Genetic Variability, Heritability, and Genotypic Correlation of Soybean Agronomic Characters

Keragaman Genetik, Heritabilitas, dan Korelasi Genetik Karakter Agronomi Kedelai

Heru Kuswantoro^{*}, Moch. Muchlish Adie, Pratanti Haksiwi Putri

Indonesian Legume and Tuber Crops Research Institute Indonesian Agency for Agricultural Research and Development Jl. Raya Kendalpayak Km. 8 Malang 65162, East Java, *E-mail: herukusw@gmail.com

Naskah diterima 07 Desember 2020 ; disetujui untuk diterbitkan 30 Oktober 2021

ABSTRACT

Genetic parameters are important in genetic improvement and variety development. This study aimed to determine the effective characters that can be applied as selection criterion in soybean breeding using genetic parameters. About 100 soybean genotypes were grown in the Muneng Agricultural Technology Research and Assessment Installation from April to July 2020. The trial was conducted using a randomized complete block design. The results showed that high genetic variability was found on days to maturity, number of branches per plant, number of productive nodes per plant, 100seed weight, and seed yield. The high heritability was shown by days to maturity, plant height, number of branches per plant, and 100-seed weight. All phenotypic correlations were significant, except for the correlation between seed yield and days to maturity, plant height, number of branches, and number of productive nodes. The seed yield had no genotypic correlation with all agronomic characters observed. The genotypic correlation was only significant for plant height and number of productive nodes, number of branches and number of filled pods, as well as number of productive nodes and 100-seed weight. Therefore, the improvement of seed yield can be conducted through direct selection using the seed yield parameter or indirectly using the 100seed weight.

Keywords: genetic variability, genotypic correlation, heritability, phenotypic correlation

ABSTRAK

Parameter genetik sangat penting dalam perbaikan genetik dan perakitan varietas tanaman. Penelitian ini bertujuan untuk menentukan karakter yang efektif digunakan sebagai kriteria seleksi dalam pemuliaan kedelai dengan menggunakan parameter genetik. Seratus genotipe kedelai ditanam di Instalasi Penelitian dan Pengkajian Teknologi Pertanian Muneng pada bulan April hingga Juli 2020 dengan rancangan acak kelompok lengkap. Hasil penelitian menunjukkan bahwa keragaman genetik yang tinggi diperoleh pada umur masak, jumlah cabang per tanaman, jumlah buku subur per tanaman, bobot 100 biji, dan hasil biji. Heritabilitas yang tinggi ditunjukkan oleh umur masak, tinggi tanaman, jumlah cabang per tanaman, dan bobot 100 biji. Korelasi fenotipik nyata diperoleh pada semua karakter agronomi, kecuali korelasi antara hasil biji dengan umur masak, tinggi tanaman, jumlah cabang, dan jumlah buku produktif. Hasil biji tidak memiliki korelasi genotipik dengan semua karakter agronomi yang diamati. Korelasi genotipik nyata diperoleh pada tinggi tanaman dengan jumlah buku subur, jumlah cabang dengan jumlah polong isi, dan jumlah buku subur dengan bobot 100 biji. Oleh karena itu, peningkatan hasil biji dapat dilakukan dengan seleksi langsung menggunakan parameter hasil biji atau secara tidak langsung melalui bobot 100 biji.

Kata kunci: heritabilitas, keragaman genetik, korelasi fenotipik, korelasi genotipik

INTRODUCTION

Yield is an important character in food crops, including soybean. Most of soybean plant breeding activities focus on improving yield character. The inheritance of low yield character is a challenge in breeding activities. Besides, yield character is strongly influenced by yield component characters such as plant height, number of branches, number of productive nodes, number of filled pods, and seed size (Dabi *et al.* 2016; Kuswantoro *et al.* 2018). The relationship between yield characters and yield component characters can be considered as selection criteria to produce high yielding soybean plants.

Selection criteria are characters that are considered in the selection process. The selection criteria for obtaining high yielding soybeans can be done by studying the relationship between yield characters and yield component characters. The phenotypic and genotypic correlation can describe the degree of association between the characters. Phenotypic correlation is the relationship between plants characters, while genotypic correlation is the relationship among inherited characters or the breeding value of different characters (Muslimin *et al.* 2013; Faot *et al.* 2019). Despite strongly influence phenotypic correlation, environment does not influence genotypic correlation because environmental factors are excluded in calculation. Genotypic correlation is considered to be more stable and expectedly can increase the accuracy in determining the selection criteria.

