## INFLUENCE OF CULTURE MEDIUM ON THE SPORULATION AND VIABILITY OF Aspergillus spp. AND Talaromyces spp. ENTOMOPATHOGENIC FUNGI

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#### ABSTRACT

Influence of Culture Medium on the Sporulation and Viability of Aspergillus spp. and Talarony pp. Enton pathogenic *Fungi.* The purpose of this study was to determine the effect of three kinds of cultures r luction and spore pro ia on viability of Aspergillus spp. (AS1, 6, 7, 9) and Talaromyces spp. (AS2-5, 8, 10) enter is study was hopathogenic gi. T arranged using Factorial-Completely Randomized Design (CRD) with 2 factors and actor was three s. The plicati kinds of cultures media (potato dextrose agar (PDA), corn meal agar (CMA). dextro e agar (SDA)) and the sab second one was isolates of Aspergillus spp. or Talaromyces spp.. Data of s e viability were tested ore product and using ANOVA and if there was significantly difference, the data then fi 's Honestly Significant nalyzed usi Difference (HSD) test at 5% of significant level. The spore production of Aspergi spp. were in the range of 0.58 - 14.27 x 10<sup>8</sup> spores mL<sup>-1</sup> (PDA);  $0.28 - 2.68 \times 10^8$  spores mL<sup>-1</sup> (SDA) and 1.85<sup>-1</sup> (CMA). The highest spore production 0<sup>8</sup> spores dlaromyces spp. were in the range of was achieved by AS1 isolate that was grown on PDA media. e spor roduced by  $2.15 - 28.62 \times 10^8$  spores mL<sup>-1</sup> (PDA);  $0.28 - 29.43 \times 10^8$  spores mL<sup>-1</sup> and  $1.88 - 16.63 \times 10^8$  spores mL<sup>-1</sup> (CMA). The highest spore production was produced by AS8 isolate which in PDA. The spore viability among isolates of the two cultur ent. The spore entomopathogenic fungi were not significantly diffe ility of Aspergillus spp. was in the range of 95.10 – 97.66% (PDA), 94.02 – 98.45% (SDA) and 92.86 ore viability of *Talaromyces* spp. was in the range of 0% (A). The 95.83-100% (PDA), 85.83-100% (SDA) Culture medium influenced spore production but not the )% (CMA 0.75of both of the entomopathogenic fungi was PDA media. spore viability. The best culture media u ed for spore pro tio

Key words: Aspergillus spp., culture midium, entomonathogenic fungi, spore production and viability, Talaromyces spp.

## ABSTRAK

Pengaruh Medium terhadap ulasi da Viabilitas Jamur Entomopatogen Aspergillus spp. dan Talaromyces spp. Penelitian ngetahui pengaruh tiga jenis media tumbuh terhadap produksi dan viabilitas spora jamur Aspergillus ini bertujuan un comyces spp. (AS2-5, 8, 10). Penelitian ini disusun menggunakan Rancangan Acak Lengkap spp. (AS1, 6, 7 an 2 falkor dan 3 ulangan. Faktor pertama adalah tiga jenis media tumbuh (*potato dextrose agar* (PDA), Faktorial (RAL) de [A), dan sabouraud dextrose agar (SDA)) dan faktor kedua adalah isolat jamur Aspergillus spp. atau corn meal agar (CI Talaromyces spp... Daa produksi dan viabilitas spora diuji menggunakan sidik ragam dan jika terdapat beda nyata, diuji lanjut menggunakan uji Beda Nyata Jujur (BNJ) pada taraf nyata 5%. Produksi spora Aspergillus spp. berada dalam kisaran 0,58 -14,27 x 10<sup>8</sup> spora mL-1 (PDA); 0,28 - 2,68 x 10<sup>8</sup> spora mL<sup>-1</sup> (SDA) dan 1,85 - 5,33 x 10<sup>8</sup> spora mL<sup>-1</sup> (CMA). Produksi spora tertinggi dihasilkan oleh isolat AS1 yang ditumbuhkan pada media PDA. Produksi spora *Talaromyces* spp. berada dalam kisaran 2,15 -28,62 x10<sup>8</sup> spora mL<sup>-1</sup> (PDA); 0,28 - 29,43 x10<sup>8</sup> spora mL<sup>-1</sup> (SDA); dan 1.88 - 16,63 x10<sup>8</sup> spora mL<sup>-1</sup> (CMA). Produksi spora tertinggi dihasilkan oleh isolat AS8 yang ditumbuhkan pada media PDA. Viabilitas spora masing masing isolat dari kedua jamur tersebut tidak berbeda nyata. Viabilitas spora Aspergillus spp. berada dalam kisaran 95,10 - 97,66% (PDA), 94,02 -98,45% (SDA) dan 92,86 - 98,20% (CMA). Viabilitas spora Talaromyces spp. berada di kisaran 95,83 - 100% (PDA), 85,83 - 100% (SDA), dan 90,75 - 100% (CMA). Media tumbuh secara nyata mempengaruhi produksi spora tetapi tidak untuk viabilitas spora. Media tumbuh terbaik untuk produksi spora kedua jamur tersebut adalah media PDA.

