



## Pla-zno nanocomposite paper for antimicrobial packaging application

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### Abstract

Many food packages (plastic wrappers) today cannot be broken down by the environment. therefore, it is necessary to add natural substances that can make the food package decompose and be resistant to contamination with bacteria. Development of biodegradable polymers from renewable sources is highly desirable for food preservation and packaging, provided they can be effective as plastics or paper that are currently used in packaging, protecting food against microbial contamination, physical damage and chemical reactions (eg oxidation). Poly lactic acid (PLA) is one of the natural polymers produced by several bacteria that grow in crops rich in carbohydrates (such as sugar beets, corn and others). This research aims to insert ZnO nanoparticles and chitosan into a plastic layer of PLA (poly lactic acid) which can improve the antibacterial properties of the resulting packaging. The method used in making PLA-ZnO-chitosan nanocomposite is the precipitation method and the heating method. PLA-ZnO nanocomposites were obtained by varying ZnO nanoparticles 0.5% by weight, 2% by weight, and 3.5% by weight. The results obtained in SEM images show that nanoparticles are homogeneously distributed on the plastic surface. Antimicrobial tests show nanocomposites work effectively in deactivating *E. coli* and *S. aureus*. where it was found that *E. coli* was more susceptible to this type of nanocomposite, where there was a reduction of 3.4 logs to 3.5% ZnO loading in the PLA layer.

**Keywords:** Nanocomposite, ZnO nanoparticles, PolyLactic Acid, Antimicrobials.

### 1. Introduction

Materials that produce environmentally friendly materials have developed rapidly. Meanwhile, utilization composites based on vegetable fiber has been mostly introduced. (1, 2, 3). For more, an advantaged approach is being introduced base on a nano-size base material.

The current trend is to direct research into developing innovative solutions for both antimicrobial functional packaging (active packaging and nanocomposite materials) and low environmental impacts (biodegradable materials, packaging can be recycled by reducing size) [4-14]. In this case, developing biodegradable polymers from renewable sources is highly desirable for preservation of food and packaging, provided that they can be effective as plastic or paper currently used in packaging, protecting food against microbial contamination, physical damage and chemical reactions (eg oxidation). Poly lactic acid (PLA) is one of the naturally occurring polymers produced by several bacteria grown in carbohydrate-rich agricultural plants (sugar beets, corn). So far, PLA has attracted attention for applications in the plastic and paper industry with the fastest growing market from 10-30% [15-16].

Zinc oxide (ZnO) nanoparticles as antimicrobial agents offer several advantages, such as

superior antimicrobial properties, no negative effects on food sensory properties, and compatibility with hard polymer processing conditions [17-20]. Due to the strong antimicrobial activity of ZnO Nanoparticle, it also holds potential applications not only in food decay control [21-23], but also in food safety control by deactivating foodborne pathogens. Among several advantages, it can be increased to use ZnO Nanoparticle for food contact applications, the benefits of which are as follows:

- ZnO non-nano forms have been authorized by EFSA (European Food Safety Authority) as additives for plastic materials and articles, with SML (Specific Migration Limits) of 25 mg / kg of food [24].
- ZnO Nanoparticle shows low toxicity to biological systems (25). In addition, zinc is an important element for human physiological activity; c.a. 10 mg / person / day is needed (CEF EFSA Panel on Food Contact Materials, 2015). Toxicity studies recommend an upper limit of 25 mg / person / day [26].
- In food packaging, transparency remains a determining factor for choosing packaging materials. On the other hand, some foods are susceptible to UV light. To overcome this problem, UV inhibitors are used in polymer processing. Studies show that loading ZnO

Nanoparticle as low as 1% by weight allows good UV blocking performance without interfering with the transparency of the host polymer [27-29].

In this study, we proposed a paper-based packaging material, which was coated with ZnO-PLA nanocomposite. This application can be a paper wrapper for food because there is a risk of relatively high microbiological contamination in cold processed foods. The focus was placed on the in vitro assessment of the antimicrobial activity of ZnO Nanoparticle.

