

Squalene Extraction: Biological Sources and Extraction Methods.

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Abstract—Squalene is a terpenoid with great importance in cosmetic, food and pharmaceutical industry; it was originally isolated from shark liver oil but is easily found in animals, vegetables and microorganisms. Nowadays shark fishing is prohibited in some countries, that is the main reason to use renewable sources for squalene extraction to protect marine life, since last decade, squalene is extracted from different sources and methods to achieve best yields at lower possible cost. Traditional extraction methods usually involve organic solvents as hexane which left residues on the extracted matrix, that can limit material use for human consumption after extraction. Separation and purification stages after extraction can elevate operations cost, one of the most interesting technology to obtain squalene from biological matrix is supercritical fluid extraction with CO₂ as solvent because of economic, safe and easy removal characteristics.

Keywords—Extraction, Renewable sources, Squalene, scale-up.

I. INTRODUCTION

Squalene is a very valuable compound common to found in vegetables and animal cells, because of its intermediate on phytoosterols and cholesterol biochemical pathways and highly appreciated by its biological importance¹. Squalene market is mainly divided into three industry sectors, cosmetics (69.2%), food (22.8%) and pharmaceutical (8%) (Fig. 1A) during 2014, squalene demand was about 267 000 ton that represents 102.4 billion dollars. Europe is the main squalene consumer followed by Asia Pacific and North America (Fig. 1B)². Several investigations have been done to search new sources of squalene by different extraction methods to achieve greatest yield at lower possible cost. The aim of this work is to gather information about common and uncommon available animal, vegetal and microbial sources to extract squalene and methods or techniques to extract it as well and scaling-up experiments.

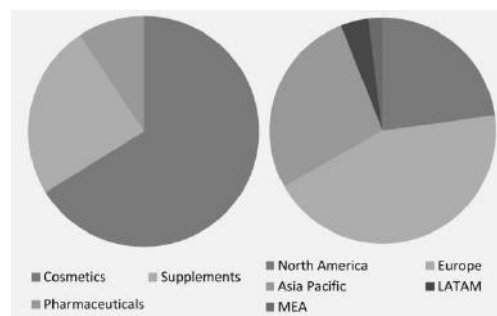


Fig.1: Market by industrial sector (A) and geographical area (B)².

II. SQUALENE

In 1916 by Matsumaru Tsujimoto³, identified a highly unsaturated hydrocarbon was identified from liver oils of the squaloid sharks, he proposes the name 'Squalene'. Squalene, is an hydrocarbonated chain (C₃₀H₅₀), a triterpene containing six unsaturated bonds with antioxidant nature⁴. Squalene has applications in various end-user industries such as cosmetic, food supplements, pharmaceuticals, and in other applications like high grade lubrication and fiber coating additives, however, the major data of commercial is referred to Shark Liver Oil (SLO). In USA SLO was used for vitamin A production but now is highly recommended in alternative medicine and ointment⁵. In Europe, the cosmetic industry demanded SLO, as mentioned before, since product as lotions, eyeliner, eyeshadows, eye makeup remover and perfumes contains 0.1-10% squalene and foundation, lipsticks and other faces preparations contains up to 50% squalene⁶, pharmaceutical, textile and leather industry also demands squalene. In Africa SLO is mainly use on fishing boats maintenance⁷.

2.1 Squalene importance

Squalene is a molecule with a long carbon chain it tends to have hydrophobic properties that is of particular interest in industry because it can be used to transport liposoluble compounds in an effective and economic ways.⁸ Squalene participates in the formation of steroid hormones, bile acids, steroids, and sterols synthesized through mevalonic acid pathway⁹.

Human epidermal sebum is composed by triacylglycerides, free fatty acids (57%), wax esters (26%)

and squalene (12%), the use of compounds present in human sebum as squalene in cosmetics reduces the possibility of allergies¹⁰ and is highly appreciated in cosmetic industry due its emollient and antioxidant properties¹. Squalene prevents H₂O₂ induced oxidative injury and protect against oxidative DNA damage¹¹. Alcohol produces lipid peroxidation although, squalene reduces fetus retina during pregnancy¹² Squalene reduces serum cholesterol due this triterpenoid may act as a substrate for HMG-CoA reductase (3-hydroxy-3-methylglutaryl Co-A)¹³. Squalene has been studied along the years and has been reported with biological activity as antioxidant^{11,14,15} chemopreventive¹⁶. The use of squalene in combination with antitumor drugs has been shown to decrease cancer cells growth^{17,18}.

2.2 Squalene biosynthesis

Squalene is found in both mammals and vegetable tissues because is an intermediary in cholesterol and sterols pathway, very important to any organism. Squalene biosynthesis initiates with enzyme thiolase that joins 2 units of Acetyl-CoA to form Acetoacetyl-CoA and by addition of another Acetyl-CoA by HMG-synthase, β -Hydroxy- β -Methylglutaryl-CoA (HMG-CoA) is formed, and by HMG-CoA reductase take place Mevalonate; then Mevalonate-5-phosphotranferse and phosphomevalonate kinase, 2 phosphates from Adenosine Triphosphate (ATP) are added and changes into Dimethylallyl pyrophosphate, Next phenyl-transferase made 2 head-to-tail unions and 3 isoprene units named as Farnesyl pyrophosphate that polymerizes by squalene synthase form squalene realizing inorganic Pyrophosphate (PPi)¹⁹⁻²¹. Those reactions can be observed in Fig. 2

