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6 Putra DF, Armaya L, El Rahimi SA

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THE EFFECTS OF RED YAM FLOUR (Ipomoea batatas L.) ON THE GROWTH, SURVIVAL

RATE AND SKIN COLOR OF GOLDFISH (Carrasius auratus)**

Dedi Fazriansyah Putra*, Lia Armaya and Sayyid Afdhal El Rahimi
 Department of Aquaculture, Faculty of Marine and Fisheries, Syiah Kuala University,
 Banda Aceh 23111, Indonesia
 *Corresponding author, e-mail: dfputra@unsyiah.ac.id
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Running title: Effects of red yam flour on goldfish

ABSTRACT

31 This study evaluated the effects of commercial diets supplemented with 0%, 3%, 6%, 32 9% and 12% red yam flour (Ipomoea batatas L.) on the growth, survival rate and skin colour of 33 goldfish (Carrasius auratus). A completely randomized experimental design was developed with 34 five treatments and three replicates. Seventy-five goldfish with the average initial length of 4.4 cm 35 were assigned to fifteen experimental tanks. The experiment had lasted for twenty-eight days. The results showed that the growth and survival rate were not significantly affected by dietary 36 37 treatments (P > 0.05). However, red yam flour significantly affected the skin color of goldfish with 38 the best concentration of 9%. It was concluded that the dietary administration of red yam flour of 39 9% was found to be a suitable dietary supplementation to ensure skin pigmentation as well as there 40 were not bad effects on fish growth and survival rate.

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42 Keywords: Carrasius auratus, Red yam flour, skin color.

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INTRODUCTION

Goldfish (*Carassius auratus*) is one of the most popular ornamental fish in Indonesia. The fish has a body color ranging from red, yellow, green, black and silvery. Since it was first discovered to be nurtured, there are approximately fifteen species of goldfish that have been recognized and favored by public. Goldfish is also one type of ornamental fish that attract many world fish markets. In order to produce the best quality goldfish, a good environment and high nutrients feed are needed. The nutritional content of feed will support color, health and quality of good fish.

For most ornamental fish, many studies focused to achieve high levels of their skin pigmentation together with body shape, fin shape and body size of the fish using sinthtetic pigments and natural sources (Kalinowski et al., 2005; Shahidi et al., 1998). These belong to the most important quality characteristic informing their market levels (Paripatananont et al. 1999). Fish, similar to other vertebrate animals, can not perform de novo synthesis of carotenoids (Goodwin, 1984). Therefore, they depend on dietary supplies to obtain their own natural pigmentation. The color of fish is produced by the presence of chromatophoral cells existing in the dermis layer of skin 59 (Putra et al, 2012). Natural carotenoid substances can be found in plants and fruits in the form of β -60 carotene. Since the high cost of synthetic colourings, many studies on natural compounds such as 61 Chlorella zofin-giensis (Bar et al. 1995) Chlorococcum sp. (Zhang et al. 1997); the green algae 62 Haematococcus pluvialis (Harker et al. 1996; Yuan & Chen 2000 and C. vulgaris (Gouveia et al. 63 1996) have been done. Other latest works have been done using paprika (Minh et al, 2014); Spirulina platensis (Mahdi et al 2013), Medicago sativa (Mahmut et al 2008); and carrot starch 64 (Pardosi et al 2016) as sources of dietary carotenoids. One of alternative sources of a cheap and 65 easily available carotenoid source in Indonesia is red yam. Red yam, among other vegetables, is 66 67 known to contain the highest source of beta carotene. According to Fatimah (2013), red yam 68 contains the highest beta-carotene among other sweet potatoes. It contains 9900 mg (32 967 SI) of 69 beta-carotene per 100 grams. The thicker the colour of red yam is, the higher the content of beta-70 carotene will be. However, studies on the effects of dietary red yam flour in fish feed on the growth 71 and skin colour of goldfish (Carassius auratus) have not been done yet.

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MATERIAL AND METHOD

74 Experimental design

The feeding trial was conducted in the Biology Laboratory of Faculty of Marine and Fisheries of Syiah Kuala University of Banda Aceh, Indonesia. It had been conducted for two months, from July to August 2017. Seventy five goldfish were purchased from a local market. Prior to the experiment, the fish had been acclimatized and fed with commercial ornamental fish for three days. Fish with the average length of 4.4 cm were randomly distributed into fifteen containers (volume 25 L each).

81 This research was done using an experimental method. The design used was Completely 82 Randomized Design (RAL) with 5 treatments and 3 replications. The treatments used were: 83 Treatment A: Control; without adding red yam flour; Treatment B: Red yam flour of 3%; Treatment 84 C: Red yam flour of 6%; Treatment D: Red yam flour of 9%; Treatment E: Red yam flour of 12%. 85 Fish maintenance had been lasted for 28 days and they had been fed three times a day at 09:00 am, 86 12:00 pm and 17:00 pm at each treatment. The amount of feed given was 5% of the average weight 87 of the fish's body.