## 2 Materials and Methods

### 2.1 Materials

This experiment is divided into three parts. First, zinc oxide nanoparticles (ZnO Nanoparticle) are synthesized. Next, the synthesized ZnO Nanoparticle is used in paper coatings. Finally, both synthesized ZnO Nanoparticle are tested for compatibility and properties. The chemicals and techniques used to synthesize ZnO Nanoparticle and paper coating films will be discussed. All chemicals used in the experiment are grade analytical reagents (AR Grade, 99.9% pure). Dehydrated zinc acetate [ $\text{Zn}(\text{CH}_3\text{COO})_2 \cdot 2\text{H}_2\text{O}$ ] was obtained from QReC Sdn. Bhd, while sodium hydroxide (NaOH) pellets are obtained from LabChem Sdn. Bhd. Ethanol is obtained from Sigma Aldrich. Poly lactic acid (PLA) (Nature Work TM PLA 3001D) in the form of pellets obtained from Nature Work ® LLC, Minnetonka, MN USA. It has a specific gravity of 1.24 g / cm<sup>3</sup> and a long flow index (MFI) of 15 g / 10 minutes (190°C / 2.16 kg). The solvents used in this study were chloroform obtained from R & M Chemistry. de-ionized water is used during the synthesis process.

### 2.2 Preparation of zno nanoparticle

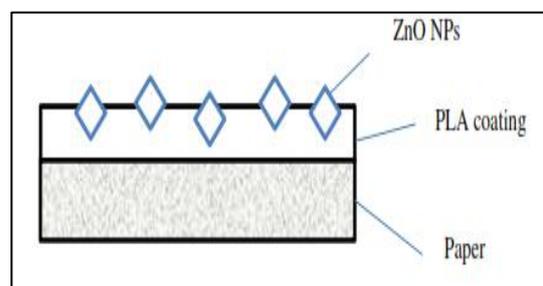
Preparation of ZnO Nanoparticle is carried out using the precipitation method mentioned in Figure. 1. First, 0.2 M of zinc acetate dihydrate is made by dissolving zinc acetate dihydrate with the molecular formula  $\text{Zn}(\text{CH}_3\text{COO})_2 \cdot 2\text{H}_2\text{O}$  in de-ionized water. Then, 0.6 M of sodium hydroxide, NaOH is prepared and added drop by drop in zinc solution dihydrate acetate with constant stirring at room temperature for 2 hours. White deposits are formed from the reaction between zinc acetate dihydrate and NaOH solution. The white sediment is filtered and rinsed with ethanol followed by de-ionized water. This is to ensure that the remaining NaOH in the white deposits is lost. Finally, the white precipitate is dried overnight in an oven at 60 ° C. The white sediments obtained are sent to XRD and FTIR analysis to ensure that pure ZnO Nanoparticle is obtained. ZnO Nanoparticle is then stored to be used to prepare the composite.

### 2.3 Nanokomposit pla / zno preparation

15 g PLA pellets (Polylactic Acid, 4060D, Natureworks) were dissolved using chloroform. Then the dissolved PLA is added with ZnO Nanoparticles at a concentration of 0.5% by weight, 2% by weight and 3.5% by weight (% by weight of PLA) and stirred with strong stirring at room temperature until it is fully dissolved. Furthermore, white kraft paper is bleached (base weight 106 g / m, 7.7% ash content, upper side size) is used as a substrate for coating. Coating is done on the size side. Coating is carried out at the film lab applicator (Elcometer 4340) using a fine rod to store wet film 50 microns to the substrate. After coating, the sample is left to dry overnight at room temperature. The structure of the final packaging material is illustrated in Figure.2.



Gambar 1. Synthesis of ZnO Nanoparticles



Gambar 2. Schematic illustration and structure of food packaging as paper coating from PLA-ZnO

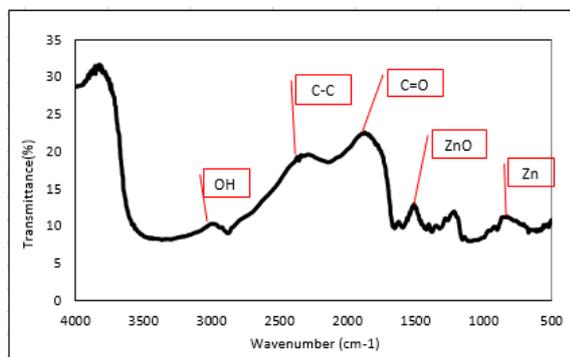
### 2.4 Bacterial activity test

To determine the effect of the addition of chitosan as an antibacterial, it was analysed by the halo zone method, namely: 1. Breeding bacteria on the media 2. Then placed each plate that has been applied to polyurethane on the surface of the media 3. Observation during the incubation period. The antibacterial properties of the plates are measured based on the wide diameter of the clear area that forms around the membrane. As reported by the study of Sawada et al. (2012) antibacterial activity on polyurethane plates using *S. Aureus* and *E.Coli*

bacteria. The sample was incubated for 8 hours at 37°C.

