# PRELIMINARY PHYTOCHEMICAL SCREENING OF Albizia lebbeck STEM BARK

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## ABSTRACT

Preliminary screening of phytochemicals is a valuable step in the detection of the bioactive chemicals in plants which may lead to drug discovery and could also be used for the treatment of various ailments. The aim of this experiment was to examine a preliminary phytochemical screening of *Albizia lebbeck* stem bark. The solvent used for the analysis includes: methanol, petroleum ether and water. The result obtained revealed the presence of alkaloids, tannins, terpenoids, saponins, phenols, flavonoids, glycosides, steroids, carbohydrates and proteins in all the solvent used. This bioactive chemicals and nutrients have therapeutic value and have been reported to be cheap, safe, and effective with no side effects in the body of animals.

**Key words:** Albizia lebbeck; bioactive chemicals; nutrients; solvents; phytochemicals.

#### 1. Introduction

Albizia lebbeck L. is a large deciduous tree belonging to the family Leguminosae (Fabaceae), it is also known as Woman's tongue. The plant is found in Africa (Nigeria, Cameroon, Tanzania, Ghana, Togo, South Africa, Kenya, Zambia, Zimbabawe, Mali, Sudan, Senegal and Niger) and some parts of Asia (Everist, 1986; Hawary *et al.*, 2011). The tree range from medium to large of multi -stemmed spreading habit. The leaves are traditionally used for feeding ruminant animals, the stem and root extracts have been traditionally used for the treatment of asthma, malaria, conjunctivitis and diarrhea (Parrota, 2003; Rashid *et al.*, 2003). Its seeds are easily extracted by hand from the pods by hand or by crushing the pods and winnowing to obtain the oil (Gupta and Chakarbarty, 1964; Hagerman and Butler, 1964; Pal *et al.*, 1995).

Scientific studies showed that the stem, roots and leaves contains tannins, alkaloids, saponins, phenols, flavonoids and terpenoids (Kirsti et al., 2010) and is capable of acting as antioxidants (Siddikov *et al.*, 2007; Sherbeiny *et al.*, 1971), antibacterial (Suzgec *et al.*, 2005; Farooq *et al.*, 1954), hypolipidemic (Sala *et al.*, 2002; Gauher *et al.*, 1968; Ghani, 2003), antifungal (Sezik *et al.*, 2004; Shikishima *et al.*, 2001), hepato-protective (Gupta and Chakarbarty, 1964; Kumar and Toky, 1994), anti-allergic (Pal and Achari, 1995; Sissikov *et al.*, 2007) and neuro-protective properties (Tada *et al.*, 2002; Suzgec *et al.*, 2005; El-Sherbeiny *et al.*, 1971).

The study was aimed to examine a preliminary phytochemical screening of *Albizia lebbeck* stem bark.

#### 2. Materials and methods

Site of the experiment

The study was carried out at the College of Agriculture, Dambartta, Kano State, Nigeria.

Sample collection and processing

Fresh stem barks of *Albizia lebbeck* were purchased from a local market in Kano State. It was identified and authenticated by a certified crop taxonomist (Mr. Bello Isyaku). Samples were washed with running tap water, air dried for 14 days and grounded into powder using a mortar and pestle and stored in an air tight well labeled container. 100 g of the grounded sample was dissolved in 500 mL each of petroleum ether, methanol and distilled water for 48 hours, the samples were stored in the refrigerator and the extracts were filtered separately with Whatman filter paper No.1 to obtain filtrates which was subjected to further analysis.

Measurements

Test for Tannins

Ferric chloride test

2 ml of test solution followed by addition of few drops of 5 % ferric chloride solution. The formation of a blue colour indicates the presence of hydrolysable tannins (Kokate, 2005; Harbone, 1998).

Lead acetate test

2 ml of test solution in a test tube, a few drops of 10 % lead acetate solution was added, a formation of a yellow or red precipitate indicated the presence of condensed tannins (Tease and Evans, 1985).

Test for Flavonoids

Pew's test

Zinc powder was added to 2 ml of extract in a test tube followed by drop wise addition of conc. HCl. The formation of purple red or cherry colour indicates the presence of flavonoids (Peach and Tracey, 1956).

Shinoda test

2 ml of the extract was added in the test tube with few fragments of magnesium metal followed by drop wise addition of conc. HCl. Formation of magentia colour indicates the presence of flavonoids (Kokate et al., 2001).