Kata kunci: Aspergillus spp., jamur entomopatogen, media tumbuh, produksi dan viabilitas spora, Talaromyces spp.

Entomopathogenic fungi is one of the promising biocontrol agents for controlling plant pest insects (Shah & Pell, 2003; Khan et al., 2012). Some species of entomopathogen such as Beauveria bassiana (Balsamo), Vuillemin (Zimmermann, 2007a) and Metarhizium anisopliae (Metschinikoft) (Zimmermann, 2007b) have been proven as excellent biological control agents against large number of pest insects. Recently, a group of fungi belongs to the genus Aspergillus have also been reported as entomopathogen (Devi et al., 2017). This fungi had been reported to cause mortality many kinds of pest insects (Shubha et al., 2014; Yang et al., 2015; Hamdani et al., 2011; Pasaru et al., 2014; Bordoloi et al., 2012). Another group of fungi, namely Talaromyces, have also been proven as potential biological control agent. The fungus showed good capability to inhibit growth of some plant pathogens, such as Verticillium dahliae (Bahramian et al., 2016), Fusarium oxysporum (Barahmian et al., 2016), Gaeumannomyces graminis var tritici (Ghanbari & Mohammadi, 2015), Sclerotinia sclerotiorum (McLaren et al., 1994) and nematodes such as Pratylenchus oryzae (Kisaakye, 2014). In our pre-tous study, we found fungi from the genus of Aspergalus and Talaromyces which had capability to cause cocoa mirid bugs (Helopeltis spp.).

For field applications, large-g ale propagatio entomopathogenic fungi is a nece sary step that mu be performed. Before mass prod tion, selection of ter of growing media (aga medium or sta entomopathogenic fun i is one of the most important eral types of culture medium things to carry out. So commonly used for en opathogenic fungi, such as

sabouraud dextrose agar (SDA), potato dextrose agar (PDA), and corn meal agar (CMA) (Ingle, 2014; Senthamizhlselvan et al., 2010; Ali et al., 2016). Those three above mentioned media has different ingredients that may give different effect to different entomopathogenic fungi. Many reports stated that different entomopathogenic fungi will not perform similar growth in the same medium (El Damir, 2006; Pandey & Kanaujia, 2006; Francisco et al., 2006, Hase & Nasreen, 2017). This study was performed to investigate effect of those three mentioned media (PDA, SDA and CMA) on the growin, spore duction and spore viability of Aspergilla pp. and *Tal* omyces spp. sult of th s study will entomopathogenic fu The give information or the suitable d are medium that can be used for *e*  $As_{I}$ gillus spp. or vati of Talaromyc

## ATERIAL AND METHOD

**Stue site.** This tudy was performed from March to June 1017. Fungal propagation and observation on their ore production and spore viability were conducted in the aboratory of Agricultural Biotechnology, Faculty of Agriculture, University of Lampung.

**Fungal isolates.** Four isolates of *Aspergillus* spp. and six isolates of *Talaromyces* spp. entomopathogenic fungi were used in this study. The isolates were obtained from three kinds of plant rhizosphere, namely corn, pineapple and chili. They were morphologically and molecularly identified using ITS1 and ITS4 (Unpublished data) (Table 1).