### 3 Result and Discussion

#### 3.1 Characteristics of ZnO nanoparticles

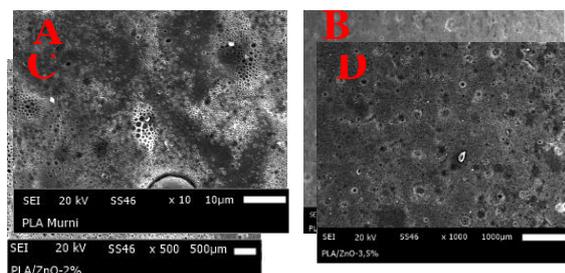


**Gambar 3.** Spectrum of FTIR ZnO Nanoparticle

Figure 3.1 shows the FTIR spectrum of the synthesized ZnO nanoparticles obtained in the range of 4000-500  $\text{cm}^{-1}$ . The band at 3378.49  $\text{cm}^{-1}$  is caused by stretching the vibration of OH groups in water, alcohol and phenol, possibly due to atmospheric humidity. The peak at 1695.5  $\text{cm}^{-1}$  is related to C = group O carboxylic derivatives, which may be due to residues of zinc acetate used in the reaction. The peak at 2328.6  $\text{cm}^{-1}$  is related to the symmetrical stretching vibrations of the carboxylate group (C-C) [30]. Absorption at 832.20  $\text{cm}^{-1}$  is due to the formation of Zn tetrahedral coordination [31]. At a wavelength of 1500.71  $\text{cm}^{-1}$ , it shows the hexagonal group of ZnO [32].

#### 3.2 Characteristics of PLA-zno paper coating nanocomposite

SEM (scanning Electron Microscope)



**Gambar 4.** SEM (A) PLA Murni (B) PLA/ZnO 1%

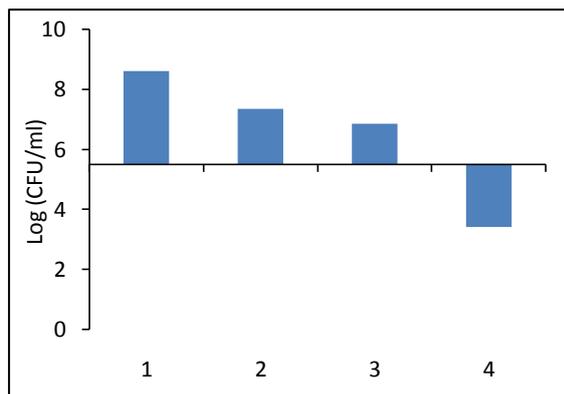
Figure 3.2 shows the morphology of the layer surface using representative SEM taken from each sample. Note that images are presented in pairs when captured at the same point of view but are produced with different detectors: secondary electrons provide more information about the morphology of the sample, while the fibrous electrons contrast with the composition of the material. In general, the surface of the paper is well covered by the coating, showing

good fineness and the absence of surface pores. This is important for the antimicrobial test following the JIS Z 2801. In addition, ZnO is distributed homogeneously, which is clearer in backscatter images (small white dots); on the other hand, some ZnO aggregates persist in the layer (larger white dots). Obviously, with increasing Nanoparticle loading, more Nanoparticle appears on the surface, and thus stronger antimicrobial activity than material can be expected.

