

Anti-Yeast Activity of Cinnamaldehyde, Eugenol and Linalool

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Abstract— It is well known that essential oils and their specific constituents have antimicrobial effects against several pathogenic and saprophytic microorganisms. Therefore, essential oils can be used as alternative or complementary antifungal agents against pathogenic yeasts, especially drug-resistant strains. In addition, essential oils can be used to reduce the effective dose of antifungal drugs. This study evaluates the anti-yeast potential of some essential oil constituents (EOCs), namely, cinnamaldehyde, eugenol and linalool. *Candida albicans*, *Candida glabrata*, *Candida tropicalis* and *Saccharomyces cerevisiae* were used as indicator test strains. EOCs exhibited significant fungicidal activity against indicator strains. The minimum fungicidal concentration (MFC) values of the EOCs ranged from 0.048 to 3.12 µl/ml, while the MFC values of amphotericin B and ketoconazole (positive controls) ranged from 0.78 to 1.56 µg/ml and 6.25–12.25 µg/ml, respectively. These compounds could be further developed into new antifungal agents, either alone or in combination with conventional antifungals.

Index Terms— Anti-yeast activity, Cinnamaldehyde, Eugenol, Linalool.

I. INTRODUCTION

Candidiasis is a fungal infection caused by yeast belonging to the *Candida* genus. More than 20 species of *Candida* can cause infections in humans [1]. Candidiasis ranges from superficial infections to deep invasive infections. Candidiasis is an important health issue not only for immunocompromised patients but also for healthy people [2]. The most common *Candida* species isolated from clinical fungal invasive infections is *Candida albicans*, followed by *C. tropicalis*, *C. parapsilosis* and *C. glabrata* [3,4]. The CDC [5] reported that increasing resistance of *Candida* species to antifungal medications is an emerging public health problem worldwide. Consequently, it is very important to discover alternative antimicrobial compounds. Many studies in the literature claim that plant extracts and essential oils (EOs) obtained from plants are one of the most promising alternative sources of antifungal agents [6-18]. Phenolics, polyphenols, terpenoids, alkaloids, lectins and polypeptides are the major groups of phytochemicals that possess antimicrobial properties [6,7,19].

Because of their natural origin, antimicrobials obtained from plants are also considered to be safer compared to synthetic compounds [19,20].

Additionally, plant-derived antifungals show promise for use against drug-resistant yeasts, because they may have different target sites than traditional antimicrobials and different mechanisms of action [19,21-23]. EO compounds are lipophilic, meaning that they can easily pass through the cell wall and cytoplasmic membrane. They disrupt the structure of the polysaccharide, fatty acid, and phospholipid layers, making the membrane permeable. Consequently, the antimicrobial effects of EOs are linked to their composition and cytotoxic effects, including cell membrane damage [24]. Thus, this study was designed to determine the antifungal activity of the essential oil constituents (EOCs) cinnamaldehyde, eugenol and linalool.

II. EXPERIMENTAL

A. Materials

Stock solutions of chemicals

Cinnamaldehyde (C₆H₅CH=CHCHO, 93+%, natural), eugenol (C₆H₁₂O₂, 98+%, natural) and linalool (C₁₀H₁₈O, 95+%, natural) were purchased from the Sigma-Aldrich Chemical Co. (Germany).

Essential oil compounds and ketoconazole were prepared by dissolving in 20% DMSO (Merck, Germany), while amphotericin B was dissolved in sterile distilled water. All stock solutions were then filter sterilized, and serial two-fold dilutions of the compounds were prepared.

B. Methods

LD₅₀ values of the EOCs

Artemia salina (Brine shrimp) acute toxicity assays were used to determine the cytotoxicity levels of the EOCs [25]. The LD₅₀ was defined as the concentration of the EOCs needed to cause half of the tested brine shrimp to die within 24 h.