The correlation between two characters describes a linear relationship. This suggests that an increase in the value of the first character, would follow by the rise in the second character value, and vice versa. Some characters are reported to have a positive phenotypic correlation with soybean yield include days to flowering, plant height, number of branches, number of productive nodes, number of pods, and 100-seed weight (Krisnawati and Adie 2016; Sulistyo et al. 2018; Kuswantoro et al. 2020). Days to flowering, plant height, and number of pods per plant were also reported to have a positive and significant genotypic correlation with seed yield (Balla and Ibrahim 2017). Faot et al. (2019) stated that days to maturity and number of branches had a positive genotypic correlation with seed yield.

The variability of breeding material population is also a determinant for the success of breeding activities. High variability increases the chance to obtain the desired character in selection process (Handayani and Hidayat 2012; Sulistyo and Mejaya 2018; Karyawati et al. 2019). The variability observed in breeding material population is phenotypic variability and can be seen from each character mean square in variance test. The significance found in variance test generally indicates that the variability exists (Kuswantoro et al. 2018). However, phenotypic variability is derived from the expression of genetic and environmental factors, thus it does not fully represent the genetic characteristics. High genetic variability described by the genetic coefficient of variability (GCV) is considered to have a more critical role as an indicator for successful of plant breeding program.

Genetic variability and heritability are genetic parameters that must be considered in breeding process. The estimated value of both parameters determines the success of breeding activities. Characters with high genetic variability and heritability values are suitable to be used as selection criteria. Heritability describes the effect of genetic factors on phenotypic variability (Akram *et al.* 2016). High heritability means that genetic factors control the character so that it is easily inherited and improved by selection (Karyawati *et al.* 2016; Ali *et al.* 2016). Genetic variability and high heritability are observed on flowering age, plant height, number of productive nodes, number of filled pods, 100seed weight, and seed yield both per plant and per unit area (Ali *et al.* 2016; Kuswantoro *et al.* 2018; Karyawati *et al.* 2019; Jandong *et al.* 2020). Each population has different genetic parameters from other populations. To increase the effectiveness of selection, the genetic parameters must be found out before the selection is conducted. The objective of the research was to determine effective characters that can be used as selection criterion.

MATERIAL AND METHOD

The research was conducted from April to July 2020 at Muneng Agricultural Technology Research and Assessment Installation, East Java, Indonesia, with coordinates of -7°48'12 ", 113°9'44" and 10 m asl. The type of soil at the research location is Alfisol. The plant materials consisted of 100 soybean genotypes consisting of local varieties and breeding lines. The genotype was considered as a fixed model. The experimental design was randomized complete block design with two replications.

The soil was cleared from weeds and previous crop residues. Then the soil was tilled to obtain a friable soil, prepared the drainage channels, and followed by planting the seeds. Each soybean genotype was planted in a 3 m \times 1.2 m (3.6 m²) plot size, with a plant spacing of 40 cm \times 15 cm, two seeds per hill. Inorganic fertilizers of 75 kg ZA + 100 kg SP 36 + 75 kg KCl/ha and organic fertilizer of 5 t/ha manure were applied at planting time. Manual weeding was done at 17 days after planting (DAP). The pest controls were applied at 16, 29, 44, 50, 59 and 68 DAP using supermetrin 50 g/l, deltametrin 25 g/l, fipronil 50 g/l, lambda cyhalothrin 25 g/l, alphamethrin 15 g/l, respectively. Observations were carried out for days to maturity, plant height, number of branches, number of productive nodes, number of filled pods, 100-seed weight, and seed yield at harvesting.

The data were analysed using analysis of variance (Anova). The phenotypic and genotypic variability coefficients and broad-sense heritability were calculated using the expected mean square (Singh and Chaudary 1985). The determination of significant genotypic variability was carried out based on genetic standard deviation. The heritability, phenotypic and genotypic correlations were calculated according to Singh and Chaudary (1985). The descriptive data were performed using Minitab 14. The significance phenotypic and genotypic correlations was determined using t-test based on Falconer (1960) and Dabi *et al.* (2016).