Table 1. Isolate, of entomopathogenic fungi used in this study

Isolate	Isolated from (rhizosphere)	Origin	Year Isolated	Identity
AS 1	Pineapple	Lampung Tengah	2015	Aspergillus sp.
AS 2	Pineapple	Lampung Tengah	2015	Talaromyces sp.
AS 3	Pineapple	Lampung Tengah	2015	Talaromyces sp.
AS 4	Pineapple	Lampung Tengah	2015	Talaromyces sp.
AS 5	Pineapple	Lampung Tengah	2015	Talaromyces sp.
AS 6	Corn	Lampung Selatan	2016	Aspergillus sp.
AS 7	Corn	Lampung Selatan	2016	Aspergillus sp.
AS 8	Corn	Pesawaran	2016	Talaromyces sp.
AS 9	Corn	Pesawaran	2016	Aspergillus sp.
AS 10	Chili	Unknown	2017	Talaromyces sp.

**Culture medium.** All the isolates were grown in sterile plastic petri dishes (90-mm diameter) contains three kinds of media i.e potato dextrose agar (PDA; HIMEDIA® India), sabouraud dextrose agar (SDA; HIMEDIA® India) and corn meal agar (CMA; HIMEDIA® India). The inoculated petri dishes were incubated at  $27 \pm 1^{\circ}$ C for 7 days.

**Culture preparation.** Mycelial plugs (4-mm diameter) of each isolates were excised from the margins of colonies (2-days-old cultures that were incubated at 27  $\pm$  1 °C) and placed in the center of a sterile plastic petri dish (90-mm diameter) containing 30 mL of media (PDA, SDA or CMA). Three replicates were prepared for each treatment.

**Preparation of the spore suspension.** Spore suspension was prepared using sterile 0.1% Tween 80. As much as 10 mL of sterile 0.1% Tween 80 was added into sterile plastic petri dish containing 7-days-old entomopathogenic fungal isolates and scraping the mycelium from plate cultures carefully. The suspension then filtered using sterile filter funnel (0.2-mm of mesh size) to remove mycelia and placed into sterile erlenmeyer flask (50 mL of volume).

Estimating spore production. One mL o or suspension was placed into haemocytomet The re production was counted using a haen beytometer und binocular microscope (Leica, Switz rland) with 400x of magnification. Observation was rformed on the presence of individual a sore on 5 are gri ls of haemocytometer (mediur size). Data of spore produced by each isolates of entom athogenic fungi was average of individual spore observe n 5 square grids. Total spore production wa palyzed us. mula described by Syahnen et al. 2011. follows;  $S = R \times K \times F$ ; S =a conjunta ( $2.5 \times 10^5$ ), F = dilution spore production, factor used.

**Spore germination.** Spore suspension  $(25 \ \mu\text{L}; 1 \times 10^6 \text{ spore mL}^{-1})$  from each isolates were placed individually (3 inoculation point) into a sterile plastic petri dish (9-mm diameter) containing 30 mL each of agar media. Each inoculation point was covered with a sterile glass coverslip (18 mm × 18 mm). The dishes were incubated at  $27 \pm 1$  °C for 10 h. Total spore geminated were calculated under binocular microscope (Leica, *Switzerland*) with 400x magnification. Spores determined to be germinated if the length of the fur is 2x length of conidia diameter (Espinel-Ingroff, 2000).

**Experimental design and statistical analysis.** This study was arranged using Factorial-Completely Randomized Design (CRD) with 2 factors and 3 replications. The first factor was three kinds of cultures media, namely PDA, CMA and SDA. The second factor was isolates of *Aspergillus* spp. or *Talaromyces* spp.. Data transformations was carried out to create nearly equal spreads or additive relationship of the data. Spore production and spore viability data were analyzed using ANOVA and if there was significantly different, the data was further investigated using Tukey's Honestly Significant Difference (HSD) test at 5% of significant level.

