a	50.01 a
3.00	3.48 ^b	0.24 b	63.05 ^a	168.90 b	95.37 ^b	-7.46 ^b	63.70 b
6.00	3.06 ^a	0.20 a	64.60 ab	170.85 b	93.59 ^c	-5.01 ^c	69.55 ^c
7.50	3.05 ^a	0.28 b	65.52 bc	171.27 ^b	92.86 ^c	-4.09 ^d	70.38 ^{cd}
15.00	3.40 b	0.21 a	64.28 ab	172.50 ^C	89.50 ^d	1.18 ^e	71.89 ^d

Different letters in the same column indicate statistical differences (p<0.05) by DMRT

Properties of Curcuminoid Powder

The character properties of curcuminoid powder were presented in Table 3. All the products had low Aw (0.21-0.27) and low moisture content (3.01-3.48%). If the data of moisture obtained

especially was comparable with the earlier results that conducted by Lim *et al.*, (2011), the moisture of curcuminoid powder is not agreement with their findings and is consistent with our results. According to Lim *et al.*, (2011), the

Curcuma longa (powder) contained moisture $14.8 \pm 0.12\%$, while our curcuminoid powder contained moisture 3.01 - 3.48%. The variations between our findings and their mentioned findings is may be due to the difference in varieties, type of materials and drying practice.

Color values L*, a* and b* (Table 3) changed depending on the amount of ground, dry turmeric used. It can be seen also that the increasing the quantity of ground dry turmeric in the extract increased the curcuminoid content as well

as redness values (a*) and yellowness values (b*); while the lightness (L*) value tended to decrease.

The properties of curcuminoid powder that dissolved in water were presented in Table 4. As shown in Table 4, dissolving the powder in distilled water produced acidic solutions with a range of pH from 3.68 – 3.75 that varied with their different color values. A deep yellow solution (high b* values) was obtained from the powder with high curcuminoid content (Table 4).

Table 4. The Color Values of Dissolved Curcuminoid Powder In Water (g/ml water)

Ground turmeric	Color Value				
used for extract (g)	L*	b*	c*		
1.50	95.21 ^e	-4.01 ^c	13.93 ^a		
3.00	93.35 ^d	-6.99 b	26.71 b		
6.00	86.74 ^c	-9.03 ^a	45.82 ^c		
7.50	84.78 ^b	-6.81 b	63.24 ^d		
15.00	68.94 ^a	2.40 d	74.40 ^e		

ns = not significance. Different letters in the same column indicate statistical differences (p<0.05) by DMRT.

Total Phenolic Content

All products (Table 2) showed significant differences in total phenolic content. Results showed that the highest value of total phenolic content was recorded in the ethanolic extract (solid/liquid ratio 1 : 50 and extraction temperature 60°C) with value 148.99 mg GAE/100 g; while the lowest for a mixture of ethanol : water 1 : 1 (solid/liquid ratio 1 : 30 and extraction temperature 30°C) with value 101.43 mg GAE/100 q.

The results of our experiment has related the different method to research work of Kaur and Kapoor (2002), i.e. It have manner of acting the present research outcomes. They evaluated the bioactive phenolic compounds and antioxidant potential of Asian spices and vegetables including *Curcuma longa* for its application in different functional foods..

It was found that total phenolic content in dry powder of turmeric were 115.30 – 148.99 mg GAE/100 mg and our results are not consistent with their work (Wojdylo *et al.*, 2007). Different extraction methods and solvent used might have a

significant effect on extraction yield as well as their antioxidant potential (Nisar *et al.*, 2015). Kim *et al.*, (2011) studied the total phenolic content in the turmeric extract and they found that total phenolic content in turmeric extract was 582.8 mg GAE/100 g extract. This different results, it might be due to the different composition of curcuma (different source), extraction condition and analytical technique in the curcuminoid quantification.

Antioxidant Capacities of Curcuminoid powder (DPPH assay)

Results of antioxidant capacity regarding DPPH scavenging activity extracts are presented in Table 5. Mean values of DPPH showed has enough significant effect of solvent and their amount of ground dry turmeric used on free radical scavenging activity of turmeric powder extract.

Mean values free radical scavenging activities of the ethanol solvent at their different concentrations (50%, 60% and 80%) and at their different ground dry turmeric used for extract (1.50, 3.00, 6.00,

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7.50, and 15.00 g) were 20.04 - 38.30%, 22.20 - 44.72% and 25.69 - 47.65% respectively. While for ascorbic acid (BHT/botylated hydroxy toluene or botylated hydroxy anisole/BHA) was 5.20%.