RESULTS AND DISCUSSION

Genotypes showed significant differences for all agronomic characters observed (Table 1). This suggests that at least one genotype differed to other genotypes. It also indicates that there was variability in genotypes based on those characters.

Days to maturity ranged from 68-78 days, with the mean of maturity was 74.8 days (Fig. 1). Days to maturity was not normally distributed (with P value <0.0005). This was due to the tested genotypes that were previously selected with early maturity as a criterion. The graph skewed to the right, meaning that most of genotypes had days to maturity above the average. All genotypes were early maturity. genotype Tgm/Brg-558 (<80 days) had the shortest days to maturity (68 days). The differences in days to maturity among soybean genotypes is closely related to day length and temperature. Long days generally result in soybean plants with long days to maturity (Liu *et al.* 2017).

Plant height was normally distributed with P value = 0.093. No selection in plant height may lead this normal distribution. The average plant height of soybean genotypes was 58.8 cm. Most of soybean

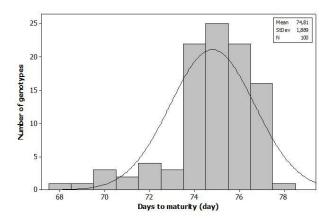


Figure 1. Days to maturity of 100 soybean genotypes. Muneng Res Station, April-July panting season

genotypes grew shorter than the average height, so the graph skewed to the left (Fig. 2). Plant height that mostly appeared was 55 cm. Based on the plant height grouping referred to Adie and Krisnawati (2013), the tested soybean genotypes had plant heights from short to very tall. The tallest genotype, namely MLGG 0103, reached 97.8 cm and it was categorized as very tall. Late maturity soybean genotypes generally show high plant height. However, all genotypes in this study belonged to early maturity. It can be caused by environmental factors such as water availability, day length, and temperature. The length of the day is related to the intensity of solar radiation and changes in temperature. High irradiation intensity and increased temperature cause reduction of stem internodes, while the number of productive nodes increased (Allen Jr. et al. 2018)

Similar to plant height, there was no selection in number of branches per plant. The number of branches per plant was also normally distributed (P value = 0.100), and it ranged from 1-4 branches. A total of 21 genotypes had four branches, and most of genotypes had three branches or lower. The average number of branches per plant in the population was 2.89 branches (Fig. 3). The number of branches and plant height may associate with seed yield per plant (Akram et al. 2011). MLGG 0523 and MLGG 0164 genotypes had four branches per plant and belonged to the tall plant group and with seed yield of 2.01 t and 1.76 t/ha, respectively. The formation of branches in soybean is also influenced by population density per unit area, thus the number of branches per plant is not a stable character. A genotype can have various branches when it was planted at different population densities (Agudamu et al. 2016).

The graph of number productive nodes did not follow a normal distribution (P value = 0.006). The tail of the graph appeared to be on the right, or the graph was leaning to the left. It differed from

 Table 1. Analysis of variance of agronomic characters of 100 soybean genotypes. Muneng Res Station, April-July planting season

Source	Replication	Genotype	Error
Days to maturity	27,380**	6,823**	1,905
Plant height	890,420**	196,241**	41,209
Number of branches per plant	1,217 *	0,813**	0,261
Number of productive nodes per plant	23,805**	5,089**	1,751
Number of filled pods per plant	1257,010**	213,069**	77,969
100-seed weight	0,041	25,259**	2,103
Seed yield	7,826**	0,151**	0,092

* Significant at level of 5%, ** Significant at level of 1%

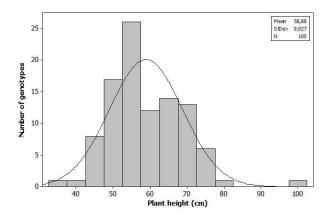


Figure 2. Plant height of 100 soybean genotypes. Muneng Res Station, April-July panting season

number of branches per plant but similar to plant height graph. This may be due to the correlation with other characters that were previously set. The average number of productive nodes was 8.88 (Fig. 4), which ranged from six to 13 nodes per plant. The highest number of productive nodes was seen ini MLGG 0714. The productive nodes are the places where pods are set up, so they play an important role in determining seed yield. The yield per unit area is largely determined by population density. Egli (2013) reported that there is a relationship between number of productive nodes and number of pods per unit area at various population densities. The number of pods per acreage is limited by the number of productive nodes in low population, and there is no correlation between number of productive nodes and number of filled pods in high population.

