Colibacillosis in poultry: A review

Yuvraj Panth

Department of Veterinary Medicine and Public Health, Agriculture and Forestry University, Rampur, Chitwan, Nepal
Correspondence: vetdoc.yuvraj@gmail.com, ORCID: https://orcid.org/0000-0002-7423-0859
Received: June 23; Accepted: September 23; Published: October 25, 2019


This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License.

ABSTRACT

Avian colibacillosis is caused by a group of pathogens designated avian pathogenic Escherichia coli (APEC). Despite being known for over a century, avian colibacillosis remains one of the major endemic diseases afflicting the poultry industry worldwide. Autologous bacterins provide limited serotype-specific protection, yet multiple serogroups are associated with disease, especially O1, O2 and O78 among many others. This article provides the vital information on the epidemiology, diagnosis, control and treatment strategy for avian colibacillosis. A better understanding of the information addressed in this review article will assist the poultry researchers and the poultry industry in continuing to make progress in reducing and eliminating avian colibacillosis from the poultry flocks.

Keywords: Colibacillosis, economics, poultry

INTRODUCTION

Any localized or systemic infection characterized by colisepticemia, hemorrhagic septicemia, coligranuloma (Hjarre's disease), swollen head syndrome, venereal colibacillosis, coliform cellulitis, peritonitis, salpingitis, orchitis, osteomyelitis/synovitis, panophthalmitis, omphalitis/yolk sac infection and enteritis, is referred to as colibacillosis. It is characterized in its acute form by septicemia resulting in death, and in its subacute form by pericarditis, perihepatitis, airsacculusit and other lesions. The presence of exudations in the peritoneal (abdominal) cavity including serum, fibrin and inflammatory cells (pus), are the characteristics. Fibrin is the product of the inflammatory response in chicken that is seen covering the surfaces of multiple organs, including the oviduct, the ovary, the intestine, the alveoli, the heart, the lungs and the liver (Linden, 2015). However, we should not only diagnose based on the lesions to infer an infection with \textit{Escherichia coli} without adding the descriptor “coli” or “coliform”, because other opportunistic bacteria can behave similarly to \textit{E. coli} in secondary infections (Barnes et al., 2008).

It has been observed that avian colibacillosis is a major infectious disease worldwide in birds of all ages, which imparts a significant economic impact on poultry production. Losses are economical, as a result of mortality and decreased productivity of affected birds, mainly around the peak egg production period and throughout the late lay period (Linden, 2015). Avian Pathogenic \textit{Escherichia coli} (APEC) infections are due to stress caused by infections with mycoplasma, infectious bursal disease (IBD) virus, infectious bronchitis (IB) virus, New Castle disease (ND) virus, and environmental influences such as temperature, humidity, ammonia and dust on farms. In addition, \textit{E. coli} can penetrate through the shell when the egg is contaminated by feces, and can spread to chickens during hatching, resulting in high early chick mortality, as well as infection of the yolk sac (Kabir, 2010).

\textit{E. coli} bacteria are a normal inhabitant of the digestive tract of birds and most strains do not cause diseases. When they occur, findings are colisepticemia (blood infection) and yolk sac infection in young chicks; and peritonitis and egg coligranuloma in adults (Vegad, 2015). Often, colibacillosis occurs simultaneously with other diseases, which makes diagnosis and management difficult (Linden, 2015). Rodents, water, flies, insects, food composition, etc. are potential risk factors for APEC (Vandekerchove et al., 2004). The intensification of poultry production and the aggressive expansion of the poultry production systems result in an increase in the incidence of colibacillosis through increased exposure of birds to pathogens and stress (Guabiraba & Schouler, 2015).

Colibacillosis’ prophylaxis is based on strict control of breeding conditions, with priority in biosafety practices and vaccination. Vaccines available in the market are more or less effective, mainly because APEC strains are very diverse and are not widely applied to date. The treatment of colibacillosis is based on antibiotherapy, however, increasing occurrence of multidrug-resistant \textit{E. coli} isolates, has challenged.

\textbf{Etiology}

The disease is caused by certain strains of avian pathogenic \textit{Escherichia coli} (APEC), of which strain O is more common. \textit{Escherichia coli} is a gram-negative, non-acid fast, non-staining,
non-spore-forming bacillus, usually $3 \times 0.6 \, \mu m$ in size, which is included in the Enterobacteriaceae family (Nolan et al., 2013).