It was shown in Table 5 that indicated free radical scavenging activities were maximum for ethanol 80% and 60% extract follow by the mixture of ethanol and water 1 : 1 (ethanol 50%) extract.

Table 5. Means Values for DPPH Assay (%) of Turmeric Extracts

Ground dry turmeric used	Antioxidant capacity as mg DPPH/g sample			
for extract (g)	Ethanol 60%	Ethanol 80%	ethanol 50%	
1.50	22.20 ^a	25.69 ^a	20.04 ^a	
3.00	29.02 ^b	32.36 b	24.52 ^b	
6.00	35.38 ^c	37.12 ^c	28.83 ^c	
7.50	38.50 ^d	40.58 ^d	34.18 ^d	
15.00	44.72 ^e	47.65 ^e	38.30 ^e	
BHT/BHA as a standard	5.20	5,20	5,20	

According to Kaur and Kapoor (2002), they assessed the free radical scavenging activity of turmeric powder by extracting it with water and ethanol. It was concluded that ethanol gave more extraction result content than a mixture of ethanol and water and measured the antioxidant potential by DPPH assay. According to them, ethanolic and the mixture of ethanolic and water (1 : 1) extracts of turmeric contained 62.45% and 41.30% antioxidant activity respectively.

Antioxidant activities were also determined by Tilak *et al.*, (2004) of some edible plants including turmeric. Plant materials extracted by using water and methanol as a solvent. According to them, it was mentioned that antioxidant potential of methanolic extract was higher than the water extract. The methanolic and water extracts of turmeric contained 52% and 38% antioxidant activity respectively.

Therefore, based on the above information can be stated that antioxidant activity our research was a little lower content than have been conducted by Kaur and Kapoor (2002) and Tilak et. al. (2004). It might be due to the different composition of curcuma (different source), and extraction condition. The extraction temperature and ethanolic strength exerted

a strong impact on the soluble solid content (Panlucci *et al.*, 2012).

CONCLUSIONS

The type of extraction solvent and extraction condition (temperature and solid/liquid ratio) affected the curcuminoid and total phenolic content, as well as the antioxidant activity of several extracts from dry turmeric (*Curcuma longa domestica Vahl*).

The best solvent used for extraction of curcuminoid content was obtained with the 1.50 extraction S/L ratio on temperature 60°C and the ethanol solvent 80% concentration with curcuminoid content value 15.20 (% w/w); and the total phenolic content value 148.99 mg GAE/100 g. While the extract that has the highest value of an antioxidant capacity was recorded in the ethanolic extract with ethanol 80% concentration, and the amount of ground turmeric used for extract 15.00 mg with antioxidant capacity value as DPPH/g sample was 47.65%.

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REFERENCES

- Anggarwal, B.B. (2010). "Targeting inflammation-induced obesity and metabolic diseases by curcumin and other nutraceuticals". *Annu. Rev. Nutr.*, 30: p. 173 199.
- Anggarwal, B.B. and Harikumar, K.B. (2009). "Potential therapeutic effects of curcumin, the anti-inflammatory agent, against neurodegenerative, cardiovascular, pulmonary, metabolic, auto-immune and neoplastic diseases". *Int. Biochem. Cell Biol.*, 41: p. 40 59.
- AOAC [Association Official Analytical Chemists]. (2006). "Official Methods of Analysis of Association of Official Analytical Chemists International". In: AOAC ed. by: Horwitz, 18 Ed. AOAC Press, Arlington, VA USA.
- Bar, A., Ommen, B.V and Timonen, M. (1995). "Metabolic disposition in rats of regular and enzymatically depolymerized sodium carboxymethylcellulose". Food Chem. Toxicol., 33: p. 901 907.
- Ching, W.Y; Yusoff, Y.B and Wan-Amarina, W.B. (2014). "Extraction of Essential Oil from *Curcuma longa"*. *J. Food Chem. & Nutr. 02 (01)*: p. 01 10.
- De, R., Kurdu, P., Swarnkar., Ramamurthy, T., Chowdhury, A., Nair, G.B and Mukhopadhyay, AK. (2009). "Antimicrobial activity of curcumin against *Heliobacter pylori* isolates from India and during infections in mice". *Antimicrob. Agents Ch. 53*: p. 1592 -1597.
- Gokhul, V.; Yuvapriya, S.; Chandramohan and Muthukumaran, P. (2015). "Isolation and Extraction of Curcumin from Three different varieties of *Curcuma longa, L* A Comparative Study". *Int. J. Pharm. Res & Allied Sci.; Vol. 4 Issue 2*: p. 79 84.