Indicator test strains

Candida glabrata (NRRL Y-1418), *Candida tropicalis* (NRRL Y-12968) and *Saccharomyces cerevisiae* (NRRL Y-11878) were obtained from the United States Department of Agriculture Agricultural Research Service (NRRL, Peoria, Illinois, USA). *Candida albicans* (ATCC 60193) was purchased from the American Type Culture Collection (LGC Standards GmbH Mercatorstr. 51 46485 Wesel Germany).

Determination of minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC)

The MIC value was determined using the broth microdilution susceptibility assay according to the NCCLS [26]. First the minimum inhibitory concentration (MIC) and then the minimum lethal/fungicidal concentrations (MFC) of the EOCs (cinnamaldehyde, eugenol and linalool) and the antifungal drugs were determined. The MIC was defined as the lowest concentration of chemical that prevented growth of

the test yeasts. The MFC was defined as the lowest concentration yielding negative subcultures. All tests were performed in duplicate and in parallel.

III. RESULTS AND DISCUSSIONS

A. Acute toxicity values of the EOCs

In this study, an *A. salina* test was used to determine the LD₅₀ values of the EOCs. This *in vivo* acute toxicity test can be used to screen the toxicity of natural and synthetic organic compounds [27] because *A. salina* is highly sensitive to a variety of chemical substances [28]. Additionally, it has been shown that the results of brine shrimp lethality tests correlate with rodent and human acute oral toxicity data [29,30]. The LD₅₀ values of the EOCs ranged from 16.9 to 70.3 µl/ml (Table 1). The assay results were then used to select the EOC dosages to be used in the anti-yeast activity studies.

Table 1. LD₅₀ values of EOCs.

EOCs	LD ₅₀ (µl/ml) values
Cinnamaldehyde	25.6
Eugenol	16.9
Linalool	70.3

B. Anti-yeast potential of the EOCs

Cinnamaldehyde, eugenol and linalool displayed promising fungistatic activity against the yeast strains used in this study (data not shown). In addition, the EOCs had fungicidal activities against all indicator strains tested. The MFCs ranged from 0.048 to 3.12 µl/ml (Figure 1). Fungicidal doses of the EOCs were significantly lower than their LD₅₀ concentrations (between 1/8 to 1/22 of the LD₅₀ concentrations). Eugenol and cinnamaldehyde displayed fungicidal activity at relatively lower doses than linalool. Supporting our results, several studies have reported that cinnamaldehyde [31], eugenol [32] and linalool [33] had significant antifungal activity. Therefore, the EOCs tested in this study have the potential for use in antifungal chemotherapy, either alone or in combination with conventional antifungals. As the MLC values of these EOCs are very low compared to the LD₅₀ values, toxicity may not be a concern; however, more detailed toxicity studies are needed.

C. Anti-yeast activity of antibiotics

Antifungals such as fluconazole, miconazole, itraconazole, nystatin, ketoconazole and amphotericin B are used for the treatment of systemic and superficial fungal infections. In this study, ketoconazole and amphotericin B were used as conventional antibiotics against the test yeasts. All the test yeasts were sensitive to the antifungals; the MFCs of ketoconazole ranged from 6.25 to 12.5 µg/ml, while the MFCs of amphotericin B ranged from 0.78 to 1.56 µg/ml (Figure 2). Additionally, fungicidal concentrations of the antibiotics were significantly higher than those of the EOCs.

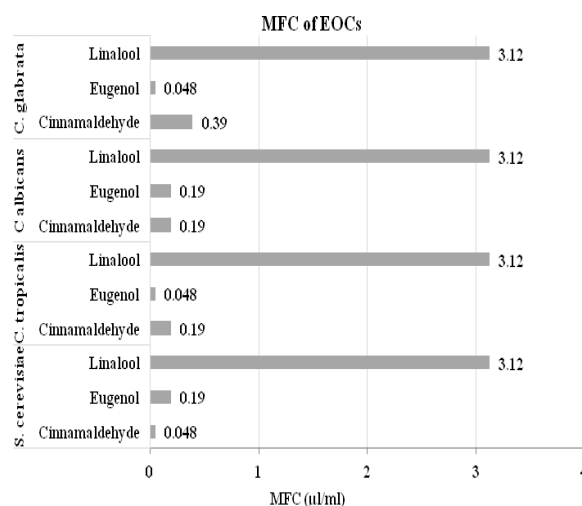


Fig. 1. Minimum fungicidal concentrations (MFCs) of EOCs.