*E. coli* is usually motile, often fimbriated, with peritrichous flagella. Somatic (O), flagellar (H) and sometimes capsular (K) antigens are used to differentiate *E. coli* serotypes. The pathogenic strains of the bacteria have virulence factors that allow them to colonize mucosal surfaces and subsequently produce diseases. Age, immune status, the nature of the diet and the strong exposure to pathogenic strains are the factors that predispose animals to develop clinical diseases after colonization by the pathogen. (Quinn et al., 2011). *E. coli* O157: H7, an important zoonotic pathogen, has been isolated from chickens and turkeys; and pigeons can transport shigatoxin-producing *E. coli* (STEC), which can affect people (Nolan et al., 2015).

**Epidemiology**

The organism is present throughout the world and is a normal inhabitant of the gastrointestinal tract of animals and birds at a concentration of $10^6$ per grams, while the dust in the poultry house can contain up to $10^5$-$10^6 / gm$ (Vegad, 2015). Garbage and feces are sources of contamination for both healthy birds and eggs. Transmission is possible through ovoids to newborn chicks. Food, rodent droppings, contaminated well water are the sources of pathogenic *E. coli*. While, isolates of pathogens for poultry commonly belong to certain serogroups, particularly serogroups O78, O1 and O2, but O15 and O55 may also be found to some extent (Chart et al., 2000). According to Amin et al., (2017), the most frequent serogroup of *E. coli* observed was O76 (15.59%), followed by 14.45% O8, 12.17% O1, 7.22% O26, 6.44% O2, 4.94% O114, 4.18% O11, 3.80% O2 and 3.04% every O45 and O84. Avian colibacillosis was found to be widely prevalent in all chicken age groups (9.52 to 36.73%) with an especially high prevalence rate in adult layer birds (36.73%) in Gazipur, Bangladesh (Rahman et al., 2004). However, the prevalence was found to be higher in broilers than in layers in the large Mymensingh district of Bangladesh (Matin et al., 2017) and in the state of Kassala, East Sudan (Omer et al., 2010). The onset of colibacillosis was 32.52% during March 23, 2014 to May 14, 2014 in Gazipur, Bangladesh (Hossain et al., 2015).

A total of 16.32% were colibacillosis alone among 3799 cases, while mixed infection with mycotoxicity, chronic respiratory disease, infectious bursal disease and coccidiosis were 4.13%, 14.71%, 1.07% and 1.23% respectively, during the year 2014/15 at the Central Veterinary Laboratory, Kathmandu (CVL report, 2014/15). It was found that the prevalence of colibacillosis in poultry was dominant, with 25.69% of the cases presented at the Veterinary Teaching Hospital of Agriculture and Forestry University in Chitwan, Nepal, during fiscal year 2014/2015 and 2015/2016 (Gautam et al., 2017). A total of 44 outbreaks of colibacillosis occurred in 2015, of which 112823 chickens were affected and 7803 died, while 155 outbreaks of colibacillosis occurred between January and June 2016, of which 77202 chickens were affected and 3183 died (VEC report, 2015; VÉC report 2016). 14.16% of cases during August 2016 to January 2017 (Shrestha & Shrestha, 2019) and 9.29% during mid-December 2014 to mid-June 2015 (Sharma & Tripathi, 2015) present at the National Laboratory of Avian Disease Research (NADIL), Chitwan, were colibacillosis positive. 31% of liver samples were positive for *E. coli* in poultry in a study conducted at the Surkhet Regional Veterinary Laboratory during September 2016 to January 2017 (Sapkota & Pandey, 2019).
Clinical signs and Lesions
In general, broilers approximately 5 weeks of age are affected (Seneviratna, 1969). Birds of any age can be affected if resistance is reduced by predisposing factors. Chicks less than 10 days old are particularly susceptible if their resistance has been reduced (Seneviratna, 1969).

The severity of the infection may be mild or persistent, and clinical signs may be absent in these cases. Birds with colisepticemia can become lethargic and stop eating and drinking. Severity of the disease may be indicated by the degree of reduced water consumption (Nolan et al., 2015). Chronically affected birds show signs of stunted growth and unthriftiness.

*E. coli* infections in poultry occur in two forms: systemic and localized.

Localized forms of colibacillosis

**Omphalitis / yolk sac infection**

The poor hygiene of the hatchery and the contamination of the eggshell contribute to the inflammation of the navel (omphalitis) of the newborn chicks and to the concurrent infection of the adjacent yolk sac (yolk sacculitis). The yolk sac infection is the most common cause of chick mortality during the first week after hatching, i.e., early chick mortality.