88 The odd red yam were purchased from traditional market in Banda Aceh city. The red yam 89 used in this study was dried first. Then, the red yam was ground using a laboratory grinder until it 90 become flour, and then mixed the red yam flour with progol as binder in accordance with the 91 dosage (2-3 g / kg of feed) in one container, stirred until evenly and adding water at a volume of 92 150 ml / kg, poured the commercial ornamental fish feed into a different dosage of mixture of red 93 yam flour and dried for 30-60 minutes. If the mixed feed color and smell changed, experimental 94 feed must be re-mixed During this period, the water temperature ranged from 25 ° C-32 ° C with 95 pH from 6 to 9 and the dissolved oxygen was approximately 5 mg/l.

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97 **Observation of fish color change**

98 The color measurements of the tested fish were adopted from Sitorus et al. (2015). Fish 99 color were measured using Toca Color Finder (TCF) observed by 5 panelists who do not have 100 visual impairments (color blindness and farsightedness). Fish color was tranforming into a scores 101 by scaling on color measuring paper. Color observation was done every 7 days for 28 days.

103 **Calculations and statistical analysis**

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103	Calculations and statistical analy	sis
104	Absolute weight growth (AWG)	= Fish weight at time-initial weight
105	Absolute lenght (AL)	= Fish length at time-initial length
106	Survival rate (SR) (%)	= 100 x (final fish number/initial fish number)
107	Specific growth rate (SGR)	= (Initial weight-initial weight) x 100/days

Results were delivered as means± SE (Standard Error). All data were analyzed by one-way 108 ANOVA using SPSS 14.0 for windows (Kalinowski et al., 2005). The level of significance was set 109

at P > 0.05, and Duncan test was used to compare the mean values. 110

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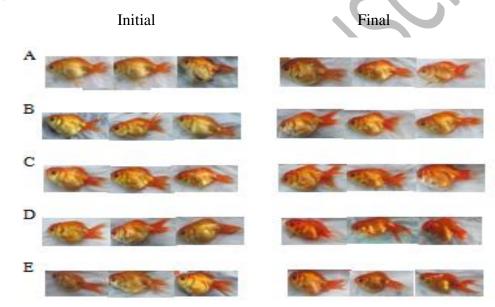
RESULTS AND DISCUSSION

The results showed that red vam flour has siginificant effect on color change of the goldfish 113 114 (*Carracius auratus*) (P<0.05) (table 1). Duncan's advanced test showed that the best color change occured in treatment D (the dose of 9%) and the lowest shown in treatment A (control) (Figure 1). 115 According to upstream et al. (2015), the color enhancement in fish varies according to the fish 116 117 absorption capacity of the pigment type and the dose administered in each treatment. Based on these changes, the color of goldfish in each treatment was different and the dose used was different 118 119 while the caratenoid material used was similar. In this study it is showed that the addition of 9% red yam flour was the best and the most effective treatment to improve the color of goldfish. According 120 121 to Barus et al. (2014), fish takes a longer period to break the carotene material into color pigments if 122 the pigment is present in large quantities. Panjaitan et al. (2016) mentioned that carotene added to 123 the fish feed with excessive doses that to some extent affect the color of fish does not improve the 124 color of the fish. Moreover, it can reduce the color value of fish.

125 An interesting finding seen on day 14, the color of the goldfish began to be brighter color at 126 day-21 (Figure 1). It is in accordance with a previous research done by Sitorus et al. (2015), which 127 stated that the administration of caratenoid in feed for two weeks showed the color improvement of goldfish. The provision of caratenoid for three weeks resulted in maximum color enhancement. 128

129 More than 3 weeks, the color will be stable due to the increase of caratenoid in the pigment cells 130 (chromatophore) of goldfish.

131 The fluctuation of color intensity is caused by the changes of chromatophore cells. The 132 changes are divided into two parts i.e. physiologically and morphologically. Physiological changes 133 are the changes caused by chromatophore cell activities that are spread and concentrated in 134 epidermis cells. The spread of chromatophore pigment stimulates the pigment to absorb the sun perfectly, which then results in an increase of intensity of the fish body. Whereas, the pigment of 135 chromatophore that concentrates or gathers near the nucleus can decrease the intensity of fish body 136 color so that it will become darker or paler. The movement of pigments in the epidermal layer is 137 138 caused by external stimulation such as temperature, pH and light intensity. The morphological 139 changes are due to the amount and composition in the feed containing the caratenoid sources (Sari 140 et al., 2012).



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- 143 Figure 1 Color changes in the goldfish (Carassius auratus) (A: Control; Red yam flour of 0%; B: Red 144 yam flour of 3%; C: Red yam flour of 6%; D: Red yam flour of 9%; E: Red yam flour of 12%)
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The result of ANOVA test showed that feeding of red yam flour has no significant effects

- on absolute weight growth, absolute length, specific growth rate and survival rate (P> 0.05) (Table 147 1).
- 148
- 149 Table 1 Changes in color, growth, survival and specific growth rate of goldfish for 28 days rearing.