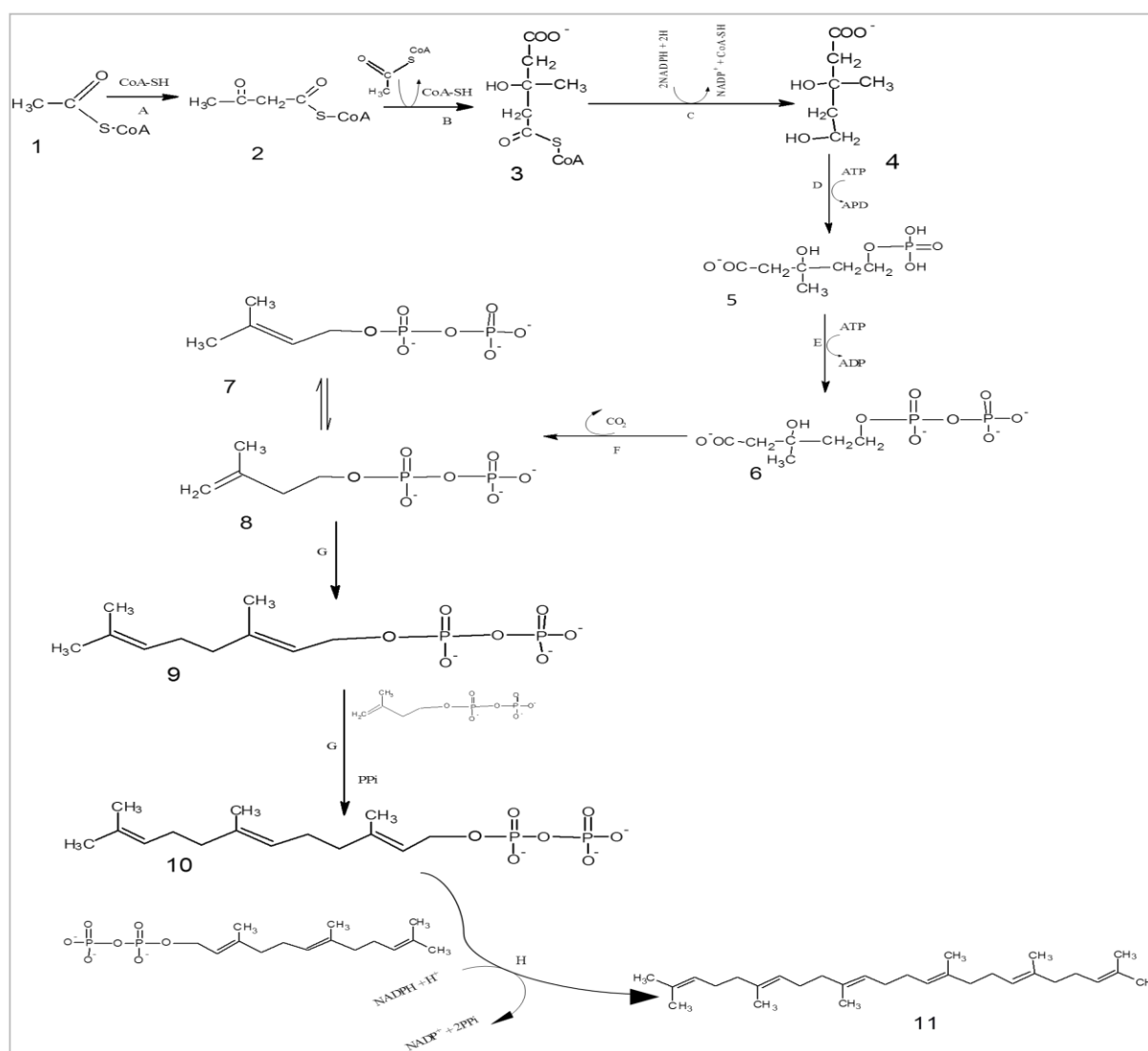


Fig. 2: Squalene synthesis pathway, adapted from reference²²

Intermediates and enzymes names, involved in squalene biosynthesis are listed in Table 1.

Table 1: intermediates and enzymes name involved in squalene biosynthesis

Intermediate	Enzymes
1 Acetyl-CoA	A Thiolase
2 Acetoacetyl-CoA	B HMG-CoA synthase
3 β -hydroxi- β -Methylglutaryl-CoA	C HMG-CoA reductase
4 Mevalonate	D Mevalonate 5-phosphotransferase
5 5-Phosphomevalonate	E phosphomevalonate kinase
6 5-Pyrophosphomevalonate	F Pyrophosphate mevalonate decarboxylase
7 Isopentenyl pyrophosphate	G Prenyl transferase
8 Dimethallyl pyrophosphate	H Squalene sintase
9 Geranyl pyrophosphate	
10 Farnesyl pyrophosphate	
11 Squalene	

III. SQUALENE SOURCES

3.1 Shark Liver Oil

The richest source of squalene is abyssal shark livers even though shallow sharks' livers had lower squalene content than cod livers. New Zealander sharks livers contains about 50% by weight squalene²³. Past decades studies were focused about shark livers and its squalene content. Some of these species are listed in Table 2.