The distribution of number of filled pods per plant was not normal (P=0.009), as the graph skewed to the left. Similar to the number of productive nodes per plant, this character was also not selected due to its correlation with other characters. The genotypes had 28 to 74 filled pods per plant, with the average of 45 per plant (Fig. 5). IAC-100/Kaba-G-67xBurr-358-T-2-261 had the largest number of filled pods, namely 74 pods. The number of productive nodes and number of filled pods generally have a positive linear relationship. The genotypes with many productive nodes had a large number of filled pods (Kuswantoro et al. 2020). The number of filled pods is a character that determines seed yield per plant. Kuswantoro et al. (2019) reported that number of filled pods associated with seed yield, while Machado et al. (2017) stated that the number of seeds per pod directly affected seed yield.

Selection of genotype resulted in abnormal distribution of 100-seed weight (P=0.005). The genotypes tested generally had 14-19 g/100 seeds,

and therefore those were categorized as medium to large seed sizes following Krisnawati and Adie (2015). The mean weight of 100-seed in the population was 17.2 g, with a minimum value of 10.2 g and a maximum value of 25.2 g (Fig. 6). The graph peak was in the middle. Less than 50% of the genotypes had medium seed size with 100seed weight was <14 g, while the rest (>50% of the genotypes) had larger seed sizes (>14 g). The major 100-seed weight values were 20 g, followed by 14 g. IAC-100/Kaba-G-67xBurr-433-T-15-286 had the largest seed size with 100-seed weight was 25.2 g.

The seed yields of population ranged from 0.95-2.1 t/ha, with the average was 2.07 t/ha (Fig. 7). The graph was skewed to the left, meaning that most of genotypes had lower yield than the average yield. The seed yield was normally distributed (P=0.543). This normal distribution may occur randomly. Genotype Glr 76 had the highest yield of 2.1 t/ha. Yield character is controlled by many

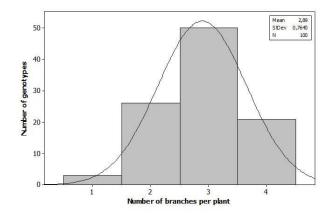


Figure 3. Number of branches per plant of 100 soybean genotypes. Muneng Res Station, April-July panting season

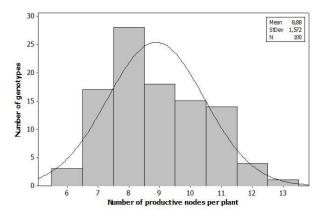


Figure 4. Number of productive nodes per plant of 100 soybean genotypes. Muneng Res Station, April-July panting season

	PCV (%)	GCV (%)	PCV-GCV (%)	GSD	Clasification [†]
Days to maturity (days)	2.80	2.10	0.70	0.52	Broad
Plant height (cm)	18.53	14.97	3.56	14.40	Narrow
Number of branches per plant	25.99	18.64	7.35	0.06	Broad
Number of productive nodes per plant	20.90	14.60	6.3	0.40	Broad
Number of filled pods per plant	26.80	18.26	8.54	16.88	Narrow
100-seed weight (g)	21.50	19.79	1.71	1.79	Broad
Seed yield (t/ha)	16.69	8.34	8.35	0.01	Broad

 Table 2. Coefficients of phenotypic and genotypic variability of agronomic characters of 100 soybean genotypes. Muneng Res Station, April-July planting season

[†]The GCV is broad if GCV> 2GSD and narrow if GCVde 2GSD, PCV = phenotypic coefficient variation, GCV = genotypic coefficient variation, GSD = genetic standard deviation

genes and strongly influenced by other characters, such as number of filled pods, 100-seed weight, and seed weight per acreage. Generally, soybean genotypes that have large seed sizes are also have high yields. However, in this study, there was a medium seeded genotype (10.7 g per 100 seeds) with high seed yield (2.01 t/ha), namely MLGG 0523. Meanwhile, IAC-100 / Kaba-G-67xBurr-433-T with the largest seed size (25.2 g per 100 seeds) vielded lower seeds per hectare, 1.6 t/ha. It shows that 100-seed weight does not always linearly associated with seed yield per hectare. This is in accordance with the study conducted by Krisnawati and Adie (2015), Kuswantoro et al. (2018), and Soares et al. (2013). Plant population also influences seed yield per hectare.