NaOH test

2-3 ml of the extract was put in a test tube and few drops of sodium hydroxide solution were added. Formation of intense yellow colour which becomes colourless on addition of few drops of dilute HCl indicates the presence of flavonoids (Khandewal, 2008).

Test for alkaloids

Iodine test

3 ml of extract was poured into a test tube, few drops of dilute iodine solution was added. Formation of a blue colour which disappears on boiling and reappears on cooling indicates the presence of alkaloids (Khandewal, 2008).

Wagner's test

2-3 ml of the extract was put in a test tube followed by few drops of Wagner's reagent. Formation of reddish brown precipitate indicates the presence of alkaloids (Kokate et al., 2001).

Dragendorff's test

2-3 ml of the extract was put in a test tube, a few drops of Dragendorff's reagent. The formation of an orange brown precipitate indicates the presence of alkaloids (Kokate et al., 2001).

Test for phenol

Ellagic acid test

3 ml of the extract was treated with few drops of 5 % (w/v) glacial acetic acid and 5 % (w/v) NaNO<sub>2</sub> solution in a test tube. Formation of a muddy or niger brown coloration indicates the presence of phenol (Gibbs, 1974).

## **Test for Saponins**

Foam test

2 ml of the extract was diluted with 20 ml of distilled water; it was shaken in a graduated cylinder for 15 minutes. A 1 cm layer of foam indicates the presence of saponin (Kokate et al., 2001).

Test for Sterol

Salkowski's test

2 ml of the extract with 2 ml of chloroform and 2 ml conc.  $H_2SO_4$  were added in a test tube and shaken well. Chloroform layer appeared red and acid layer showed greenish yellow fluorescence indicated the presence of sterols (Kokate et al., 2001).

Test for glycosides

Keller-Kiliani test

3 ml of the extract and glacial acetic acid were put in a test tube followed by one drop of  $FeCl_3$  and conc.  $H_2SO_4$ . Reddish brown appears at the junction of the two liquid layers and upper layer bluish green colour indicates the presence of glycosides (Kokate et al., 2001).

Test for Terpenoids

5 ml of the extract was mixed with 2 ml of chloroform in a test tube and 3 ml of conc.  $H_2SO_4$  was added to form a layer. A reddish brown colouration at the interface is an indication of terpenoids.

Concentrated H<sub>2</sub>SO<sub>4</sub> test

5 ml of the test material was mixed with 2 ml glycial acetic acid in a test tube, followed by the addition of one drop of 5 % FeCl<sub>3</sub> and conc. H<sub>2</sub>SO<sub>4</sub> Formation of a brown ring indicates the presence of glycosides (Kokate et al., 2001).

# Test for carbohydrates

Molisch's test

2 ml of extract in a test tube, add a few drops of 1 % alpha-napthol followed by 2 ml of conc.  $H_2SO_4$ . A reddish violet or purple ring indicates the presence of carbohydrates (Harbone, 1998).

Seliwanoff's test

1 ml of extract in a test tube was added to 3 ml of Seliwanoff's reagent and it was heated on water bath for 10 minutes and cooled followed by 1 % nitrate solution. A formation of red colour confirmed the presence of carbohydrates (Harbone, 1998; Kokate et al., 2001).

Test for protein

Biuret test

3 ml of extract was placed in a test tube followed with the additions 5 drops of melons reagent and 2 ml of 10 % NaOH and mixed thoroughly. Purple or violet colour is an indication of the presence of protein (Harbone, 1998).

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<b>J.</b>	Results

Test	Aqueous	Methanol	Petroleum ether
Test for tannins			
Ferric chloride test	+ve	+ve	+ve
Lead acetate test	+ve	+ve	+ve

# Test for flavonoids

Pew's test	+ve	+ve	+ve
Shinoda test	-ve	+ve	+ve
NaOH test	-ve	+ve	+ve
Test for alkaloids			
Iodine test	-ve	+ve	-+ve
Wagner's test	-ve	+ve	+ve
Dragendorff's test	-ve	+ve	+ve
Test for phenols			
Ellagic acid test	LVA	Lva	Lva
Eliagic acid test	+ve	+ve	+ve
Test for Terpenoids			
Salkowski test	+ve	+ve	+ve
Test for Phlobatannins			
Test for saponins			
Foam test	+ve	+ve	+ve
Test for steroids			
Salkowski's test	+ve	+ve	+ve
Salkowski s test	TVC	TVC	TVC
Test for glycosides	+ve	+ve	+ve
Keller-Kiliani test	+ve	+ve	+ve
Conc. H <sub>2</sub> SO <sub>4</sub> test	+ve	+ve	+ve
Test for carbohydrates			
Molisch's test	+ve	+ve	+ve
Seliwanoff's test	+ve	+ve	+ve
Test for protein			
Biuret test	+ve	+ve	+ve
Dialet test	TYC	TYC	TVC