- Affected birds show depression, swollen abdomen and tendency to cluster (huddling)
- Affected birds are often dehydrated, stunted, and may have a vent pasting and an enlarged gall bladder
- Unabsorbed yolk sac
- Distended and smelly yolk sac with abnormality in color and consistency
- The tissue around the navel is often moist and red (inflamed); so the disease is often called "mushy" chick or poult disease
- Peritonitis and hemorrhage on the surface of the intestine

**Swollen head syndrome**

An acute to subacute form of cellulitis that affects the subcutaneous tissues of the periorbital area, giving a swollen appearance to the chicken's face, is swollen head syndrome. Swelling of the head is caused by inflammatory exudate under the skin that accumulates in response to bacteria, usually *E. coli*, after viral infections of the upper respiratory tract, such as avian metapneumovirus, infectious bronchitis virus, etc. (Nolan et al., 2013) and is worse in flocks exposed to high levels of ammonia in the air (Nolan et al., 2015). Mainly broilers and their parents are affected, characterized by the swollen appearance of the face, respiratory symptoms (coughing and sneezing), tracheitis, opisthotonus, green diarrhea with a foul odour (Vegad, 2015).

**Coliform Cellulitis (Inflammatory Process)**

Cellulitis is the inflammation of the loose connective tissue located under the skin. Serosanguinous to caseous exudate plaques in subcutaneous tissues are commonly located on the abdomen or between the thigh and midline. The clinical signs are usually absent. Lesions are
observed in the processing after deburring or during autopsy. The fast, heavy and aggressive fattening lines, more often, are likely to have scratches on the skin, which predisposes to coliform infection and then cellulitis (Nolan et al., 2015).

Males are more often affected than females (Al-Ankari et al., 2001; Blanco et al., 1997) that could be due to its association with weight and aggressiveness. Other risk factors include poor feathering, high flock density, litter quality, as they make birds prone to scratches.

Diarrheal disease (enteritis)

Primary enteritis is a common manifestation of E. coli infections in mammals, including humans, however, it is considered rare in poultry. Diarrhea is the result of enterotoxigenic (ETEC), enterohemorrhagic (EHEC), enteropathogenic (EPEC) or enteroinvasive E. coli (EIEC) infections; with each type having certain virulence factors that determine the characteristics of enteric disease (Nolan et al., 2013). Strains that cause enteritis are able to bind and erase the intestinal epithelium. The birds are dehydrated and show diarrhea. The intestine and the blind of the affected birds are pale and distended with fluid and mucus spots. Characteristic “pits and pedestals” that adhere to blurred lesions covered with adhered E. coli are observed in the intestines (Nolan et al., 2015).

Venereal colibacillosis (acute vaginitis)

Venereal colibacillosis is more common in turkey breeding hens. Vaginitis is acute and most often causes mortality. Cloacal and intestinal prolapse, peritonitis often accompanies vaginitis. The mucosa of the affected hens is thickened, ulcerated and covered with a caseonecrotic membrane. Egg production is reduced with a larger number of small eggs.

Salpingitis and peritonitis

Oviduct inflammation (salpingitis) due to E. coli infection results in reduced egg production with the death of laying hens and breeders. Mortality is sporadic and infections can spread to the peritoneum causing peritonitis. Oviducts are presented with a firm mass or masses of caseous exudate. Cloaca, infected air sacs, vent pecking and prolapse are responsible for infection (Vegad, 2015; Linden, 2015). When E. coli spreads from the oviduct to the abdomen, salpingoperitonitis occurs. In immature birds, the oviduct is infected by extension of the air saculitis that affects the left abdominal air sac (Nolan et al., 2013). Affected birds show signs of sporadic loss of laying, damaged vents, urate leaks, distended abdomen and death.

Coliform orchitis / epididymitis / epididymo-orchitis

The testicles and epididymis of the affected roosters are swollen, firm, irregularly shaped and may have adhesions to adjacent tissues; the necrosis is extensive. The lesions are typically unilateral.

Egg peritonitis

Almost always, egg peritonitis is caused by E. coli. Egg peritonitis is the inflammation of the peritoneum caused by the presence of a broken egg in the abdominal cavity. It is a common cause of sporadic death in laying or breeding hens, but in some flocks it can become the main cause of death before or after reaching maximum production and giving the appearance of a
contagious disease (Rosales, 2019). Dead birds show scattered pieces of yolk, thick yolk, cheese-like material or milky fluid in the abdominal cavity (Vegad, 2015). A complete or partially formed egg may be impacted in the oviduct.