The phenotypic and genotypic coefficients of variability are presented in Table 2. The phenotypic coefficient of variability of all observed characters was greater than the genotypic coefficient of variability. The largest phenotypic coefficient of variability was achieved by number of filled pods per plant, followed by number of branches per plant. The genotypic coefficient of variability does not always follow the magnitude of phenotypic coefficient of variability. The 100-seed weight achieved the highest genotypic coefficient of variability. Small differences between CVP and CVG values showed that the influence of environmental factors is considered to be small, and the appearance of phenotype indicates a genetic value (Reni and Rao, 2013; Malek et al. 2014; Ekka and Lal, 2016).

The criterion using CVP-CVG is less relevant because if CVP of a character is low, CVP-CVG is also low. For example, CVP of days to maturity was low (2.80%); therefore, CVP-CVG was also low (0.70%). It was different from number of filled pods per plant, which had a relatively high CVP (26.80%); although CVG was also high (18.26%), CVP-CVG was also high (8.54%). Such determination of variability is irrelevant because the character is

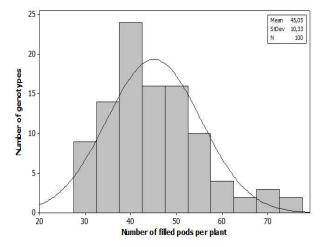


Figure 5. Number of filled pods per plant of 100 soybean genotypes. Muneng Res Station, April-July panting season

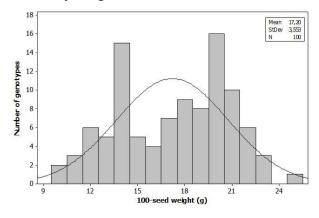


Figure 6. The 100-seed weight of 100 soybean genotypes. Muneng Res Station, April-July panting season

compared to other characters with different environmental influences. Days to maturity and 100seed weight that are less influenced by environment will give a lower CVP-CVG than other characters, such as number of filled pods and number of branches per plant, which are more influenced by environment.

In contrast to coefficient of variability, in determining genetic variability using genetic standard

Table 3. Phenotypic and genotypic variations and heritability of agronomic characters of soybean genotypes. Muneng ResStation, April-July planting season

	Vp	Vg	Ve	H bs	Classification ^a
Days to maturity (days)	4.37	2.46	1.91	0.56	
Plant height (cm)	118.73	77.52	41.21	0.65	High
Number of branches per plant	0.54	0.28	0.26	0.51	High
Number of productive nodes per plant	3.42	1.67	1.75	0.49	Medium
Number of filled pods per plant	145.52	67.55	77.97	0.46	Medium
100-seed weight (g)	13.68	11.58	2.10	0.85	High
Seed yield (t/ha)	0.12	0.03	0.09	0.25	Medium

^aBased on Stansfield (1991)

Table 4. Phenotypic and genotypic correlations of agronomic characters of soybean genotypes. Muneng Res Station, April-July planting season

	HIG	BRC	NOD	POD	W100	YLD
DTM	0.276**	0.284**	0.247*	0.329**	-0.218*	0.118
	0.477	0.465	0.433	0.467	-0.274	0.290
HIG		0.416**	0.774**	0.430**	-0.585**	0.012
		0.640	0.991**	0.747	-0.799	-0.052
BRC			0.440**	0.444**	-0.530**	0.049
		0.790	0.941*	-0.815	0.100	
NOD				0.487**	-0.592**	-0.048
				0.814	-0.939**	-0.241
POD					-0.567**	0.354**
					-0.840	0.771
W100						0.215*
						0.325

Upper = phenotypic correlation, Lower = genotypic correlation, **significant at level of 1%, * significant at level of 5%, DTM = days to maturity, HIG = plant height, BRC = number of branches/plant, NOD = number of reproductive nodes/plant, POD = number of filled pods/plant, W100 = 100-seed weight, YLD = seed yield/ha

deviation, determination criteria are measured based on the character itself, thus environmental factors can be completely eliminated. Although the difference in coefficient of variability was small, based on genetic standard deviation of variability in maturity and 100-seed weight in this study, it was broad due to different measurement unit of the characters. The number of filled pods that had the highest difference in coefficient of variability had narrow criteria based on genetic standard deviation. Therefore, it seems that the use of genetic standard deviation is more relevant in determining the genetic variability of a character.