Table 2. Squalene content in different shark liver oil

Shark specie	Squalene liver content (%)	Reference
<i>Centrosymnuscrepidater</i>	35.7-59.4	
<i>Centrosymnusowstoni</i>	37.1-53.1	
<i>Centrosymnuscoelolepis</i>	31.1-47.1	
<i>Deaniacalcea</i>	43.4-66.1	24
<i>Etmopterusbaxteri</i>	14.3-51.5	
<i>Etmopterus sp. nov.</i>	20.8	
<i>Dalatiaslichia</i>	43.4	
<i>Centrophorussquamosus</i>	<0.01	
<i>Centrosymnusplunketi</i>	0.9*	
<i>Etmopterusgranulosus</i>	50.3-60.5*	25
<i>Deaniacalcea</i>	69.6*	
<i>Centrosymnuscrepidater</i>	73*	
New Zealander shark	50-55*	26
<i>Centrophorussquamosus</i>	65.5	27
Cuban sharks	0.03	28

*Expressed as Hydrocarbon (predominantly squalene)

Cuban sharks, squalene determination was performed from a liver mixture of three species *Ginglymostomacirratum*, *Carcharhinuslongimanus* and *Carcharhinusfalciformis*. Nowadays trade volumes of fishing sharks are close to exceed sustainable levels²⁹. Onwards it become necessary to extract squalene from renewable sources³⁰.

3.2 Vegetable Sources

Squalene is present in all vegetable oils but in small amounts³¹. Olive is a well know squalene source and its content depends of it is associated with fruit maturation as autumn begins reach the higher concentration of squalene and the end of season there are no significant changes³². Nowadays olive oil become one of most vegetable squalene source commercially exploited, but its content is not enough to satisfy the demand³³. Deodorized olive oil contains about 28% squalene³⁴. Olive pomace which has been considered like a by-product in the olive oil production has residual (0.0023%) amount of squalene³⁵. In other hand olive leaves that were found containing 0.0038-0.0152% squalene in hexane extracts³⁶. Other products as palm oil contains only 1.8-2.3 % of squalene however, it is produced in huge mounts and so it can be use as squalene source³⁷.

Recently some other crops have been tested as possible new source of squalene. Cucurbit seeds squalene content reported is 10.97-40.27%, differences are due to variety of cucurbit, although is suggested that cucurbit seeds can have hypocholesterolemic effect on human diet³⁸.

Tobacco plant that, contains approximately 2% but; like it continues growing it accumulates up to 20 % in 8 years³⁹. Residues from winery industry (lees) may be also valorized trough squalene recovery, yield achieved was $0.06 \pm 0.008\%$, although seasonal production of raw material, labor requirement may limits its potential as a squalene source⁴⁰.

In Asia, ginseng is important because not need to grow in warm weather, and seeds oil content between 514-569 mg/100g squalene and represents about 60% of unsaponifiable matter.⁴¹ Nuts are an excellent source of vegetable oil; but some of them such as brazil, pecan, pine, pistachio and cashew nuts have a great squalene content due this nuts should be added to dairy diet.⁴² Essential oil obtained by hydrodistillation of the *Strychnos spinosa* leaves contains about 0.5% of squalene in oil fraction.⁴³ Using deodorized soy oil can be extracted 100% squalene content and up to 93% purity by solvent modified extraction⁴⁴. Rice bran is a co-product of milled rice and its contains approximately 20% and about 8.5% of squalene⁴⁵. Bee pollen of lotus (*Nelumbonucifera* Gaertn) content 0.0084% of squalene extracted by supercritical fluid extraction⁴⁶. Some

authors³⁰ have explored unconventional squalene sources as pumpkin, amaranth seeds, borage and walnut reported 0.52, 5.22, 0.022, 2.83, % squalene in oil respectively.

In contrast there are crops that have been underestimated with industrial purpose but in some communities is common to cultivate as cultural heritage and its consumption is local, as amaranth, a pre-Columbian crop that contains relatively high amounts of squalene⁴⁷. *Amaranthus cruentus*, oil extracts reports squalene content 6.95, 5.0 and 8%^{30,48,49}, respectively.

Five varieties of *A. cruentus* were cultivated at different altitudes and reported different squalene content ranged from 2.26 to 5.94% of the oil and statistical analysis showed significant difference for localities but not for varieties of plant so it is suggested that environmental conditions, such as temperature and water availability, may lead to a greater accumulation of squalene in the grain.⁵⁰

Table 3: Squalene vegetable sources

Oil source	Squalene content in oil (%)	Reference
Olive oil deodorized	28	34
Cucurbit seeds	10.97-40.27	38
Olive pomace	0.0023	35
Olive leaves	0.0038-0.0152	36
Tobacco plant	2.00-20.00	39
Wine lees	0.06 ± 0.008	40
Gingseng seed	0.51-0.56	41
Brazilian nut	137.78	
Pecan nut	15.7	
Pistachio	9.14	42
Cashews	8.94	
Pine seed	3.95	
<i>Strychnos spinosa</i>	0.5	43
Deodorized Soy oil	1.83	44
Rice bran	11.75	45
<i>Nelumbonucifera Gaertn</i>	0.0084	46
Palm oil	1.8-2.3	37
Olive oil	0.5-0.65	33
<i>Camellia olifeira</i>	7.62	52
Pumpkin seeds	0.52	
Amaranth seeds	5.22	30
Borage	0.022	
Walnut	2.83	
<i>Amaranthus cruentus</i>	5-8	30,48,49
<i>Amaranthus hybridus</i>	5.27-7.21*	51

*Expressed as unsaponifiable matter

Amaranthus hybridus is reported to have between 5.27±0.47–7.21±0.57% of unsaponifiable matter and can be

assumed that it should contain squalene⁵¹. Table 3 summarizes squalene content in vegetable sources previously mentioned. Some of these renewable sources are not widely harvested or used industrially even when its squalene content is important, and others are widely produced and made them better alternatives than shark livers.

3.3 Microorganism sources

Microorganisms are an interesting squalene source since they do not need to be harvested in huge portions of land. Microalgae (*Schizochytrium mangrovei*) represents a viable alternative source of squalene reaching 33 mg/g of cell dry weight, even when biomass is a residue from biodiesel production⁵³.