#### 3.3 Characteristics of antibacterial tests

The *S. aureus* (Gram-positive) susceptibility test with respect to the active ingredient is shown in Figure 5. For convenience of comparison, the control bacterial concentration at 0 hours was used as the baseline. As expected, the susceptibility of *S. aureus* to follow the concentration of ZnO Nanoparticle - higher concentrations provide a stronger antimicrobial effect. If in log reduction, this corresponds to 1.16, 1.66, and 5.18. Reduction of log greater than 2 indicates success of killing greater than 99%, and hence the material can be described as "antimicrobial" [33-39]. So, in this sense, only a sample of 3.5% -NOO Nanoparticle can be said to be antimicrobial against *S. aureus*. These results are comparable to those reported in previous publications: for example, in the study [40], starch-based coatings containing 1.37% by weight ZnO Nanoparticles induced 1.6 log reductions to *S. aureus*; in a similar study [41], 3% of Nanoparticle weight achieved a 4.3 log reduction against the same bacterial strain; while in work [28], it takes longer (7 days, not 24 hours) to achieve a satisfactory log reduction. The difference between results from different sources can be related to microbe activity. ZnO Nanoparticle in certain morphologies has stronger activity than others [42]. In addition, the smaller the particle size the stronger the antimicrobial efficacy [43].

For the *E. coli* susceptibility test, as shown in Figure 6, a stronger inhibitory effect was observed with all samples containing ZnO Nanoparticle, which each induced 3.15, 3.58 and 4.30 log reduction. Obviously, this test provides evidence that *E. coli* (Gram-negative) is more susceptible to ZnO Nanoparticles than *S. aureus* (Gram-positive). Note that the detection limit of the coating method used is 3.4 log (CFU / mL).



**Gambar 5.** Antimicrobial test to determine S.Aureus



**Gambar 6.** Antimicrobial tests to determine E. Coli

#### 4 Conclusion

The results obtained are in SEM images showing that the nanoparticles are distributed homogeneously on the surface. Antimicrobial tests showed that nanocomposite worked effectively in deactivating E. coli and S. aureus. Furthermore, E. coli was found to be more susceptible to this type of nanocomposite, showing a reduction of 3.4 log to 3.5% ZnO loading in the PLA layer. Based on the results, it can be explained that the ZnO nanoparticles have several active mechanisms, and different antimicrobial testing methods can only activate some of the mechanisms.

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#### References

- [1] Z. Zulkifli dan I.B. Dharmawan, "Analisa Pengaruh Perlakuan Alkalisasi dan Hydrogen Peroksida terhadap Kekuatan Mekanik Komposit Serat Sabut Kelapa Bermatriks Epoxy", *J.POLIMESIN*, Vol. 17, No. 1, pp 41-46, 2019.
- [2] A. Dabet, I. Indra dan H. Hafli, "Aplikasi Teknik Manufaktur Vacuum Assisted Resin Infusion (VARI) untuk Peningkatan Sifat Mekanik Komposit Plastik Berpenguat Serat Abaca (AFRP)", *J.POLIMESIN*. Vol. 16, No. 1, pp. 19-24, 2018.
- [3] I. Mawardi, A. Azwar dan A. Rizal, "Kajian Perlakuan Serat Sabut Kelapa terhadap Sifat Mekanis Komposit Epoksi Serat Sabut Kelapa", *J.POLIMESIN*, Vol. 15, No. 1, pp. 22-29, 2017.
- [4] S. Khalloufi, J. Giasson, C. Ratti, "Activity of freeze-dried mushrooms and fruit", *Can Agr Eng*, Vol.42, pp. 51-56. 2000.
- [5] A. Cagri, Z. Ustunol, ET Rysers, "Antimicrobial edible films and coatings", *J Food Protect*, Vol. 67, pp. 833-843, 2004.
- [6] M. Vargas, A. Albors, A. Chiralt, C. Gonzalez-Martinez, "Characterization of chitosan-oleic acid composite films", *Food Hydrocolloids*, vol. 23, pp. 536-547. 2009.
- [7] G. Findenig; S. Leimgruber; R. Kargl; S. Spirk; K. Stana-Kleinschek; V. Ribitsch, "Making Water Barrier Coatings from Hydrophilic Components", *ACS Appl. Mater. Interface*, vol. 4, pp. 3199-3206, 2012.
- [8] Bourtoom, MS Chinnan, "Preparation and properties of rice starch-chitosan mixed biodegradable film", *LWT-Food Sci Technol*, vol. 41, pp. 1633-1641, 2008.
- [9] Bertuzzi, EF Castro Vidaurre, M. Armada, JC Gottifredi, "Water-Permeability Edible Starch-Based Films", *Journal Of Food Engineering*, vol. 80, pp. 972-978. 2007.
- [10] Perez-Mateos, P. Montero, MC Gomez-Guillen, "Formulation and stability of biodegradable films made from cod gelatin and sunflower oil mixtures", *Food Hydrocolloids*, vol. 23, pp. 53-61. 2009.
- [11] Sivarooban, NS Hettiarachchy, MG Johnson, "Physical and antimicrobial properties of grape seed extract, nicin, and EDTA incorporated edible film soy protein", *International Food Research*, vol. 41, pp. 781-785. 2008.
- [12] MA Del Nobile, A. Conte, AL Incoronato, O. Panza, "Antimicrobial efficacy and release of thymol kinetics from zein films", *Journal of Food Engineering*, vol. 89, Pp. 57-63. 2008.
- [13] P. Suppakul, J. Miltz, K. Sonneveld, SW Bigger, "Active packaging technology with an emphasis on antimicrobial packaging and