Heritability is a comparison of genetic variance against phenotypic variance (genotypic and environmental variances). Of seven characters observed, four characters (days to maturity, plant height, and number of branches per plant) had high heritability. In comparison, three characters had low heritability (number of productive nodes per plant, number of filled pods per plant, and seed yield). The highest heritability was achieved by 100-seed weight, while the lowest value was achieved by seed yield per plant (Table 3). High heritability shows that genetic factors are more dominant than environmental factors. Kuswantoro (2017) also reported high heritability on days to maturity, seed size, and seed yield. The changes in heritability value could be influenced by plant material as genetic factor and environmental conditions.

The phenotypic and genotypic correlations of the various agronomic characters were tested. In general, the phenotypic correlation was significant, except for the correlation between seed yield and days to maturity, plant height, number of branches, and number of productive nodes per plant (Table 4). The seed yield significantly correlated with number of filled pods and 100-seed weight. Several studies also reported similar results. Machado *et al.* (2017) stated that plant height associated with number of productive nodes, and number of pods per plant correlated with plant height and seed yield. Jiang *et al.* (2018) and (Mahbub *et al.* 2015) also stated that 100-seed weight had a positive phenotypic correlation with seed yield. Seed yield correlated with number of pods was reported by Bekele and Alemahu (2011). Mahbub *et al.* (2015) said that plant height, number branches per plant, number of pods per plant, 100-seed weight significantly had a positive genotypic and phenotypic correlation with seed yield.

In contrast to the phenotypic correlation, genotypic correlation was generally insignificant (Table 4). There were only three significant genotypic correlations, namely correlation between plant height and number of productive nodes, number of branches and number of filled pods, and number of productive nodes and 100-seed weight. Machado et al. (2017) also reported significant genotypic correlations of plant height with number of productive nodes and number of pods, and number of pods with seed yield. Similarly, Machikowa and Laosuwan (2011) investigated the correlation between number of pods, number of nodes, and number of seeds per plant. Bekele and Alemahu (2011) stated that a significant genotypic correlation was obtained in seed yield with maturity and number of pods.

Interestingly, phenotypic and genotypic correlations of 100-seed weight in this study were significantly and negatively correlated with all agronomic characters, except for seed yield. It means that soybean genotypes with large seed sizes had early maturity, short plant, fewer branches, fewer productive nodes and pods. Bekele and Alemahu (2011) also reported a negative genotypic correlation between 100-seed weight with number of branches and productive nodes. The trend of negative values on phenotypic and genotypic correlations in 100seed weight was also observed by Machikowa and Laosuwan (2011). The linear correlation also shows a significant negative correlation between 100-seed weight and other agronomic characters (Krisnawati and Adie 2016; Kuswantoro 2017).

Inspite of genotypic correlation value was greater than phenotypic correlation, genotypic correlation was not always significant, even though phenotypic correlation was very significant. It occurs because the determination of significant genotypic correlation and phenotypic correlation was different. Determining the significance genotypic correlation involved heritability value, while phenotypic correlation was based on linear correlation (Falconer 1960; Debi *et al.* 2016).

CONCLUSION

Abnormal distribution of a character was caused by the selection of plant materials based on this character or the correlation between the characters with a selected character. Significant genetic diversity was obtained in days to maturity, number of branches per plant, number of productive nodes per plant, 100-seed weight, and seed yield. Not all significant phenotypic correlations were followed by significant genotypic correlations. Significant genotypic correlation was shown by the plant height with number of productive nodes per plant, number of branches per plant with number of filled pods per plant, and number of productive nodes per plant with seed yield. The 100-seed weight could be used as indirect selection criterion for high yield because the phenotypic correlation was significant.

ACKNOWLEDGEMENT

Thanks to the Ministry of Research and Technology, Indonesia which has provided the funding for this research through the program of National Research Priority, Ministry of Research and Technology/National Research and Innovation Agency with the fund from Innovative and Productive Research (Rispro) Mandatory 2020/2021, Lembaga Pengelola Dana Pendidikan (LPDP), Ministry of Finance Republic of Indonesia, with Contract No. 21/E1/PRN/2020.