A novel yeast strain classified in *Pseudozyma* genus, isolated from seawater is also an interesting squalene source producing 340.52 mg squalene/L with 40 g/L of glucose and sodium nitrogen as nitrogen source⁵⁴.

The strain *Schizochytrium* sp. CCTCC M209059 reports similar squalene content as in fish oil. Due to its fast growing and productivity is an alternative source to obtain squalene. High aeration is recommended to increase squalene synthesis, same authors determined squalene keeps oil stable⁵⁵.

Wild-type *Saccharomyces cerevisiae* can accumulate between 0.62 mg/L of squalene during the stationary growth phase and 3.4 mg/L of squalene until the exponential growth but an engineered strain (named FOH-2) can accumulate more than wild-type strain, since squalene biosynthesis mechanism is overexpressed⁵⁶.

3.4 Squalene Localization

Squalene (and other polyisoprenes) has the function of inhibiting proton leakage through cell membrane, but its localization in cell membrane was not clear until neutron diffraction experiments were performed⁵⁷. Cell membrane is composed by hydrophobic/hydrophilic lipid bilayer, where squalene, a structural analogue of squalene with same number of carbon atoms (C₃₀H₆₂) as squalene, was found to be located between membrane monolayers (Fig. 3). Due to squalene is a saturated compound it may have less stability between the bilayer than unsaturated molecule as squalene⁵⁷. According to this, during squalene extraction, saturated and unsaturated fatty acids could also be extracted.

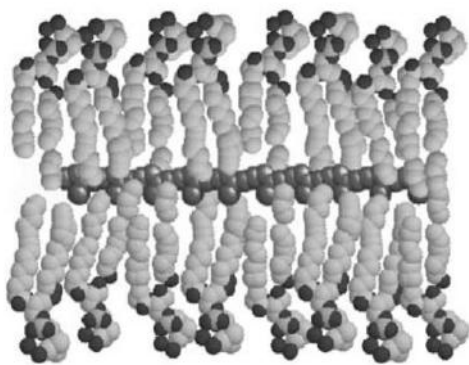


Fig. 3: Schematic representation of squalene in the middle the bilayer of the cell membrane⁵⁷

Evidence of squalene enzymes obtained by immunofluorescence microscopy, suggested that squalene is synthesized in the smooth endoplasmic reticulum subsequently accumulated in small vesicles, some this material is incorporated to plasma membrane⁵⁸. Some squalene vegetables sources as amaranth seeds (*Amaranthushypochondriacus*), lipids fraction have been identified in embryonic cells (Fig 4), surrounding protein bodies (Pb) and cell nucleus (N). A considerable lipid fraction identified with selective colorants as Sudan Black B⁵⁹. is possible to content squalene lipid bodies

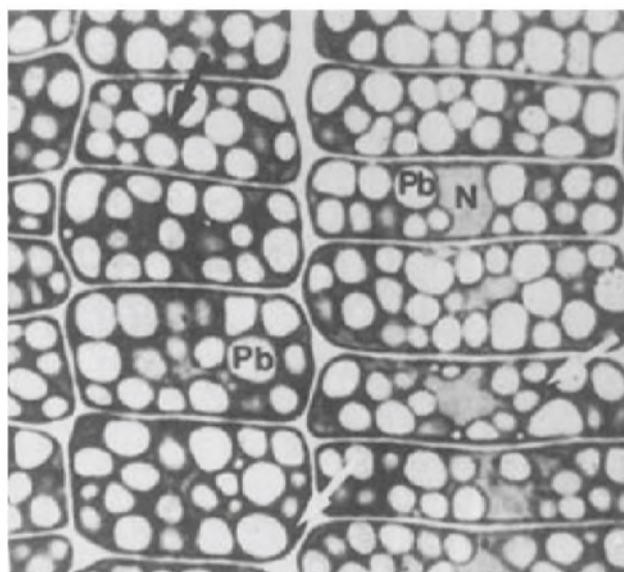


Fig. 4 Light micrograph of amaranth cells peripheric embryo of cytoplasm surrounding the protein bodies is full of lipids (arrows) which stains dark with Sudan Black B⁵⁹

Identifying lipids reserves in vegetables matrix, which probably contains squalene are important to select suitable methods to extract squalene.

IV. EXTRACTION METHODS

Many techniques can be used to recover lipids from biological matrix, and obtain specific

compounds⁶⁰. Soxhlet extraction (organic solvent extraction) is the most common method used as standard and extract is considered to be 100 % of extractable matter⁶¹. Hexane is typically the solvent used for large scale extractions due to its relatively low cost and high extraction efficiency⁶².

Lipid extraction usually involves organic solvents, at industrial scale is commonly used cold press to avoid thermolabile compounds degradation, since this methods are at low pressures, yield might be low, so development of new techniques at higher pressures may aid to increase yield and process time⁶³. Ultrasonic extraction combined with organic extraction can achieve higher yield⁶⁴.

Cold press with new mechanisms that replaces hammer crusher achieved 90.1% extraction and oil reported till 65g/kg of squalene³³. Cold press, organic solvent and Supercritical fluids extraction, were tested in order to compare its yields and the conclusion was supercritical fluids extraction reached the highest yield and purity³¹.

Other separation method, is silver ion complexation based on the complexation reaction between Ag^+ and unsaturated carbon double bonds, it was tested on Camellia oil obtained from seeds of *C. oleifera*, optimal condition was 70% methanol (v/v), 0.6 mol/L AgNO_3 , for 12 h, at 0 °C. Purity of squalene extract reach 37.8%. advantages of this method are low cost, recycle of reagents and continuous operation⁶⁵, an disadvantage of this method, is saponification and esterified before extraction and chemical reagents removed from extract after extraction.