# RESULTS AND SCUSSIO

Four isolates *cl* Aspergillus six isolates p. ané pathoge fungi (Figure of Talaromyces sp entor heir spore production and spore 1) were investigated o tivated on ee kinds of cultures media, viability *y* DA), sabouraud dextrose lextrose aga namely potal orn meal agar (CMA). Those isolates agar (SDA) and now the capal tw to infect and cause mortality of irid bugs (*Helopeltis* spp.) (Unpublished data). coa

**Spon production.** Spore production among *Talaro yces* spp. isolates and *Aspergillus* spp. isolates was significantly different. The spore generated by both *spergillus* spp. and *Talaromyces* spp. was significantly influenced by the cultures media used as well as the isolates (Table 2).

Aspergillus spp.. Spore production of each isolate was in the range of  $0.58 - 14.27 \times 10^8$  spores mL<sup>-1</sup> (PDA);  $0.28 - 2.68 \times 10^8$  spores mL<sup>-1</sup> (SDA) and  $1.85 - 5.33 \times 10^8$  spores mL<sup>-1</sup> (CMA). The highest spore production was achieved by AS1 isolate grown on PDA media, and it was significantly higher than AS1 isolate on SDA and CMA or all isolates which were cultured on PDA, SDA and CMA. The lowest spore production was obtained by AS6 isolate on SDA media and it was statistically not different from AS7 isolate that was cultured on PDA media also AS 1 and AS9 isolates grown on SDA media (Table 3).

Based on the data of average of spore production by all isolates in the three cultures medium, the highest spore production was obtained by isolates that were grown on PDA media ( $8.78 \times 10^8$  spore mL<sup>-1</sup>) followed by CMA ( $3.28 \times 10^8$  spore mL<sup>-1</sup>) and SDA media ( $1.23 \times 10^8$  spores mL<sup>-1</sup>) (Figure 2).

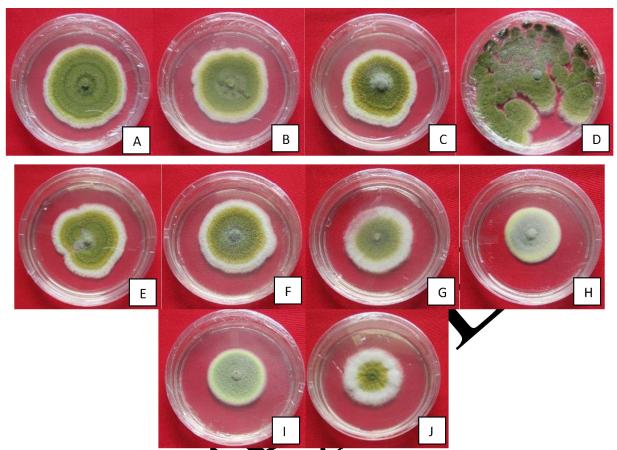


Figure 1. Isolates of *Aspergillus* spp (in Lisolate: A–D) and *Talaromyces* spp. (n=6 isolates; E–J) used in this study. A. AS1, B. AS6, G. AS7, D. AS9, AS2, F, AS3, G. AS4, H. AS5, I. AS8, J. AS10

Table 2. Analysis of variance of sport and the sport of t

Source	df	Anova ss	Mean square	Fvalue	Pvalue	
		Aspergillu	s spp.			
Cultures me	2	15.12	7.56	203.78	<.0001	
Isolates	3	2.20	0.73	19.78	<.0001	
Cultures media * 1solat	6	14.01	2.34	62.94	<.0001	
		Talaromyc	es spp.			
Cultures media	2	8.44	4.22	163.82	<.0001	
Isolates	5	72.15	14.43	560.22	<.0001	
Cultures media * isolates	s 10	23.92	2.39	92.86	<.0001	

*Talaromyces* spp.. Spore produced by all *Talaromyces* spp. isolates were in the range of  $2.15 - 28.62 \times 10^8$  spores mL<sup>-1</sup> (PDA);  $0.28 - 29.43 \times 10^8$  spores mL<sup>-1</sup> (SDA); and  $1.88 - 16.63 \times 10^8$  spores mL<sup>-1</sup> (CMA). The highest spore production was produced by AS8 isolate which were cultured on PDA and it was not significantly different with AS8 isolate grown in SDA and CMA media. This AS8 isolate was significantly different than