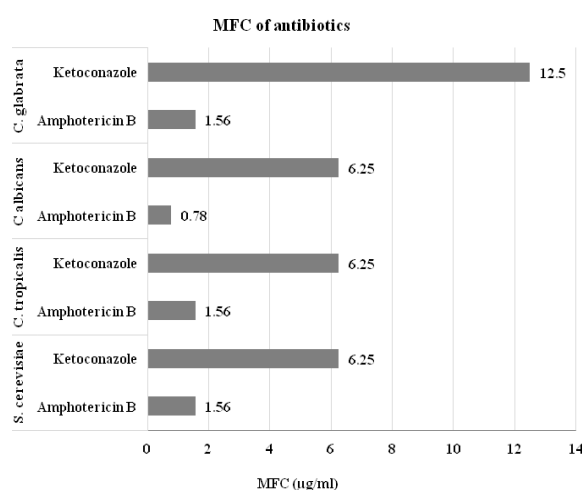


Fig. 2. Minimum fungicidal concentrations (MFCs) of conventional antibiotics.

IV. CONCLUSIONS

The increasing resistance of human pathogens to current antimicrobial agents is a significant problem in modern healthcare. Therefore, there is a real need for the development of new types of antimicrobial substances to treat individuals infected with multidrug-resistant bacteria and fungi. In this study, cinnamaldehyde, eugenol and linalool were evaluated for their antifungal activity. The results showed that cinnamaldehyde, eugenol and linalool exhibited significant fungicidal activity against all indicator test yeasts. Furthermore, these compounds were not toxic at their effective antimicrobial concentrations. In conclusion, these compounds could be used for future development of anti-candidal agents, either alone or combination with conventional antifungal therapeutics. However, the *in vivo* effects of these compounds must be determined to safely use these compounds.

REFERENCES

- [1] CDC, Centers for disease Control, (2017a). <https://www.cdc.gov/fungal/diseases/candidiasis/index.html>
- [2] T.P. McCarty, P.G. Pappas, "Invasive Candidiasis", Infectious Disease Clinics of North America, vol 30(1), pp. 103–124 (2016).