+ve : positive; -ve : negetive

# 4. Discussions

The results obtained showed the presence of tannins, alkaloids, terpenoids, flavonoids, phenols, steroids, saponins, glycosides, carbohydrates and protein. According to Cowan (1999); Alagbe et al. (2020); Omokore and Alagbe (2019); Shittu et al. (2019) various chemicals such as alkaloids, tannins, saponins, terpenoids and glycosides are often naturally found in plants have been implicated as being responsible for the antimicrobial activities of plants containing them. These chemicals or phytochemicals allows plant to perform multiple biological activities such as: antimicrobial, antifungal, antiviral, antioxidant etc. (Alagbe, 2017, 2018; 2019; Alagbe and Grace, 2019). Medicinal plants contain numerous biologically active compounds such as nutrients and phytochemicals which have physiological actions on the human body (Olowokudejo *et al.*, 2008). Terpenoids are attractants for insects and mites and also exhibit various pharmacological activities i.e., anti-inflammatory, anticancer, antimalarial and antiviral (Knappers *et al.*, 2005; Mahalo and Sen, 1997; Oluwafemi *et al.*, 2020; Shittu *et al.*, 2020). Flavonoids, tannins and saponins have been suggested to posses' antibacterial, anti-thrombitic, antiplasmodic, anti-malaria and antioxidant properties (Saleem *et al.*, 2005; Enzo, 2007; Williams *et al.*, 2000; Cheeke, 2000). Phenols are strong antioxidants and exhibit hypolipidemic properties (Hollman, 2001). Proteins play a key role in tissue maintenance and growth of animals; carbohydrates are needed for energy in the body (Musa *et al.*, 2020)

#### 5. Conclusion

Albizia lebbeck stem bark is rich in various nutrients and useful chemicals which have therapeutic properties without side effects in the body of human and animals. Further research needs to be carried out to ascertain the bioactive chemicals in other parts of the plants.

### 6. Funding

This research received no external funding.