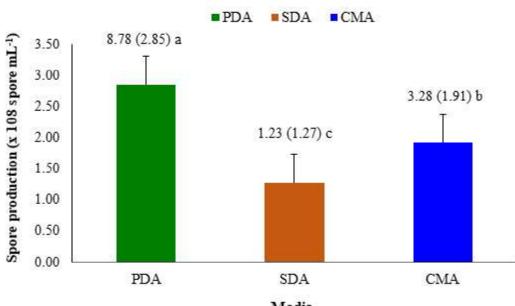
the other isolates grown in the three cultures media. The lowest spore production was produced by AS4 cultured in SDA and it was not significantly different with AS2, AS3 and AS 10 grown in SDA (Table 4).

Based on the average of the spore production of all the isolates that were cultured in the three kinds of medium showed that the isolates which were grown in PDA media produced the highest spore  $(10.34 \times 10^8)$ 

Media	Isolates	Spore production (x 10 <sup>8</sup> spore/mL <sup>-1</sup> )
	AS 1	14.27 (3.84) a
PDA	AS 6	10.20 (3.26) b
PDA	AS 7	0.58 (1.04) fg
	AS 9	10.05 (3.24) b
	AS 1	1.12 (1.27) efg
	AS 6	0.28 (0.88) g
SDA	AS 7	2.68 (1.78) de
	AS 9	0.83 (1.15) fg
	AS 1	1.85 (1.53) def
CMA	AS 6	5.33 (2.38) c
CMA	AS 7	3.33 (1.96) cd
	AS 9	2.58 (1.75) de
Pvalue		<.0001
HSD 5%		0.57

Table 3. Spore production of Aspergillus spp. in three kinds of different cultures media

Number in one column followed by the same letter (s) was not significantly different based of Tukey's Honestly Significant Difference (HSD) test at 5% of significant level. Number in parentheses were the routh of transformation  $\sqrt{x+0.5}$ .



Media

Figure 2. Average of spore production of *Aspergillus* spp. cultured in three kinds of different cultures media. The best spore production was obtained by isolates grown on PDA media, followed by CMA media and SDA media. Number in parentheses are the result of transformation  $\sqrt{x+0.5}$  of spore production data. Number with the same letter was not significantly different based on Tukey's Honestly Significant Difference (HSD) test at 5% of significant level

spores mL<sup>-1</sup>), followed by CMA (6.81 x10<sup>8</sup> spores mL<sup>-1</sup>) and SDA media (6.52 x10<sup>8</sup> spores mL<sup>-1</sup>) (Figure 3). SDA media has been reported as the best media for *Aspergillus* spp. followed by PDA media and the last was CMA media (Ali et al., 2016). Ingle (2014) noted

that SDA media can provide colony growth and sporulation of entomopathogenic fungi *Nomuraea rileyi* better than PDA media. Senthamizhlselvan et al. (2010) revealed *B. bassiana* isolates BbMdKKL 2106 had maximum sporulation on SDA media (8.95 x 10<sup>8</sup> spore

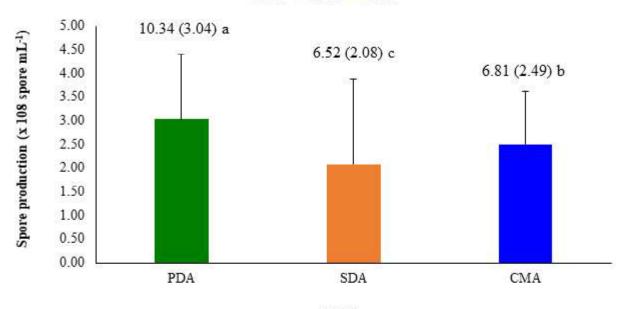
Media	Isolate	Spore production (x $10^8$ spore mL <sup>-1</sup> )
	AS 2	13.67 (3.76) bc
	AS 3	6.83 (2.71) d
PDA	AS 4	2.15 (1.63) fg
PDA	AS 5	7.43 (2.81) d
	AS 8	28.62 (5.40) a
	AS 10	3.33 (1.95) ef
	AS 2	0.92 (1.18) ghi
	AS 3	1.00 (1.22) ghi
	AS 4	0.28 (0.88) i
SDA	AS 5	7.10 (2.75) d
	AS 8	29.43 (5.47) a
	AS 10	0.40(0.95) hi
	AS 2	3.17 (1.91)ef
	AS 3	5.05 (2.35) de
	AS 4	1.50 (1.41) sn
СМА	AS 5	1.88(1.5) fg
	AS 8	16.63 4.14
	AS 10	<u>12.60</u> (3.62) c
Pvalue		001
HSD 5%		