Supercritical fluid extraction (SCFE) can be used to extract polar compounds. Supercritical fluids have diffusivity as gas so can penetrate solid materials, high density and solvation power as liquids, these fluids are compressible and little pressure changes its properties. SCFE have been studied due its advantages against conventional extraction and extract have better quality, biostability and easy to remove from extracted matrix⁶⁶. CO_2 is used to extract oil due its convenience characteristics as non-toxic, non-flammable, easy to remove and economic solvent and also reduces thermolabile degradation in extracted compounds⁴⁸. Squalene SCFE have been performed by several researchers even when is considered as an expensive technology and achieved extract with high purity. Amaranth seeds have been mainly tested by squalene SCE, some conditions are the next: 35MPa and yield was 0.305% and by adding a co-solvent is possible obtain more squalene⁶⁷. In other work, CO_2 were used at 50°C and 300 bar reach 7.95% in oil⁶¹ other optimal conditions reported to extract squalene were 30MPa and 40°C by 90-120 min in order to allow highest and faster oil and squalene extraction from *Amaranthus cruentus*.⁴⁸ At

higher temperature (100°C) best yield is reported to be at 55 MPa and 1.5 h extraction time from *Amaranthus paniculatus*.

V. SCALED-UP EXPERIMENTS

At bench scale, embryonic tissue (as bran from amaranth) was separated from hole seed but amaranth bran was fine thin, therefore pellets were obtained by extrusion to be extracted. Large amounts of pellets (15 Kg) was immersed in hexane for 10 min, solvent was removed at vacuum at 65°C and then filtered⁶⁸ oil recovery reach up to 97.7%.

Scale up studies allows to establish methodology translate SCFE process from laboratory-scale to industrial scale, this behavior is not always approached or predicted, this is the mainly reason to observe differences at studies to avoid serious under or over estimates⁶⁹. Solubility of volatile solutes increases with temperature due an effect in their vapour pressure, this effect is pronounced multicomponent systems than binary systems. Pressure, temperature and solvent density had an effect on the extraction yield, due to the “enhanced solubility effect”. SCFE Laboratory-scale units have a bed length/diameter of vessel ratio relatively high with those greater capacity units, these may affect overall lipid yield, reduced superficial velocity by increasing retention time consequently solvent is saturated⁶¹. Maintaining optimum condition extraction, solvent flow and biomass ratio not affect significantly the process efficiency even at 8 fold scaled-up⁷⁰.

Scaling-up SCFE process depends on extraction efficiency, a model capable to predict the extraction process and time operation which also depends of extracted matrix, batch size, retention time, time for load and unload extraction matrix and cycle of pressuring and depressing extractor to calculate extraction time cycle.⁷¹

VI. CONCLUSION

Squalene is a natural antioxidant very valuable in cosmetic, food and pharmaceutical industries, squalene is also an important intermediate in animal and vegetables cells pathways, accordant to this there are several alternative sources for squalene extraction, many investigations have focused on obtaining best yield as possible, some of the most profitable squalene sources, would be by-products from industrial processes, since squalene is mostly used in human products for human consumption, is important to consider safe extraction methods as supercritical fluid extraction. Scaling-up experiments are important to estimate extraction yield and extract cost. The best source of squalene extraction will depend on the bioproduct and the available technology.

ACKNOWLEDGEMENTS

The authors would like to thank the Instituto Politécnico Nacional and CONACyT for financial support of this work.

REFERENCES

- [1] Huang, Z.-R.; Lin, Y.-K.; Fang, J.-Y. Biological and pharmacological activities of squalene and related compounds: potential uses in cosmetic dermatology. *Molecules* **2009**, *14* (1), 540–554.
- [2] Global Market Insight. Squalene Market Size, Share, Price - Industry Report, 2022 <https://www.gminsights.com/industry-analysis/squalene-market> (accessed May 16, 2017).
- [3] Gnes, F. Medical Use of squalene as a natural antioxidant. *J. Marmara Univ. Inst. Heal. Sci.* **2013**, *3* (4), 1.
- [4] Reddy, L. H.; Couvreur, P. Squalene: A natural triterpene for use in disease management and therapy. *Adv. Drug Deliv. Rev.* **2009**, *61* (15), 1412–1426.
- [5] Rose, D. A.; International Traffic Network.; International Union for Conservation of Nature and Natural Resources. Species Survival Commission. *An Overview of World Trade in Sharks and Other Cartilaginous Fishes*; TRAFFIC International, 1996.
- [6] Kaiya, A. The use of natural squalene and squalane, and the latest situation of the raw materials. *J. Japan Oil Chem. Soc.* **1990**, *39* (8), 525–529.
- [7] Vannuccini, S. *Shark Utilization, Marketing, and Trade*, No. 389.; Food and Agriculture Organization of the United Nations: Rome, Italy, 1999.
- [8] Ball, E. Liposomas En: Dermatología. *Dermatología Venezolana*. **1995**, *33*, 15–23.
- [9] Kim, S.-K.; Karadeniz, F. Biological Importance and Applications of Squalene and Squalane., 1st ed.; Elsevier Inc., **2012**; 65:223-33
- [10] Blasco, L.; Duracher, L.; Forestier, J.; Vian, L.; Marti-Mestres, G. Skin constituents as cosmetic ingredients. part iii: a molecular modeling study of bio-mimetic monoglycerides behavior at the squalene-water interface. *J. Dispers. Sci. Technol.* **2006**, *27* (6), 817–824.
- [11] Warleta, F.; Campos, M.; Allouche, Y.; Sánchez-Quesada, C.; Ruiz-Mora, J.; Beltrán, G.; Gaforio, J. J. Squalene protects against oxidative DNA Damage in MCF10A Human Mammary Epithelial Cells but not in MCF7 and MDA-MB-231 Human Breast Cancer Cells. *Food Chem. Toxicol.* **2010**, *48* (4), 1092–1100.
- [12] Aguilera, Y.; Dorado, M. E.; Prada, F. A.; Marti, J. J.; Quesada, A.; Ruiz-gutie, V. The Protective role of squalene in alcohol damage in the chick embryo