- [3] N. Yapar, H. Pullukcu, V. Avkan-Oguz, S. Sayin-Kutlu, B. Ertugrul, S. Sacar, B. Cetin, O. Kaya, "Evaluation of species distribution and risk factors of candidemia: a multicenter case-control study", *Medical Mycology*, vol 49, pp. 26–31 (2011).
- [4] J. Zirkeel, H. Klinker, A. Kuhn, M. Abele-Horn, D. Tappe, D. Turnwald, H. Einsele, W.J. Heinz, "Epidemiology of *Candida* blood stream infections in patients with hematological malignancies or solid tumors", *Medical Mycology*, vol 50, pp. 50–55 (2012).
- [5] CDC, Centers for disease Control, (2017b). <https://www.cdc.gov/fungal/antifungal-resistance.html>
- [6] A.C. Abreu, A.J. McBain, M. Simões, "Plants as sources of new antimicrobials and resistance-modifying agents", *Natural Product Reports*, vol 29, pp. 1007–1021 (2012).
- [7] M.M. Cowan, "Plant products as antimicrobial agents", *Clinical Microbiology Reviews*, vol 12, pp. 564–582. (1999).
- [8] D. Savoia, "Plant-derived antimicrobial compounds: alternatives to antibiotics", *Future Microbiology*, vol 7(8), pp. 979–990 (2012).
- [9] D. Jasso de Rodríguez, D. Hernández-Castillo, R. Rodríguez-García, J.L. Angulo-Sánchez, "Antifungal activity in vitro of *Aloe vera* pulp and liquid fraction against plant pathogenic fungi", *Industrial Crops and Products*, vol 21, pp. 81–87 (2005).
- [10] D. Jasso de Rodríguez, D. Hernández-Castillo, J.L. Angulo-Sánchez, R. Rodríguez-García, J.A. Villarreal Quintanilla, R.H. Lira-Saldivar, "Antifungal activity in vitro of *Flourensia* spp. extracts on *Alternaria* sp., *Rhizoctonia solani*, and *Fusarium oxysporum*", *Industrial Crops and Products*, vol 25, pp. 111–116 (2007).
- [11] D. Jasso de Rodríguez, F.A. Trejo-González, R. Rodríguez-García, M.L.V. Díaz-Jimenez, A. Sáenz-Galindo, F.D. Hernández-Castillo, J.A. Villarreal-Quintanilla, F.M. Peña-Ramos, "Antifungal activity in vitro of *Rhus muelleri* against *Fusarium oxysporum* f. sp. Lycopersici", *Industrial Crops and Products*, vol 75, pp. 150–158 (2015).
- [12] V.K. Bajpai, S. Shukla, S.C. Kang, "Chemical composition and antifungal activity of essential oil and various extract of *Silene armeria* L.", *Bioresource Technology*, vol 99, pp. 8903–8908 (2008).
- [13] H.T. Chang, Y.H. Cheng, C.L. Wu, S.T. Chang, T.T. Chang, Y.C. Su, "Antifungal activity of essential oil and its constituents from *Calocedrus macrolepis* var. *formosana* Florin leaf against plant pathogenic fungi", *Bioresource Technology*, vol 99, pp. 6266–6270 (2008).
- [14] S. Kordali, A. Cakir, H. Ozer, R. Cakmakci, M. Kesdek, E. Mete, "Antifungal, phytotoxic and insecticidal properties of essential oil isolated from Turkish *Origanum acutidens* and its three components carvacrol, thymol and p-cymene", *Bioresource Technology*, vol 99, pp. 8788–8795 (2008).
- [15] M. Zabka, R. Pavela, L. Slezakova, "Antifungal effect of *Pimenta dioica* essential oil against dangerous pathogenic and toxinogenic fungi", *Industrial Crops and Products*, vol 230, pp. 250–253 (2009).
- [16] L.C. Cordova-Albores, Y.M. Rios, L.L. Barrera-Necha, S. Bautista-Baños, "Chemical compounds of a native *Jatropha curcas* seed oil from Mexico and their antifungal effect on *Fusarium oxysporum* f. sp. *gladioli*", *Industrial Crops and Products*, vol 62, pp. 166–172 (2014).
- [17] N. Khaledi, P. Taheri, S. Tarighi, "Antifungal activity of various essential oils against *Rhizoctonia solani* and *Macrophomina phaseolina* as major bean pathogens" *Journal of Applied Microbiology*, vol 118, pp. 704–717 (2015).
- [18] D. Rongai, P. Pulcini, B. Pesce, F. Milano, "Antifungal activity of somebotanical extracts on *Fusarium oxysporum*". *Open Life Sciences*, vol 10, pp. 409–416 (2015).