Table 4. Spore production of Talaromyces spp. in three different cultures media

Number in one column followed by the same letter (s) was not significantly different based on Tukey's Honestly Significant Difference (HSD) test at 5% of significant ovel. Number in parentheses were the result of transformation

√x+0.5

■PDA ■SDA ■CMA



Media

Figure 3. Average of spore production of *Talaromyces* spp. cultured in three different cultures media. The best spore production was obtained by isolates grown on PDA media, followed by CMA media and SDA media. Number in parentheses are the result of transformation √x+0.5 of spore production data. Number with the same letter was not significantly different based on Tukey's Honestly Significant Difference (HSD) test at 5% of significant level.

mL<sup>-1</sup>) while FpCmKKL 1526 isolate had minimum sporulation on PDA media (0.28 x 10<sup>8</sup> spore mL<sup>-1</sup>).

In this study, the isolates of *Aspergillus* spp. and *Talaromyces* spp. which were grown on PDA media were able to produce the highest spores followed by CMA and SDA (Figure 2; 3). It is suggested that PDA media is more suitable to optimize the spore production of *Aspergillus* spp. and *Talaromyces* spp. compared with SDA and CMA.

In line with this study, some reports also stated that fungal isolates grown on PDA media have improved their ability and showed better spore production than other media. Gupta et al. (2012) described that *Aspergillus niger* grown on PDA media showed better growth compared to CYA (Czapek's Dox + Yeast Extract Agar) and LCA (Lignocellulose Agar). PDA media are also reported to be able to produce better growth and sporulation of *Rhizoctonia solani*, *Uromyces appendiculatus*, *Cercospora beticola*, Alternaria alternata, Alternaria helianthi and Aspergillus fumigatus than Czapek's agar (CZA) media, CMA (corn meal agar), NA (nutrient agar) and SDA (sabouraud dextrose agar) (Hase & Nasreen, 2017).

The results of this study indicated that culture medium influenced spore production of *Aspergillus* spp. and *Talaromyces* spp.. All solates of both entomopathogenic fungi grown in three kinds of medium (PDA, SDA and CMA) produced spores which were significantly different (Tabel 2, 3, 4; Figure 2, 3).

**Spore viability.** Spore viability among all *Talaromyces* spp., as well as Aspergillus spr, was no gnificantly different. Each of the isolat produced ores with similar viability. The spece viab of both spergillus spp. and *Talaromyc* nfluenced by the was n spp. cultures media or isolat (Table

Source	df	Anova ss	M yn sgaare	Fvalue	Pvalue	
		Aspergilas	spp.			
Cultures media	2	0.002	0.001	0.02	0.98	
Isolates	3	0.14	0.05	0.91	0.45	
Cultures media * isolates	6		0.02	0.43	0.85	
		Talaromyc	spp.			
Cultures media	2	0.49	0.24	3.07	0.06	
Isolates	<b>)</b> 5	0.69	0.14	1.74	0.15	
Cultures media * isolates	-10	0.84	0.08	1.06	0.42	

Table 5. Analysis of variance of spore viability of *Aspergiller* pp. and *Naromyces* spp.