# 7. Conflicts of Interest

The author declare no conflict of interest

#### References

- 1. Alagbe, J.O and Oluwafemi, R.A. (2019). Hematology and serum biochemical indices of growing rabbits fed diet supplemented with different levels of *Indigofera zollingeriana* leaf meal. *Progress in Chemical and Biochemical Research*. 2(4): 170-177.
- 2. Alagbe, J.O and Grace, F.R. (2019). Effect of *Albizia lebbeck* seed oil dietary supplementation on the haematological and serum biochemical parameters of weaner rabbits. *Sumerianz Journal of Agriculture and Veterinary*. 2(10): 96-100.
- 3. Alagbe, J.O., Sadiq, M.R., Anaso, E.U and Grace, F.R. (2019). Efficacy of *Albizia lebbeck* seed oil on the growth performance and carcass characteristics of weaner rabbits. *Sumerianz Journal of Agriculture and Veterinary*. 2(12): 116-122.
- 4. Shittu, M.D., Adejumo, D.O., Ewuola, E.O., Alaba, O., Alagbe, J.O and Ojebiyi, O.O. (2020). Gut morphometric characteristic and ecological response of broiler starter fed varied levels of protein. *Asian Journal of Animal Science*, 14(1):33-39.
- 5. Alagbe, J.O. (2020). Performance, hematology and serum biochemical parameters of weaner rabbits fed different levels of fermented *Lagenaria brevifora* whole fruit extract. *Advances in Research and Reviews*, 2020, 1:5.
- 6. Oluwafemi, R.A., Isiaka Olawale and Alagbe, J.O. (2020). Recent trends in the utilization of medicinal plants as growth promoters in poultry nutrition- A review. *Research in: Agricultural and Veterinary Sciences*. 4(1): 5-11
- 7. **Musa, B.,** Alagbe, J.O., Adegbite Motunrade Betty, Omokore, E.A. (2020). Growth performance, caeca microbial population and immune response of broiler chicks fed aqueous extract of *Balanites aegyptiaca* and *Alchornea cordifolia* stem bark mixture. *United Journal for Research and Technology*, 2(2):13-21.
- 8. Alagbe, J.O., Sharma, R., Eunice Abidemi Ojo, Shittu, M.D and Bello Kamoru Atanda (2020). Chemical evaluation of the proximate, minerals, vitamins and phytochemical analysis of *Daniellia oliveri* stem bark. *International Journal of Biological, Physical and Chemical Studies*, 2(1):16-22.
- 9. Oluwafemi, R.A., Oluwayinka, E.O and Alagbe, J.O. (2020). Effect of dietary supplementation of neem oil (*Azadirachtia indica*) on the growth performance and nutrient digestibility of weaned rabbits. *European Journal of Biotechnology and Bioscience*. 8(5): 6-10.
- 10. Shittu, M.D., Adesina, G.O., Ademola, S.G and Alagbe, J.O. (2020). Intestinal histomorphometry, relative organs weight and apparent digestibility of rabbits fed varied replacement levels of Soyabeans meal with *Ipeomea asirifolia* leaf meal. *Global Academic Journal of Agriculture and Bio sciences*.
- 11. Omokore, E.O and Alagbe, J.O. (2019). Efficacy of dried *Phyllantus amarus* leaf meal as an herbal feed additive on the growth performance, haematology and serum biochemistry of growing rabbits. *International Journal of Academic Research and Development*. 4(3): 97-104.
- 12. Everist, F. L. (1986). "Antioxidant activity and phenolic compounds in 32 selected herbs." Food Chem., 105: 940-949.
- 13. Kirsti, T., Kettunen, H., Bento, M. H., Saarienen, M., Lahtinen, S., and Routonen, N. (2010). "The effect of feeding essential oil on broiler performance and gut microbiota." British Poultry Science, 51: 381-392.
- 14. Alagbe, J.O. (2019). Effects of dried *Centella asiatica* leaf meal as a herbal feed additive on the growth performance, haematology and serum biochemistry of broiler chicken. *International Journal of Animal Research*. 3(23): 1-12.
- 15. Alagbe, J.O. (2019). Role of *Moringa olifera* leaf meal on the growth performance of Poultry/African catfish A review. *International Journal of Advanced Biological and Biomedical Research*. 7(3):249-257.
- 16. Cheeke, P. R. (2000). Actual and potential applications of Yucca schidigera and Quillaja saponaria saponins in human and animal nutrition. *Journal of Animal Science*. 77:1-10.
- 17. Hollman, P. C. (2001). Evidence for health benefits of plant phenols: Local or systemic effects? *Journal of Science, Food and Agriculture*, 81: 842-852.
- 18. Khandelwa, K. R. (2008). Practical Pharmacognosy. 19th edition: Nirali Prakashan, Pune
- 19. Kokate, C. K. (1994). Practical Pharmacognosy, 4th ed: Vallabh Prakasan, Delhi
- 20. Kokate, C. K., Purohit, A. P., Gokhale, S. B. (2001). Carbohydrate and derived Products, drugs containing glycosides, drugs containing tannin Hs, lipids and protein alkaloids. Text book of Pharmacognosy, 7, edition, 2: 133-166.
- 21. Peach, K and Tracey, M. V. (1956). Modern methods of plant analysis. Vol. 3, Springer Verlag, Berlin.
- 22. Shashank, B., Dhyani, D. (2012). Preliminary Phytochemical Screening of Ailanthus excelsa Roxb. *International Journal of Current Pharmaceutical Research*, 4(1): 87-89.
- 23. Treare, G. E., Evans, W. C. (1985). Pharmacognosy 17th edn: Bahiv Tinal, London
- 24. Vasudevan, M. D. and Sreekumari, S. (2007). *Textbook of biochemistry for medical students*. 5th Ed., Jaypee Brothers Medical Publishers (P) Ltd. New Delhi, India
- 25. Gibbs, R. D. (1974). *Chemotaxonomy of Flowering Plants*. Vol. 1, McGill Queen's University Press, Montreal and London.
- 26. Alagbe, J.O. (2018). Effect of different levels of feed added Coriander (Coriandrum sativum) leaves meal on