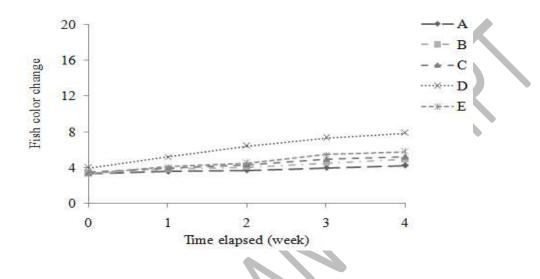
	Test parameters					
Treatment	Color change	Absolute weight growth (gram)	Absolute length (mm)	Specific growth rate (% / day)	Survival rate (%)	
A (control)	0.93 ± 0.31^{a}	2.44 ± 0.57^{a}	11.99 ± 1.60^{a}	2.28 ± 0.26^{a}	100 ± 0.00^{a}	
B (3%)	1.20 ± 0.00^{ab}	2.90 ± 0.33^{a}	13.75 ± 0.75^{a}	2.71 ± 0.46^{a}	100 ± 0.00^{a}	
C (6%)	1.80 ± 0.53^{b}	2.35 ± 0.42^{a}	13.08 ± 2.76^{a}	1.93 ± 0.50^{a}	93.33 ± 11.55^{a}	

D (9%)	3.87 ± 0.31^{d}	2.17 ± 0.65^{a}	12.46 ± 3.22^{a}	2.21 ± 0.53^{a}	100 ± 0.00^{a}
E (12%)	$2.47 \pm 0.31^{\circ}$	2.18 ± 0.43^{a}	10.48 ± 2.93^{a}	2.24 ± 0.22^{a}	100 ± 0.00^{a}

150 Description: Different superscripts in the column show significant differences (P < 0.05) between 151 treatments based on Duncan Test.

152 The results of the study of color changes of goldfish during the study period of 28 days in 153 each treatment can be seen in Figure 2.

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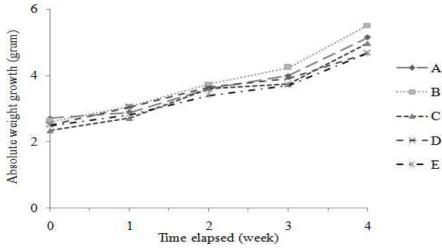


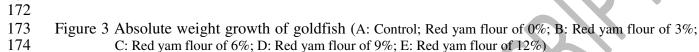
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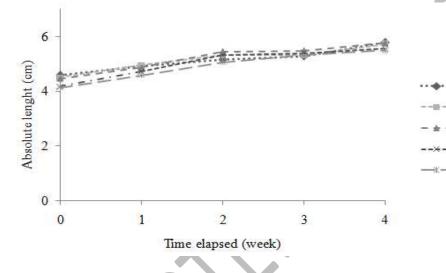
Figure 2 Fish color changes of goldfish (A: Control; Red yam flour of 0%; B: Red yam flour of 3%; C: Red yam flour of 6%; D: Red yam flour of 9%; E: Red yam flour of 12%)

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The growth parameters (absolute weight growth, absolute length growth and specific growth 160 rate) of goldfish increase in each treatment; both control feed and artificial feed with the addition of 161 red yam flour (Figure 3 and 4) but no significant different (P > 0.05). This indicates that fish are able 162 to utilize the feed given for their growth and body maintenance (Putra et al, 2016). Feeding 163 164 increasing the growth of goldfish is considered to meet the nutritional needs of goldfish. It can be 165 seen from the nutritional content in the feed and the resulted performance. According to Subamia et 166 al. (2013), growth occurs when there are excess energies that once; the available energy was used 167 for metabolism, digestion and activities. The result of ANOVA analyses showed that the treatment 168 of red yam flour with different doses did not give significant effect to the growth parameters of 169 goldfish, namely absolute weight growth, absolute length growth and specific growth rate (P> 170 0.05).







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Figure 4 Absolute length growth of goldfish (A: Control; Red yam flour of 0%; B: Red yam flour of 3%; C:
 Red yam flour of 6%; D: Red yam flour of 9%; E: Red yam flour of 12%)

B

-D -E

The survival rates of goldfish during the 28-day study period gave no significant effects in 181 182 each treatment. It is allegedly due to the lack of protein content in red yam flour. The survival of 183 goldfish is also influenced by water quality. According to Noviyanti et al. (2015), the optimal water temperature for the goldfish is around 25 ° C-32 ° C. The temperatures obtained in maintaining 184 media ranged between 27 ° C-29 ° C. The optimal degree of acidity (pH) of water for goldfish is 185 186 around 6-9. The water quality parameters measured during the study were DO, temperature and pH 187 within feasibility limits for the maintenance of goldfish. The over tolerance of water quality of fish may cause fish stress. It is in accordance to Barus et al. (2015), stated that an ornamental fish in a 188 189 state of stress can cause its color to fade. It is due to the gathering of pigment granules in the middle 190 of the cell, so that the fish loses its color.

CONCLUSION
The administration of red yam flour (Ipomoea batatas L.) at different concentration in fish
feed has a significant effect on color improvement with the optimal dose in D treatment (dose of
9%). However, the administration does not significantly affect the increases of absolute weight,
absolute length, daily growth rate and the survival of the goldfish (Carassius auratus). Red yam
flour can be considered as a potential feed supplement to enhance the economical value in
ornamental fish markets.
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