- [19] A. Upadhyay, I. Upadhyaya, A. Kollanoor-Johny, K. Venkitanarayanan, "Combating pathogenic microorganisms using plant-derived antimicrobials: A minireview of the mechanistic basis", *Biomed Research International*, vol 2014, 18p., Article ID761741, <http://dx.doi.org/10.1155/2014/761741>
- [20] M.A.B. Rajeh, Z. Zuraini, S. Sasidharan, L.Y. Latha, S. Amutha, "Assessment of *Euphorbia hirta* L. leaf, flower, stem and root extracts for their antibacterial and antifungal activity and brine shrimp lethality" *Molecules*, vol 15, pp. 6008–6018 (2010).
- [21] I. Ahmad, A.Z. Beg, "Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multi-drug resistant human pathogens", *Journal of Ethnopharmacology*, vol 74, pp. 113–123 (2001).
- [22] J.N. Eloff, "Which extractant should be used for the screening and isolation of antimicrobial components from plants?" *Journal of Ethnopharmacology*, vol 60, pp. 1–8 (1998).
- [23] M. Petrosyan, Y. Sherbakovs, N. Sahakyan, Z. Vardanyan, A. Poladyan, Y. Popov, A. Trchounian, "*Alkanna orientalis* (L.) Boiss. plant isolated cultures and antimicrobial activity of their extracts: Phenomenon, dependence on different factors and effects on some membrane-associated properties of bacteria", *Plant Cell Tissue Organ Culture*, vol 122(3), pp. 727–38 (2015).
- [24] F. Bakkali, S. Averbeck, D. Averbeck, M. Idaomar, "Biological effects of essential oils – A review", *Food and Chemical Toxicology*, vol 46, pp. 446–475 (2008).
- [25] J.L. McLaughlin, C.J. Chang, D.L. Smith, "Bench-top bioassays for the discovery of bioactive natural products: An update" In: Atta-Ur-Rahman (Ed.), *Studies in Natural Products Chemistry*. Oxford, Elsevier, pp. 383–409 (1991).
- [26] NCCLS (National Committee for Clinical Laboratory Standards) "Performance Standards for Antimicrobial Susceptibility Testing" Fourteenth Informational Supplement. NCCLS document M100–S14, NCCLS, Wayne, Pa. USA (2004).
- [27] Atta-Ur-Rahman, M.I. Choudhary, W.J. Thomsen, "Bioassay Techniques for Drug Development", Harwood Academic Publishers, CRC Press, 240 p. (2001).
- [28] L.Y. Latha, S. Sasidharan, Z. Zuraini, S. Suryani, L. Shirley, S. Sangetha, "Antibacterial activity and toxicity of *Psophocarpus tetragonolobus*", *Pharmaceutical Biology*, vol 45, pp. 31–36 (2007).
- [29] M.C. Calleja, G. Persoone, "Cyst-based toxicity tests IV, The potential of ecotoxicological tests for the prediction of acute toxicity in man as evaluated on the first ten chemicals of the MEIC programme", *ATLA*, 20, pp. 396–405 (1992).
- [30] A.L. Parra, R.S. Yhebra, I.G. Sardiñas, L.I. Buela, "Comparative study of the assay of *Artemia salina* L. and the estimate of the medium lethal dose (LD50 value) in mice to determine oral acute toxicity of plant extract", *Phytomedicine*, vol 8, pp. 395–400 (2001).
- [31] A.D. Thakre, S.V. Mulange, S.S. Kodgire, G.B. Zore, S.M. Karuppaiyil, "Effects of Cinnamaldehyde, Ocimene, Camphene, Curcumin and Farnesene on *Candida albicans*", *Advances in Microbiology*, vol 6, pp. 627–643 (2016).
- [32] I.C.G. da Silva, H.B. de Pontes Santos, Y.W. Cavalcanti, C.F.W. Nonaka, S.A. de Sousa, R.D. de Castro, "Antifungal activity of eugenol and its association with nystatin on *Candida albicans*", *Brazilian Research in Pediatric Dentistry and Integrated Clinic*, vol 17, e3235. DOI: <http://dx.doi.org/10.4034/PBOCI.2017.171.16> (2017).
- [33] I.J. Dias, E.R.I.S. Trajano, R.D. Castro, G.L.S. Ferreira, H.C.M. Medeiros, D.Q.C. Gomes, "Antifungal activity of linalool in cases of *Candida* spp. isolated from individuals with oral candidiasis", *Brazilian Journal of Biology*, <http://dx.doi.org/10.1590/1519-6984.171054>, (2016).

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