- the performance, carcass quality, immune response and blood profile of quails (*Corturnix cortunix japonica*). *Pacific International Journal*. 1(3):40-53.
- 27. Alagbe, J.O. (2017). Nutrient evaluation of sweet orange (*Citrus sinensis*) fruit peel as a replacement for maize in the diets of weaner grass cutters. *Scholarly Journal of Agricultural Science*. 6(8):277-282.
- 28. El-Sherbeiny, W.E., W.Y. Haylett and D.S. Awrty, 1971. Assay of the tannin content in *Albizia lebbeck* L. J. Biochem., 32: 1515-1521.
- 29. Farooqi, M.I.H. and K.N. Kaul, 1962. Investigation of the chemical composition of the gum of *Albizia lebbeck* L. trees. J. Sci. Ind. Res., 21: 454-455.
- 30. Farooq, M.O. and I.P. Varshney, 1954. Isolation of palmitic, stearic, arachidic, oleic and linoleic acids from the oil of *Albizia lebbeck* L. Indian J. Chem., 8: 535-536.
- 31. Gaulier, Y.T., 1968. Assay of the amino acids content of *Albizia lebbeck* L. legumes. Arch. Pharm., 313: 18-25.
- 32. Ghani, A., 2003. Medicinal Plants of Bangladesh with Chemical Constituents and Uses. 2nd Edn., Asiatic Society of Bangladesh, Dhaka, Bangladesh, Pages: 603.
- 33. Gupta, S.R. and H.L. Chakarbarty, 1964. Analysis the oil of *Albizia lebbeck* L. seeds grown in India. Indian J. Chem., 9: 756-758.
- 34. Hagerman, A.E. and L.G. Butler, 1994. Assay of condensed tannins or flavonoid oligomers and related flavonoids in plants. Methods Enzymol., 234: 429-437.
- 35. Kumar, N. and O.P. Toky, 1994. Variation in chemical contents of seed and foliage in Albizia lebbek (L.) Benth. of Agroforestry Syst., 25: 217-225.
- 36. Pal, B.C., B. Achari, K. Yoshikawa and S. Arihara, 1995. Saponins from *Albizzia lebbek*. Phytochemistry, 38: 1287-1291.
- 37. Parrotta, J.A., 2003. Albizia Lebbeck L. Forestation, Nurseries and Genetics Resources. RNGR, USDA, USA.
- 38. Rashid, R.B., R. Chowdhury, C.M. Hasan and M.A. Rashid, 2003. Constituents of Albizzia lebbek and antibacterial activity of an isolated flavone derivative. Saudi Pharm. J., 11: 52-56.
- 39. Sagdullaev, B.T., Shakhidoyatov, R.H and Chernenko, M.T. (2001). Lipids and carbohydrates from Althaea nudifolia and A. armeniaca roots. Chem. Nat. Comp. 37(3): 208-212.
- 40. Enzo, A.P. (2007). Traditional plants and herbal remedies used in the treatment of diarrheal disease: Mode of action, quality, efficacy and safety considerations. In: Ahmad I, Aqil F, Owais M, editors. Modern Phytomedicine Turning Medicinal Plants in to Drugs. WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim, 248-260
- 41. Williams, R.J., Spencer, J.P. and Rice-Evans, C. (2004). Flavonoids: antioxidants or signalling molecules? Free Rad. Biol. Med. 36(7): 838-849.
- 42. Saleem, R., Ahmed, M., Ahmed, S. I., Azeem, M., Khan, R. A., Rasool, N. et al. (2005). Hypotensive activity and toxicology of constituents from root bark of Polyalthia longifolia var.pendula. Phytother Res. 19:881–4.
- 43. Sala, A., Recio, M.D., Giner, S., Manez, H and Rios, J.L. (2002). Antiinflammatory and antioxidant properties of Helichysum italicum. J. Pharm and Pharmacol. 54(3): 365-371.
- 44. Sezik, E., Yesilada, K., Shahidoyatov, Z., Nigmatullaev, H and Honda, G. (2004). Traditional Medicine in Uzebekistan. J. Ethnophar. 92: 197-207.
- 45. Shikishima, Y., Takaishi, G., Ito, Y and Ashurmetov, O. (2001b). Terpenoids and g-pyrone derivatives from Prangos tshimganica. Phytochemistry 57: 135-141.
- 46. Suzgec, S., Mericli, A.H., Houghton, P. and Cubuku, B. (2005). Flavonoids of Helichrysum compactum and their antibacterial activity. Fitoterapia 76:269-272.
- 47. Tada, Y., Shikishima, Y., Takaishi, Y and Sibabata, H. (2002). Coumarins and gamma pyrone derivatives from Prangos pabularia. Phytochemistry 59: 649-654.