- application”, *Journal of Food Science*, vol. 68. Pp. 408-420. 2003.
- [14] Javidi, Z., Hosseini, S. F., & Rezaei, M. “Development of flexible bactericidal films based on poly(lactic acid) and essential oil and its effectiveness to reduce microbial growth of refrigerated rainbow trout”. *LWT – Food Science and Technology*, vol. 72, pp.251– 260. 2016.
- [15] Marra, A., Silvestre, C., Duraccio, D., & Cimmino, S. “Polylactic acid/zinc oxide biocomposite films for food packaging application”. *International Journal of Biological Macromolecules*, vol. 88, pp. 254– 262. 2016.
- [16] Duncan, T. V. “Applications of nanotechnology in food packaging and food safety: Barrier materials, antimicrobials and sensors”, *Journal of colloid and interface science*, vol. 363(1), pp. 1-24. 2011.
- [17] Liu, Y., He, L., Mustapha, A., Li, H., Hu, Z., & Lin, M. “Antibacterial activities of zinc oxide nanoparticles against *Escherichia coli* O157: H7”. *Journal of applied microbiology*, vol. 107(4), pp. 1193-1201. 2009.
- [18] Llorens, A., Lloret, E., Picouet, P. A., Trbojevich, R., & Fernandez, A. “Metallic-based micro and nanocomposites in food contact materials and active food packaging”. *Trends in Food Science & Technology*, vol. 24(1), pp. 19-29. 2012.
- [19] Martins, N. C., Freire, C. S., Neto, C. P., Silvestre, A. J., Causio, J., Baldi, G., Trindade, T. “Antibacterial paper based on composite coatings of nanofibrillated cellulose and ZnO. *Colloids and Surfaces A*”, *Physicochemical and Engineering Aspects*, vol. 417, pp. 111-119. 2013.
- [20] Fernández, A., Picouet, P., & Lloret, E. “Cellulose-silver nanoparticle hybrid materials to control spoilage-related microflora in absorbent pads located in trays of fresh-cut melon”. *International Journal of Food Microbiology*, vol. 142(1), pp. 222-228. 2010.
- [21] Li, X., Xing, Y., Jiang, Y., Ding, Y., & Li, W. “Antimicrobial activities of ZnO powder - coated PVC film to inactivate food pathogens”. *International journal of food science & technology*, vol. 44(11), pp. 2161-2168. 2010.
- [22] Maneerat, C., & Hayata, Y. “Antifungal activity of TiO<sub>2</sub> photocatalysis against *Penicillium expansum* in vitro and in fruit tests”. *International journal of food microbiology*, vol. 107(2), pp. 99-103. 2006.
- [23] Reddy, K. M., Feris, K., Bell, J., Wingett, D. G., Hanley, C., & Punnoose, A. “Selective toxicity of zinc oxide nanoparticles to prokaryotic and eukaryotic systems”. *Applied physics letters*, vol. 90(21), pp. 213-902. 2007.
- [24] El-Feky, O. M., Hassan, E. A., Fadel, S. M., & Hassan, M. L. “Use of ZnO nanoparticles for protecting oil paintings on paper support against dirt, fungal attack, and UV aging”. *Journal of Cultural Heritage*, vol. 15(2), pp. 165-172. 2014.
- [25] Murariu, M., Doumbia, A., Bonnaud, L., Dechief, A. L., Paint, Y., Ferreira, M., Dubois, P. “High-performance polylactide/ZnO nanocomposites designed for films and fibers with special end-use properties”. *Biomacromolecules*, vol. 12(5), pp. 1762-1771. 2011.
- [26] Therias, S., Larché, J.-F., Bussière, P.-O., Gardette, J.-L., Murariu, M., & Dubois, P. “Photochemical behavior of polylactide/ZnO nanocomposite films”. *Biomacromolecules*, vol. 13(10), pp. 3283-3291. 2012.
- [27] F. Gu, et al., “Structural Evaluation and greatly improved luminescence from Dy<sup>3+</sup> + doped ZnO nanocrystals by Li + doping via the combustion method”, *Langmuir*, vol. 20 (9), pp. 3528 – 3531. 2004.
- [28] K. Ravichandrika, P. Kiranmayi, R. Ravikumar, “Synthesis, characterization and antibacterial activity of ZnO nanoparticles”, *International Journal of Pharmacy Sciences*, vol. 4. 2012.
- [29] AK Zak, et al., “Effects the annealing temperature on several structural and optical properties of ZnO nanoparticles”, *Ceram. Int.* vol. 37 (1). Pp. 393 – 398. 2011.
- [30] Pantani, R., Gorrasi, G., Vigliotta, G., Murariu, M., & Dubois, P. “PLA-ZnO 400 nanocomposite films: Water vapor barrier properties and specific end-use characteristics”. *European Polymer Journal*, vol. 49(11), pp. 3471-3482. 2013.
- [31] K. Leja and G. Lewandowicz, “Polymer Biodegradation and Biodegradable Polymers – a Review,” vol. 19, no. 2, pp. 255–266, 2010.
- [32] J. Nanosains, “Modifikasi Bentonit ( Clay ) menjadi Organoclay dengan Penambahan Surfaktan,” vol. 2, no. 1, 2009.
- [33] K. Fukushima, D. Tabuani, and G. Camino, “Poly lactic acid )/ clay nanocomposites : effect of nature and content of clay on morphology , thermal and thermo-mechanical properties,” *Mater. Sci. Eng. C*, vol. 32, no. 7, pp. 1790–1795, 2012.
- [34] A. Araújo, G. Botelho, M. Oliveira, and A. V Machado, “Applied Clay Science Influence of clay organic modifier on the thermal-stability of PLA based nanocomposites,” vol. 89, pp. 144–150, 2014.
- [35] N. P. Resin, “1. Identification Of The Substance / Preparation And The Company / Undertaking,” no. C, pp. 1–10, 2003.
- [36] M. Jamshidian, E. A. Tehrani, M. Imran, and M. Jacquot, “Poly-Lactic Acid : Production ,