**Systemic forms of colibacillosis**

**Colisepticemia**

Colisepticemia is defined by the presence of virulent *E. coli* in the bloodstream of birds. The duration, degree and outcome of the disease plus the pattern and severity of the lesions are determined by virulence and the number of organisms, balanced with the effectiveness of the host's defenses. The most common form of colibacillosis is characterized by an initial respiratory infection (air sacculitis), often followed by a generalized infection (perihepatitis, pericarditis and septicemia) (Mellata *et al.*, 2003). Colisepticemia progresses through the following stages: acute septicemia, subacute polyserositis and chronic granulomatous inflammation (Nolan *et al.*, 2013).

![Figure 4: Osteoarthrosis; Source: Nolan et al., 2015](image1)

![Figure 3: Multiple focal lesions in liver; Source: Nolan et al., 2015](image2)

![Figure 2: Perihepatitis; Source: Nolan et al., 2015](image3)

Pericarditis, characteristic of colisepticemia, is frequently observed and may be associated with myocarditis. The pericardium becomes cloudy and edematous. Initially, fluid and soft masses of pale exudate accumulate within the pericardial sac, followed by fibrinous exudate. With the progression of the disease, the exudate increases, becomes more cellular and undergoes marriage. (Nolan *et al.*, 2013). Then, the inflamed adherent pericardial sac undergoes organization (fibrosis), resulting in constrictive pericarditis and heart failure. Fibrous perihepatitis and splenomegaly with necrotic tissues in the liver and spleen may be evident.

There is air sacculitis of varying severity and, therefore, respiratory signs such as rales, sneezing and cough are observed. Osteomyelitis, arthritis, tenosynovitis and spondylitis can be sequelae of colisepticemia.

On PM examination, the infected air sacs thicken, are opaque and may contain caseous exudate. Affected birds are normal in appearance and are often found dead with a complete culture, sometimes in acute colisepticemia.
In layers and breeders, most outbreaks are associated with the onset of egg production, but less frequently they occur at a more advanced age. A rapid onset of deep depression, lameness and diarrhea was observed (Zanella et al., 2000). At necropsy, polyserositis (perihepatitis, pericarditis) and peritonitis associated with free yolk in the peritoneal cavity, oophoritis and salpingitis were present in most birds (Nolan et al., 2013).

The chicks are affected within 24-48 hours after hatching. A total of 10-20% mortality is observed with elevation for 2-3 weeks. Initial lesions in chicks consist of congested lungs and splenomegaly. When the bacterium is located in the central nervous system, meningitis and encephalitis are evident in young chicks. Affected birds have neurological signs of rowing and/or neck twisting. Unilateral panophthalmitis is occasionally observed and is characterized by severe inflammation and damage to the internal eye tissue.

**Hemorrhagic Septicemia**

This form occurs in turkeys and is characterized by widespread circulatory alterations, discoloration of subserosal fat, bloody fluid on serous surfaces, pulmonary edema and hemorrhage, and enlargement of the liver, spleen and kidneys. Generalized necrosis of the liver and multifocal necrosis of the spleen are evident on the cut surfaces. (Nolan et al., 2013).

**Coligranuloma**

Coligranuloma (Hjarre's disease) is a sporadic form of colibacillosis that affects chickens, turkeys and quails. Multiple granulomas occur in the liver, proventriculus, ventricle, small intestine, cecum and mesentery.

**Diagnosis**

The diagnosis is based on the isolation and identification of the *E. coli* organism of the typical colibacillosis lesions. Fibrinous pericarditis is highly suggestive of the disease (Seneviratna, 1969). A presumptive diagnosis of *E. coli* infection can be made if the majority of the colonies are characteristically dark with a metallic glow on eosin-methylene blue (EMB) agar, bright pink, with a precipitate around the colonies on MacConkey agar, or yellow in tergitol-7 agar. Differentiation between APEC and commensal *E. coli* isolates can be made by using molecular diagnoses such as PCR (Nolan et al., 2013).

Acute septic diseases caused by Pasteurellae (*Pasteurella, Ornithobacterium, Riemerella*), Salmonellae, Streptococci and other organisms, should be taken in consideration to differentiate the disease. *Chlamydophila*, pasteurellae or streptococci (*Streptococcus, Enterococcus*) can cause pericarditis or peritonitis, and other bacteria, mycoplasmas and *Chlamydophila* can cause air sacculitis. (Nolan et al., 2013).