- retina. *Exp. Eye Res.* **2005**, 80, 535–543.
- [13] Strandberg, T. E.; Tilvis, R. S.; Miettinen, T. A. Metabolic variables of cholesterol during squalene feeding in humans: comparison with cholestyramine treatment. *J. Lipid Res.* **1990**, 31 (9), 1637–1643.
- [14] Kohno, Y.; Egawa, Y.; Itoh, S.; Nagaoka, S.; Takahashi, M.; Mukai, K. kinetic study of quenching reaction of singlet oxygen and scavenging reaction of free radical by squalene in N-Butanol. *Biochim. Biophys. Acta* **1995**, 1256, 52–56.
- [15] Aguilara, Y.; Dorado, M. E.; Prada, F. A.; Martí, J. J.; Quesada, A.; Ruiz-gutie, V.; Martínez, J. J.; Quesada, A.; Ruiz-Gutiérrez, V. The protective role of squalene in alcohol damage in the chick embryo retina. *Exp. Eye Res.* **2005**, 80 (4), 535–543.
- [16] Rao, C. V.; Newmark, H. L.; Reddy, B. S. Chemopreventive effect of squalene on colon cancer. *Carcinogenesis* **1998**, 19 (2), 287–290.
- [17] Murakoshi, M.; Nishino, H.; Tokuda, H.; Iwashima, A.; Okuzumi, J.; Kitano, H.; Iwasaki, R. Inhibition by squalene of the tumor-promoting activity of 12-o-tetradecanoylphorbol-13-acetate in mouse-skin carcinogenesis. *Int. J. Cancer* **1992**, 52 (6), 950–952.
- [18] Smith, T.; Yang, G.; Seril, D.; Liao, J.; Kim, S. Inhibition of 4-(Methylnitrosamino)-1-(3-Pyridyl)-1-Butanone-induced lung tumorigenesis by dietary olive oil and squalene. *Carcinogenesis* **1998**.
- [19] Lynen, F. Biosynthetic pathways from acetate to natural products. *Pure Appl. Chem.* **1967**, 14 (1), 137–167.
- [20] Bloch, K. the biological synthesis of cholesterol. *Science* **1965**, 150 (3692), 19–28.
- [21] Gershbein, L. L.; Singh, E. J. Hydrocarbons of dogfish and cod livers and herring oil. *J. Am. Oil Chem. Soc.* **1969**, 46 (10), 554–557.
- [22] Lehninger, A. L.; Nelson, D. L. (David and Cox) *Lehninger Principles of Biochemistry*; W.H. Freeman, Macmillan Learning, 2005.
- [23] Catchpole, O. J.; von Kamp, J.-C.; Grey, J. B. Extraction of squalene from shark liver oil in a packed column using supercritical carbon dioxide. *Ind. Eng. Chem. Res.* **1997**, 36 (10), 4318–4324.
- [24] Deprez, P. P.; Volkman, J. K.; Davenport, S. R. Squalene content and neutral lipid composition of livers from deep-sea sharks caught in tasmanian waters. *Mar. Freshw. Res.* **1990**, 41 (3), 375–387.
- [25] Bakes, M. J.; Nichols, P. D. Lipid, fatty acid and squalene composition of liver oil from six species of deep-sea sharks collected in southern australian waters. *Comp. Biochem. Physiol. - B Biochem. Mol. Biol.* **1995**, 110 (1), 267–275.
- [26] Catchpole, O. J.; von Kamp, J.-C. Phase equilibrium for the extraction of squalene from shark liver oil using supercritical carbon dioxide. *Ind. Eng. Chem. Res.* **1997**, 36 (9), 3762–3768.
- [27] Kjerstad, M.; Fossen, I.; Willemsen, H. M. Utilization of deep-sea sharks at hatton bank in the north atlantic. *J. Northwest Atl. Fish. Sci.* **2003**, 31, 333–338.
- [28] Cruz-Núñez, G.; Palmadóttir, H.; Jónsdóttir, R.; García-Rodríguez, E. Quality of cuban shark liver oil. comparison with icelandic cod liver oil (Calidad del aceite de hígado de tiburón cubano. Comparación con el Aceite de Hígado de Bacalao Islandés). **2009**, 10(2)1-10.
- [29] Clarke, S. C.; McAllister, M. K.; Milner-Gulland, E. J.; Kirkwood, G. P.; Michielsens, C. G. J.; Agnew, D. J.; Pikitch, E. K.; Nakano, H.; Shivji, M. S. Global Estimates of shark catches using trade records from commercial markets. *Ecol. Lett.* **2006**, 9 (10), 1115–1126.
- [30] Czaplicki, S.; Ogrodowska, D.; Derewiaka, D.; Tańska, M.; Zadernowski, R. Bioactive compounds in unsaponifiable fraction of oils from unconventional sources. *Eur. J. Lipid Sci. Technol.* **2011**, 113 (12), 1456–1464.
- [31] Czaplicki, S.; Ogrodowska, D.; Zadernowski, R.; Derewiaka, D. Characteristics of biologically-active substances of amaranth oil obtained by various techniques. *Polish J. Food Nutr. Sci.* **2012**, 62 (4), 235–239.
- [32] Fernández-Cuesta, a.; León, L.; Velasco, L.; De la Rosa, R. Changes in squalene and sterols associated with olive maturation. *Food Res. Int.* **2013**, 54 (2), 1885–1889.
- [33] Samaniego-Sánchez, C.; Quesada-Granados, J. J.; de la Serrana, H. L.-G.; López-Martínez, M. C. β -carotene, squalene and waxes determined by chromatographic method in picual extra virgin olive oil obtained by a new cold extraction system. *J. Food Compos. Anal.* **2010**, 23 (7), 671–676.
- [34] Bondioli, P.; Mariani, C.; Lanzani, A.; Fedeli, E.; Muller, A. Squalene recovery from olive oil deodorizer distillates. *J. Am. Oil Chem. Soc.* **1993**, 70 (8), 763–766.
- [35] Stavroulias, S.; Panayiotou, C. Determination of optimum conditions for the extraction of squalene. *Chem. Biochem. Eng. Q.* **2005**, 19 (4), 373–381.
- [36] Guinda, P. Á.; Lanzón, A.; Albi, J. J. R. T. Aislamiento y cuantificación de los componentes de la hoja del olivo: extracto de hexano. *Grasas y aceites* **2002**, 53 (4), 419–422.
- [37] Al-darmaki, N. I. K. Extraction and enrichment of minor lipid components of palm fatty acid distillate using supercritical carbon dioxide, thesis, University