- Gupta, S.C; Patchva, S., Koh, W. And Anggarwal, B.B. (2012). "Discovery of curcumin, a component of golden spice and its miraculous biological activities". *Clin Exp. Pharmacol. P.,* 39: p. 283 299.
- Hadiyanto, H.; Sutanto, A.A and Suharto, Y. (2014). "Ultrasound assisted extraction of antioxidant from *Cloleus tuberosus* peels". *Carphatian J. Food Sci. Technol. 6* (1): p. 58 65.
- Jayaprakasha, KG., Jaganmohan-Rao, L. and Sakariah, KK. (2005). "Chemistry and Biological activities of *Curcuma longa"*. *Trends Food Sci. Technol.*, *16*: p. 533 548.
- JECFA [Joint Expert Commission of Food Additive] (1990). "Toxicological Evaluation of certain food additives". *WHO Food Additive Series 26*: p. 104 123.
- Johnson, JJ and Mukhtar, H. (2007). "Curcumin for chemoprevention of colon cancer". *Cancer Lett. 255*: p. 170 181.
- Jouyban, A., Soltanpour, S and Chan H.K. (2004. "A simple relationship between dielectric constant of mixed solvents with solvent composition and temperature". *Int. J. Pharm. 269*: p. 353-360.
- .Jurenka, J.S. (2009). "Anti-inflammatory properties of curcumin, A major constituent of *Curcuma longa*: A review of preclinical and clinical research". *Altern. Med. Rev. 14*: p. 141 153.
- Kaur, C. and Kapoor, H.C. (2002). "Antioxidant activity and total phenolic content of some Asian vegetables". *Int. J. Food Sci. & Technol., 37*: p. 153 161.
- Kim, D.C., Kim, S.H., Choi, B.H., Back, N.I., Kim. D., Kim, M.J and Kim, K.T. (2005). "*Curcuma longa* extract properties against Gastric Ulcers by blocking H₂ histamine receptors". *Biol. Phar. Bull., 28*: p. 2220 2224.

- Kim, S., Yang, M., Lee, M. and Kang, S. (2011). "Antioxidant activities of hot water extracts from various spices". *Int. J, Mol. Sci. 12*: p. 4120 4131.
- Kulkarni, S.K., Dhir, A. and Akula, KK. (2009). "Potentials of curcumin as an antidepressant". *Sci. Wolrd J. 9*: p. 1233 1241.
- Lim, H.S., Park, S.H., Ghafoor, K., Hwang, S.Y and Park, J. (2011). "Quality and antioxidant properties of bread containing turmeric (*Curcuma longa, L.*) cultivated in South Korea". *J. food Chem., 124*: p. 1577 1582.
- Maizura, M., Aminah, A., and Wan-Aida, W.M. (2011). "Total phenolic content and antioxidant activity of Kesum (*Polygonum minus*), ginger (*Zingiber officinale*) and turmeric (*Curcuma longa*) extract". *Int. Food Res. J., 18*: p. 529 534.
- Markom, M.; Hasan, M.; Daud, W.R.W; Singh, H. and Jaim, J.M. (2007). "Extraction of hydrolysable tannins from *Phyllanthus niruri Linn*: Effects of solvents and extraction methods". *Sep. Purif. Technol. 52*: p. 487 496.
- Ministry of Public Health. (2011).

 Supplement to Thai Herbal
 Pharmacopoeia Vol. I.
 Prachachon, Bangkok Thailand.
- Montar, C., Charoenchai, L., Suksaeree, J. and Sueree, L. (2016)."Quantitation of curcuminoid contents, dissolution profile, and volatile oil content of turmeric capsules produced at some secondary government hospitals". J. Food Drug Anal. 24: p. 493 -499.
- Muhktar, S. and Ghoni, I. (2012). "Antibacterial activity of aqueous and ethanolic extracts of garlic, cinnamon and turmeric against *Escherichia coli ATCC* 25922 and *Bacillus substilisin* DSM". *Int. J.*