- Applications , Nanocomposites , and Release Studies,” vol. 9, pp. 552–571, 2010.
- [37] R. Mansa, C. Huang, A. Quintela, F. Rocha, and C. Detellier, “Applied Clay Science Preparation and characterization of novel clay / PLA nanocomposites,” *Appl. Clay Sci.*, vol. 115, pp. 87–96, 2015.
- [38] L. Avérous, “Polylactic Acid: Synthesis , Properties and Applications,” pp. 2006–2008, 2008.
- [39] C. Kingsland, “*PLA: A Critical Analysis*,” 2010.
- [40] B. Ayana, S. Suin, and B. B. Khatua, “Highly exfoliated eco-friendly thermoplastic starch (TPS) poly (lactic acid)(PLA)/ clay nanocomposites using unmodified nanoclay,” *Carbohydr. Polym.*, vol. 110, pp. 430–439, 2014.
- [41] J. Ren, *Biodegradable Poly (Lactic Acid): Synthesis, Modification, Processing and Applications*
- [42] Esmailzadeh, H., Sangpour, P., Shahraz, F., Hejazi, J., & Khaksar, R. “Effect of nanocomposite packaging containing ZnO on growth of *Bacillus subtilis* and *Enterobacter aerogenes*”. *Materials Science and Engineering*, vol. 58, pp. 1058-1063. 2016.
- [43] O. Yamamoto, “Influence of particle size on the antibacterial activity of zinc oxide,” *International Journal of Inorganic Materials*, 3(7), pp. 643-646. 2001.