**Prevention and control**

**Management**

The prevention of colibacillosis is largely a matter of good sanitation and hygiene. Predisposing factors and sources of infection should be ruled out and resolved accordingly. Fumigation of litter with methyl bromide and formaldehyde is effective in destroying *E. coli* (Seneviratna, 1969). Fecal contamination of eggs can be reduced by collecting eggs frequently,
keeping nest material clean, without using eggs from the floor, discarding broken eggs and spraying or disinfecting eggs within 2 hours after laying. The water test for coliform count and treatment with disinfectants is essential. Hot granulation processes destroy \textit{E. coli} (Ekperigin et al., 1990), but care must be taken not to contaminate the finished food again. Colibacillosis has been controlled by the chlorination of drinking water and the use of closed irrigation systems (nipple) (Dhillon & Jack, 1996). Maintaining good air and litter quality is essential to reduce the risk of a flock developing colibacillosis (Davis & Morishita, 2005). Proper ventilation minimizes damage to the respiratory tract by ammonia and, therefore, reduces exposure to bacterial and aerial endotoxins.

Nutritional management should be manipulated, such as food additives that support a healthy immune system and improve survival capacity, adequate protein ratios, increased selenium, increased levels of vitamins A and E and probiotics to promote competitive exclusion (Linden, 2015). Appropriate care should be taken to restrict immunosuppression from stress or other diseases.

### Vaccination

#### Table 1: \textit{E. coli} vaccine types and characteristics (Linden, 2015)

<table>
<thead>
<tr>
<th>Type of vaccine</th>
<th>Description</th>
<th>Results</th>
</tr>
</thead>
</table>
| **Autogenous inactivated** (killed) | • Provides protection against homologous \textit{E. coli} strains  
• No cross protection  
• Breast injection | • Reduced mortality and morbidity due to \textit{E. coli} infection |
| **Commercial modified-live** | • Poulvac \textit{E. coli} 078 (by Zoetis)  
• Cross protection against serotypes O1, O2 and O18  
• Spray | • Reduced morbidity and mortality due to \textit{E. coli} infection  
• Enhanced bird productivity |

### Treatment

Treatment strategies should include actions to control predisposing infections or environmental factors and the early use of antibacterials indicated by susceptibility testing. Different antimicrobial drugs have been used to treat and control colibacillosis. The fortuitous use of antimicrobials has produced a threat of resistance to antimicrobials and, therefore, the availability of effective antimicrobials has decreased. In addition, the development of new drugs is also lacking.

Neomycin (38.00\%) was mainly used for the treatment of colibacillosis, with an efficacy of 92.10\%, with resistance of \textit{E. coli} to neomycin, oxytetracycline, amoxicillin, enrofloxacin and ciprofloxacin, 38.88\%, 50.55\%, 60.68\%, 50.00\% and 30.55\% respectively, during March 23, 2014 to May 14, 2014 in Gazipur, Bangladesh (Hossain et al., 2015). Gentamicin (100\%) was highly sensitive for the treatment of colibacillosis (Hossain et al., 2015).

For an effective treatment, an antibiotic sensitivity test is desired to determine the susceptibility of the bacterial isolate to an antimicrobial therapy. The sensitivity of \textit{E. coli} to gentamicin, neomycin, levofloxacin, tetracycline, enrofloxacin, cephalaxin, azithromycin,
bacitracin, chloramphenicol, doxycycline, amoxicillin and ampicillin was 100%, 100%, 100%, 89%, 84%, 81%, 80%, 80%, 80%, 71%, 50% and 20% respectively (CVL report 2014/15).

Alternatives to antimicrobials such as prebiotics, probiotics, enzymes, digestive acidifiers, vitamins, immune enhancers and anti-inflammatory drugs have been suggested; however, the use of these has not been done so excitingly. Lactobacillus, Bacillus species have been used as probiotics to inhibit colonization of E. coli in the digestive tract.

Conclusion

Colibacillosis is primarily a disease that occurs when birds are stressed or infected with other diseases such as CRD, IBD, etc., that suppress the immune system. E. coli, a normal commensal of the gastrointestinal tract, finds possibilities of multiplication and alters the health of the birds when the birds are predisposed to stress. Care must be taken to prevent the disease by maintaining strict biosecurity measures.

Author Contributions

YP wrote whole article.

Conflicts of Interest

The author declares that there is no conflict of interest.

REFERENCES