- *Applied Bio. Pharma. Tech., 3* : p. 132 137.
- Mussatto, SI;Ballesteros, LF; Martins, S.; and Teixeira, JA. (2011). "Extraction of Antioxidant phenolic compounds from spent coffee grounds". *Sep. & Purif Technol.* 83: p. 173 179.
- Nabati, M.; Mahkam, M. and Heidari, H. (2014). "Isolation and Characterization from powdered rhizomes of turmeric plant marketed in Maragheh City of Iran with Soxhlet Technique". *Iranian Chem. Commun. 2*: p. 236 243.
- Nisar, T., Iqbal, M., Raza, A., Safdar, M., Iftikhar, F. and Waheed, M. (2015). "Free radical scavenging of turmeric (*Curcuma longa*)". *Amer. Eurassion J. Agric & Environ. Sci.* 15 (7): p. 1272 1277.
- Noriega, P., Mafud, DF., Souza, B., Soares-Scott, M., Rivelli, DP., Barros, SBM and Bacchi, EM. (2012). "Applying design of experiments (DOE) to flavonoid extraction from *Passiflora alata* and *Passiflora. edulis*". *Rev. Bras. Farmacogn*: DOI 10.1590/S01002-695X2012. 005.000036.
- Panlucci, VP; Couto, RO, Teixeira, CC and Freitas, LAP (2012). "Optimization of the extraction of curcumin from *Curcuma longa* rhizomes". *Rev. Braz. Farmacogn. 23 (1)*: p. 1 9.
- Peres-Almeida, L., Cherubino, A., and Alves, RJ. (2005). "Separation and Determination of the physicochemical characteristics of curcumin, demethoxy curcumin and bisdemethoxy curcumin". Food Res. Int. 38: p. 1039 1044.
- Priyadarsini, KV. (2009). "Photophysics, Photochemistry and Photobiology of Curcumin: Studies from organic solutions, bio-mimetics and living cells". J. Photochem. Photobiol. C. 10: p. 81-96.

- Priyadarsini, KV. (2014). "The Chemistry of Curcumin: From Extraction to Therapeutic agent". *Molecules 19*: p. 1 22.
- Pusphakumari, KN; Varghese, N. and Kottol, K. (2014). "Purification and Separation of Individual Curcuminoids from spent turmeric oleoresin, a by product from Curcumin Production Industry". *Int. J. Pharm. Sci. & Res.*: p. 3246 3254.
- Revathy, S.; Elamalai, S., Benny, M. and Antony, B. (2011). "Isolation, Purification and Identification of Curcuminoids from Turmeric (*Curcuma longa, L.*) by Column Chromatography". *J. Exper. Sci. 2* (7): p. 21 25.
- Rocha, L., Lucio, EMA., Franca, HS and Sharapin, N. (2008). "Mikania glomerata spreng: Desenvolminto de um produco fitoterapio". *Rev. Bras. Farmacogn., 18*: p. 744 747.
- Rogers, NM., Kireta, S. and Coates, PTH. (2010). "Curcumin induces maturation-arrested dendritic cells that expand regulatory T cells in vitro and in vivo". *Clin. Exp. Immunol., 162*: p. 460 473.
- Sogi, DS., Sharma, S., Oheroi, DPS and Wani, IA. (2010). "Effect of extraction parameters on curcumin yield from turmeric". *J. Food Sci. Technol. Mys., 47*: p. 300 304.
- Thornfeldt, C. (2005). "Cosmeceuticals, containing herbs: Fact, Fiction and future". *Dermatol. Surg., 31* (7): p. 873 880.
- Tilak, JC., Benerjee, M., Mohan, H., and Devasagayan, TPA. (2004). "Antioxidant availability of turmeric in relation to its medicinal and culinary uses". *Phytother., 18*: p. 798 804.
- Wakte, PS., Sachin, BS., Patil, AA., Mohato, DM., band TH and Shinde, DB. (2011). "Optimization of microwave, ultrasonic and

- supercritical carbon dioxide assisted extraction techniques for curcumin from *Curcuma longa*". *Sep. Purif. Technol. 79*: p. 50 55.
- Waghmare, P.; Patingrao, D. and Kadu, P. (2015). "Extraction, Isolation, Purification and Identification of Curcumin: A Review Article". *Europ. J. Biomedical & Pharm. Sci., Vol. 2 Issue 3*: p. 108 123.
- Wickenberg, J., Inge masson, SL and Hlebowicz, J. (2010). "Effects of *Curcuma longa* (turmeric) on post prandial plasma glucose and insulin in healthy subjects". *Nutr. J. 9*: p. 1 5.
- Wilken, R., Veena, MS., Wang, MB and Srivatsan, ES. (2011). "Curcumin: A review of anti-cancer properties and therapeutic activity in head and neck squamous cell carcinoma". *Mol. Cancer 10*: p. 1 19.
- Wojdylo, A., Oszmian'ski, J., and Czemerys, R. (2007). "Antioxidant activity and phenolic compounds in 32 selected herbs". *Food Chem., 105*: p. 940 949.
- Yang, JH., Tseng, YH., Lee, JL and mau, JL. (2006). "Antioxidant properties of methanolic extracts from monascal rice". *LWT Food Sci. & Technol.*, 39: p. 740 747.
- Yen, GC. And Chen, HY. (1995). "Antioxidant activity of various tea extracts in relation to their antimutagenicity". *J. Agric. Food Chem., 43*: p. 27 32.