REFERENCES

- Adie MM, Krisnawati A. 2013. Biologi tanaman kedelai. dalam Sumarno, Suyamto, Widjono A, Kasim H. Kedelai teknik Produksi dan Pengembangan. Puslitbangtan. Bogor: 45-73.
- Agudamu, Yoshihira T, Shiraiwa T. 2016. Branch development responses to planting density and yield stability in soybean cultivars. Plant Production Science 19 (3): 331-339. DOI: 10.1080/ 1343943X.2016.1157443.
- Akram S, Hussain BMN, Al Bari MdA, Burritt DJ, Hossain MA. 2016. Genetic variability and association analysis of soybean (*Glycine* max(L.)Merrill) for yield and yield attributing traits. Plant Gene and Trait 7(13): 1-11. DOI: 10.5376/ pgt.2016.07.0013
- Ali A, Khan SA, Ehsanullah, Ali N, Husain I. 2016. Estimation of genetic parameters in soybean for yield and morphological characters. Pak. J. Agri., Agril. Engg., Vet. Sci.32(2): 162-168
- Allen Jr. LH, Zhang L, Boote KJ, Hauser BA. 2018. Elevated temperature intensity, timing, and duration of exposure affect soybean internode elongation, mainstem node number, and pod number per plant. The Crop Journal 6(2): 148-161.https://doi.org/ 10.2016/j.cj.2017.10.005
- Anderson RL, Bancroft TA. 1952. Statistical Theory in Research. McGraw Hill Book Company, New York USA.

- Balla MY, Ibrahim SE. 2017. Genotypic correlation and path coefficient analysis of soybean [*Glycine max* (L.) Merr.] for yield and its components. Agricultural Research and Technology 7(3):001-005. DOI: 10.19080/ARTOAJ.2017.07.555715
- Bekele A, Alemahu G. 2011. Desirable traits influencing grain yield in soybean. Innovative Systems Design and Engineering 2(3): https://iiste.org/Journals/index.php/ ISDE/article/view/209/94
- Dabi A, Mkebib F, Desalegen T. 2016. Estimation of genetic and phenotypic correlation coeficient and path analysis of yield and yield contributing of bread wheat (*Triticum aseticum* L.) genotypes. International Journal of Natural Resource Ecology and Management. 1(4): 145-154.
- Egli DB. 2013. The relationship between the number of nodes and pods in soybean communities. Journal of Crop Sciences 53 (4): 1668. DOI: 10.2135/ cropsci2012.11.0663.
- Ekka NP, Lal GM. 2016. Study on genetic variability and character association in soybean [*Glycine max* (L.) Merrill] germplasm at vindhyan zone of Uttar Pradesh. AgricSci Digest 36 (1): 69-71.
- Falconer DS. 1960. Introduction to quantitative genetics. pp. 317-318. https://archive.org/details/ introductiontoq00falc/page/318/mode/2up
- Faot MM, Zubaidah S, Kuswantoro H. 2019. Genotypic correlation and path analysis of agronomical traits of soybean (*Glycine max*) lines infected by CpMMV. Biodiversitas 20(6): 1496-1503. DOI: 10.13057/biodiv/ d200602
- Handayani T, Hidayat IM. 2012. Keragaman genetik dan heritabilitas beberapa karakter utama pada kedelai sayur dan implikasinya untuk seleksi perbaikan produksi. Journal of Horticulture 22(4): 327-333
- Jandong EA, Uguru MI. Okechukwu EC. 2020. Estimates of genetic variability, heritability, and genetic advance for agronomic and yield traits in soybean (*Glycine max* (L.)). African Journal of Biotechnology 19(4): 201-206. DOI: 105897/AJB2019.16910
- Jiang GL, Rutto LK, Ren SX. 2018. Evaluation of soybean lines for edamame yield traits and trait genotypic correlation. HortScience 53(12): 1732– 1736.
- Karyawati AS, Sari GN, Waluyo B. 2019. Variabilitas genetik, heritabilitas, dan kemajuan genetik beberapa karakter kuantitatif galur $\rm F_3$ kedelai hasil persilangan. Jurnal Agro. 6(2): 134-143. https://doi.org/10.15575/5175
- Krisnawati A, Adie MM. 2015.Selection of soybean genotypes by seed size and its prospects for industrial raw material in Indonesia.Procedia Food Science 3: 355-363.
- Krisnawati A, Adie MM. 2016. Hubungan antarkomponen morfologi dengan karakter hasil biji kedelai. *Buletin Palawija* 14(2): 49-54