- of Birmingham, 2012.
- [38] Aguilar, Y. M.; Yero, O. M.; López, J. C.; Navarro, M. V.; Espinosa, M. E. Fitoesteroles yescualeno como hipocolesterolémicos en cinco variedades de semillas de *Cucurbita maxima* y *Cucurbita moschata* (Calabaza). *Rev. Cuba. Plantas Med.***2011**, 16 (1), 72–81.
- [39] Maisashvili, A.; Bryant, H. L.; Richardson, J. W. Economic feasibility of tobacco leaves for biofuel production and high value squalene. International Food and Agribusiness Management Review. *Int. Food Agribus. Manag. Rev.***2016**, 19 (4).
- [40] Naziri, E.; Mantzouridou, F.; Tsimidou, M. Z. Recovery of squalene from wine lees using ultrasound assisted extraction-a feasibility study. *J. Agric. Food Chem.***2012**, 60 (36), 9195–9201.
- [41] Beveridge, T. H. J.; Li, T. S. C.; Drover, J. C. G. Phytosterol content in american ginseng seed oil. *J. Agric. Food Chem.***2002**, 50 (4), 744–750.
- [42] Ryan, E.; Galvin, K.; O'Connor, T. P.; Maguire, a R.; O'Brien, N. M. Fatty acid profile, tocopherol, squalene and phytosterol content of brazil, pecan, pine, pistachio and cashew nuts. *Int. J. Food Sci. Nutr.***2006**, 57 (3–4), 219–228.
- [43] Hoet, S.; Stévigny, C.; Hérent, M.-F.; Quetin-Leclercq, J. Antitrypanosomal compounds from the leaf essential oil of *strychnos spinosa*. *Planta Med.***2006**, 72 (5), 480–482.
- [44] Gunawan, S.; Kasim, N.; Ju, Y. Separation and purification of squalene from soybean oil deodorizer distillate. *Sep. Purif. Technol.***2008**, 60 (2), 128–135.
- [45] Yamamoto, Y.; Hara, S. Novel fractionation method for squalene and phytosterols contained in the deodorization distillate of rice bran oil. *J. Oleo Sci.***2010**, 59 (2), 65–70.
- [46] Xu, X.; Dong, J.; Mu, X.; Sun, L. Supercritical CO₂ extraction of oil, carotenoids, squalene and sterols from lotus (*Nelumbo Nucifera Gaertn*) Bee Pollen. *Food Bioprod. Process.***2011**, 89 (1), 47–52.
- [47] Rodas, B.; Bressani, R. Contenido de aceite, ácidos grasos y escualeno en variedades crudas y procesadas de grano de amaranto. *Arch. lationamericanos Nutr.***2009**, 59 (4), 82–87.
- [48] Wejnerowska, G.; Heinrich, P.; Gaca, J. Separation of squalene and oil from amaranthus seeds by supercritical carbon dioxide. *Sep. Purif. Technol.***2013**, 110, 39–43.
- [49] Singhal, R. S.; Kulkarni, P. R. Effect of puffing on oil characteristics of Amaranth (Rajgeera) Seeds. *gamell***1990**, 380 (13), 952–954.
- [50] Berganza, B. E.; Moran, A. W.; Rodríguez M., G.; Coto, N. M.; Santamaría, M.; Bressani, R.; Coto, M.; Rodr, G.; On, N.; Bressani, R. Effect of variety and location on the total fat, fatty acids and squalene content of amaranth. *Plant Foods Hum. Nutr.***2003**, 58 (3), 1–6.
- [51] Dhellot, R.; Matouba, E.; Maloumbi, M. G.; Nzikou, J. M.; Ngoma, D. G. S.; Linder, M.; Desobry, S.; Parmentier, M. Extraction , chemical composition and nutritional characterization of vegetable oils : case of *Amaranthus Hybridus* (Var 1 and 2) of Congo Brazzaville. *African J. Biotechnol.***2006**, 5, 1095–1101.
- [52] Li, D.; Wang, J.; bi, L.; Zhao, Z. Influence of Extraction method on content of bioactive component squalene in seed oil of *Camellia oleifera* Abel. *Biomass Chem. Eng.***2006**, 40 (1), 9–12.
- [53] Hoang, M. H.; Ha, N. C.; Thom, L. T.; Tam, L. T.; Thi, H.; Anh, L.; Thi, N.; Thu, H.; Ioeng, J. B. I. B. Extraction of squalene as value-added product from the residual biomass of *schizochytrium mangrovei* pq6 during biodiesel producing process. *J. Biosci. Bioeng.***2014**, 118(6), 632–639.
- [54] Chang, M.-H.; Kim, H.-J.; Jahng, K.-Y.; Hong, S.-C. The isolation and characterization of *Pseudozyma* sp. JCC 207, a Novel Producer of Squalene. *Appl Microbiol Biotechnol.* **2008** Apr;78(6):963–72.
- [55] Ren, L. J.; Ji, X. J.; Huang, H.; Qu, L.; Feng, Y.; Tong, Q. Q.; Ouyang, P. K. Development of a stepwise aeration control strategy for efficient docosahexaenoic acid production by *Schizochytrium*Sp. *Appl. Microbiol. Biotechnol.***2010**, 87 (5), 1649–1656.
- [56] Rasool, A.; Ahmed, M. S.; Li, C. Overproduction of squalene synergistically downregulates ethanol production in *Saccharomyces cerevisiae*. *Chem. Eng. Sci.***2016**, 152, 370–380.
- [57] Hauss, T.; Dante, S.; Dencher, N. A.; Haines, T. H. Squalane is in the midplane of the lipid bilayer : implications for its function as a proton permeability barrier. **2002**, 1556(2-3) 149–154.
- [58] Leber, R.; Landl, K.; Zinser, E.; Ahorn, H.; Spo, A.; Kohlwein, S. D.; Turnowsky, F. Dual Localization of squalene epoxidase , Erg1p , in yeast reflects a relationship between the endoplasmic reticulum and lipid particles. **1998**, 9 (February), 375–386.
- [59] Coimbra, S.; Salema, R. *Amaranthus hypochondriacus* Seed structure and localization of seed reserve. *Ann. Bot.***1994**, 74, 373–379.
- [60] Hawthorne, S. B.; Grabanski, C. B.; Martin, E.; Miller, D. J. Comparisons of soxhlet extraction, pressurized liquid extraction, supercritical fluid extraction and subcritical water extraction for environmental solids: recovery, selectivity and effects on sample matrix. *J. Chromatogr. A***2000**, 892 (1), 421–433.