## Table 6. Spore viability of spergilly spp. in three different cultures media

Media	Isolates	Spore viability (%)	
	AS 1	95.10	
	AS 6	95.30	
PDA	AS 7	97.66	
	AS 9	97.14	
	AS 1	94.02	
	AS 6	96.67	
SDA	AS 7	98.45	
	AS 9	95.24	
	AS 1	96.58	
	AS 6	98.20	
CMA	AS 7	97.88	
	AS 9	92.86	
Pvalue		0.89 <sup>nd)</sup>	
HSD 5%		0.29	

<sup>nd)</sup> Not significantly different

Aspergillus spp.. The spore viability of Aspergillus spp. was in the range of 95.10 - 97.66% (PDA), 94.02 - 98.45% (SDA) and 92.86 - 98.20% (CMA) (Table 6). The highest average of spore viability produced by the isolates grown on CMA media (96.38%), PDA Smedia (96.30%) and the lowest on SDA media (96.10%) (Figure 4). *Talaromyces* spp.. *Talaromyces* spp. showed spore viability in the range of 95.83 - 100% (PDA), 85.83 - 100% (SDA), and 90.75 - 100% (CMA) (Table 7). Based on the average of spore viability, the highest viability was obtained by isolate grown in PDA media (98.07%) followed by CMA (96.71%) and the lowest was SDA (93.74%) (Figure 5).

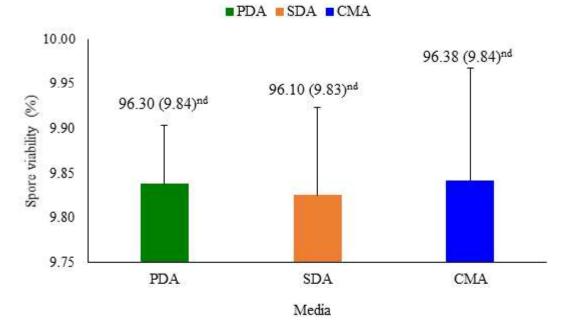
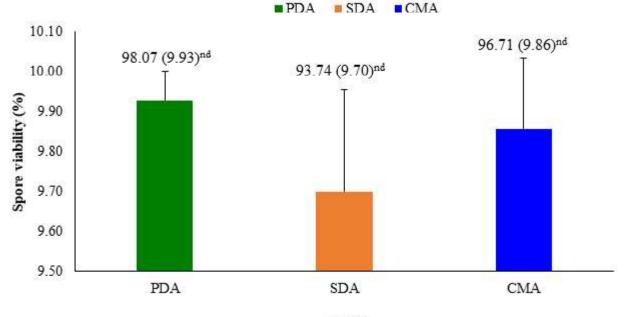


Figure 4. Spore viability of *Aspergillus* spp. cultured in three different cultures media. Number in parentheses are the result of transformation  $\sqrt{x+0.5}$  of spore viability data. Spore viability of the isolates grown in the three kinds of different cultures media was not significantly different (nd) based on Tukey's Honestly Significant I ofference (HSD) test at 5% of significant level

Table 7. Sport v. ility of Talaromyces spp. in three different cultures media

Media	Isolates	Spore viability (%)
	AS 1	95.10
	AS 6	95.30
<b>P</b> DA	AS 7	97.66
	AS 9	97.14
	AS 1	94.02
	AS 6	96.67
SDA	AS 7	98.45
	AS 9	95.24
	AS 1	96.58
	AS 6	98.20
CMA	AS 7	97.88
	AS 9	92.86
Pvalue		0.15 <sup>nd)</sup>
HSD 5%		0.40

<sup>nd)</sup> Not significantly different



Media

Figure 5. Spore viability of *Talaromyces* spp. cultured in three different cultures media. Number in parentheses are the result of transformation  $\sqrt{x+0.5}$  of spore viable y data. Speceviability of the isolates grown in the three kinds of different cultures media was no significantly different (nd) based on Tukey's Honestly Significant Difference (HSD) test at 5% of significant level.

The fact that spore viability is not affected by type of media was also reported in Vrticillium lecanit Derakhshan et al. (2008) explained the V. lecanii grown on MYB (molasses yeast broth) me PCB (p tato carrot broth), JYB (jagge y yeast broth). sucrose yeast broth), PSB (pot o sucrose broth) and PDB (potato dextrose broth) similar spore a relative viability, rangi from 89 to



Culture medium (PDA, SDA and CMA) influenced spore production of *Aspergillus* spp. and *Talaromyces* spp.. However, the medium did not affect the viability of spore produced by those two entomopathogenic fungi. Spore viability produced among all the isolate of the two entomopathogenic fungi were not significantly different. PDA was the best culture media used for spore production for both entomopathogenic fungi.

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