- Kuswantoro H, Artari R, Iswanto R, Imani H. 2020. Family structure of F_5 soybean lines derived from soybean varieties with the main differences on seed size and maturity traits. Biodiversitas 21(6): 2576-85. DOI: 10.13057/biodiv/d210630
- Kuswantoro H, Artari R, Rahajeng W, Ginting E, Supeno A. 2018. Genetic variability, heritability, and correlation of some agronomical characters of soybean varieties. Biosaintifika. Journal of Biology and Biology Education 10(1):9-15. DOI: 10.15294/ Biosaintifika.v10i1.9939
- Kuswantoro H. 2017. The role of heritability and genetic variability in estimated selection response of soybean lines on tidal swamp land. Pertanika Journal of Tropical Agricultural Sciences 40(2): 319-328
- Kuswantoro H. 2019. Performance, similarity and genetic parameters of agronomical characters of soybean [*Glycine max* (L) Merrill.] germplasms. Agr.Nat. Resour. 53(3) 228–236.
- Liu X, Wu J, Ren H, Qi Y, Li C, Cao J, Zhang X, Zhang Z, Cai Z, Gai J. 2017. Genetic variation of worl soybean maturity date and geographic distribution of maturity groups.Breeding Science Preview: 1-12.Doi: 10.1270/jsbbs.16167.
- Machado BQV, Nogueira APO, Hamawaki OT, Rezende GF, Jorge GL, Silveira IC, Medeiros LA, Hamawaki RL, Hamawaki CDL. 2017. Phenotypic and genotypic correlations between soybean agronomic traits and path analysis. Genet. Mol. Res. 16(2): gmr16029696
- Machikowa T and Laosuwan P. 2011. Path coefficient analysis for yield of early maturing soybean.Songklanakarin J. Sci. Technol. 33 (4): 365-368.
- Mahbub MM. Rahman MM, Hossain MS, Mahmud F, Kabir MMM. 2015. Genetic variability, correlation and path analysis for yield and yield components in soybean. American-Eurasian Journal of Agricultural and Environmental Sciences 15(2):231-236.
- Malek MA, Rafii MY, Afroz MSS, Nath UK, Mondal MMA. 2014. Morphological characterization and assessment of genetic variability, character association, and divergence in soybean mutants. *The Scientific World Journal* 2014. https://dx.doi.org/ 10.1155/2014/968796
- Muslimin I, Sofyan A, Islam S. 2013. Parameter genetik pada uji klon jati (*Tectona grandis* L. F) umur 5,5 tahun di Sumatera Selatan. Jurnal Pemuliaan Tanaman Hutan 7(2): 97-106
- Reni YP, Rao YK.2013. Genetic variability in soybean [*Glycine max* (L) Merrill]. International Journal of Plant, Animal and Environmental Sciences 3(4): 35-38.
- Singh RK, Chaudary BD. 1985. Biometrical Methods in Quantitative Genetics Analysis.Kalyani Publishers.318 p.

- Soares MM, Oliveira GL, Soriano PE, Sekita MC, Sediyama T. 2013. Performance of soybean plants as function of seed size: II. Nutritional stress. Journal of Seed Science 35(4): 419-427.
- Stansfiled WD.1991. Teori dan Soal-soal Genetika. Erlangga. Jakarta. 417 p.
- Sulistyo A, Mejaya MJ. 2018. Genetic parameters estimation of agronomic traits in soybean population resistant to whitefly. IOP Conf. Series: Earth and Environmental Science197(2018) 012041. The 2nd International Conference on Biosciences (ICoBio). Doi: 10.1088/1755-1315/197/1/012041
- Sulistyo A, Purwantoro, Sari KP. 2018. Correlation, path analysis and heritability estimation for agronomic traits contribute to yield on soybean. International Symposium on Food and Agro-biodiversity (ISFA). IOP Conf. series: Earth and Environmental Science 102 (2018) 012034. Doi: 10.1088/1755-1315/102/1/ 012034