- [61] Westerman, D.; Santos, R. C. D.; Bosley, J. A.; Rogers, J. S.; Al-Duri, B. Extraction of amaranth seed oil by supercritical carbon dioxide. *J. Supercrit. Fluids***2006**, 37 (1), 38–52.
- [62] Mercer, P.; Armenta, R. E. Developments in oil extraction from microalgae. *Eur. J. Lipid Sci. Technol.***2011**, 113 (5), 539–547.
- [63] Vázquez, L.; Torres, C. F.; Fornari, T.; Señoráns, F. J.; Reglero, G. Recovery of squalene from vegetable oil sources using countercurrent supercritical carbon dioxide extraction. *J. Supercrit. Fluids***2007**, 40 (1), 59–66.
- [64] Chung, ki won; Kim, won il; Hong, in kwon; Park, kyung ai. Ultrasonic energy effects on squalene extraction from Amaranth Seed. *Appl. Chem.***2000**, 4, 149–152.
- [65] Xiao, H.; Yao, Z.; Peng, Q.; Ni, F.; Sun, Y.; Zhang, C. X.; Zhong, Z. X. Extraction of squalene from camellia oil by silver ion complexation. *Sep. Purif. Technol.***2016**, 169, 196–201.
- [66] Yin, J.; Wang, A.; Wei, W.; Liu, Y.; Shi, W. Analysis of the operation conditions for supercritical fluid extraction of seed oil. *Sep. Purif. Technol.***2005**, 43 (2), 163–167.
- [67] Kraujalis, P.; Venskutonis, P. R. Supercritical Carbon Dioxide Extraction of squalene and tocopherols from amaranth and assessment of extracts antioxidant activity. *J. Supercrit. Fluids***2013**, 80, 78–85.
- [68] Sun, H.; Wiesenborn, D.; Rayas-Duarte, P.; Mohamed, A.; Hagen, K. Bench-Scale Processing of Amaranth seed for oil. *J. Am. Oil Chem. Soc.***1995**, 72 (12), 1551–1555.
- [69] Prado, J. M.; Dalmolin, I.; Carareto, N. D. D.; Basso, R. C.; Meirelles, A. J. A.; Oliveira, J. V.; Batista, E. A. C.; Angela, M.; Meireles, A. Supercritical Fluid Extraction of grape seed: process scale-up, extract chemical composition and economic evaluation. *J. Food Eng.***2012**, 109, 249–257.
- [70] Taher, H.; Al-Zuhair, S.; Al-Marzouqi, A. H.; Haik, Y.; Farid, M.; Tariq, S. Supercritical Carbon Dioxide Extraction of microalgae lipid: process optimization and laboratory scale-up. *J. Supercrit. Fluids***2014**, 86, 57–66.
- [71] Fiori, L. Supercritical Extraction of Grape Seed Oil at Industrial-Scale: Plant and process design, modeling, economic feasibility. *Chem. Eng. Process. Process Intensif.***2010**, 49 (8), 866–872.