

# Characterization of $\alpha$ -Amylase Produced by the Endophytic Strain of *Penicillium digitatum* in Solid State Fermentation (SSF) and Submerged Fermentation (SmF)

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**Abstract**—  $\alpha$ -Amylases are enzymes responsible for breaking the  $\alpha$ -1.4 bond in polysaccharides with three or more glucose units, occupying the second place in the most widely employed enzymes in industry in the world. The objective of this study was to compare the yields of  $\alpha$ -amylase produced by the endophytic fungus, *Penicillium digitatum*, strain D1-FB, isolated from *Baccharis dracunculifolia* D.C. (Asteraceae), through the solid state fermentation (SSM) and submerged fermentation (SmF) processes, in addition to characterizing the produced enzyme. The two fermentations were conducted for 120 hours, taking samples every 24 hours to obtain the peaks of production. The enzymes were characterized according to their optimal pH and temperature for performance and stability regarding the incubation in the presence of ions, variations in pH and temperature. The maximum yield of the enzyme was observed with SSF, using rice bran as substrate after 72 hours of fermentation, with 1,625 U/mL. The  $\alpha$ -amylase had an optimal pH at 6.5 and optimal temperature at 45°C. All the ions resulted in a decrease in the activity of  $\alpha$ -amylase in the concentration of 5mM. The enzyme proved to be quite stable in a pH range of 6.0 to 7.5 and up to the temperature of 37°C, but it presented great drops in activity with temperatures above 45°C and in the presence of ions at the concentration of 5 mM.

**Keywords**— *Penicillium digitatum*,  $\alpha$ -amylase, starch, enzymes, endophytics.

## I. INTRODUCTION

Starch is a polymer consisting of glucose molecules joined by  $\alpha$ -1.4 and  $\alpha$ -1.6 bonds. Two polysaccharides

comprise the structure of starch: amylose and amylopectin (Figures 1A and 1B). The first (1A) is a linear molecule containing more than 6000 glucose units connected by glycosidic  $\alpha$ -1.4 bonds, and the second (1B), very similar to glycogen (Myers *et al.*, 2000), is highly branched, containing  $\alpha$ -1.4 bonds between the of glucose monomers, and  $\alpha$ -1.6 in the branching points at each 24-30 glucose residue (Brena *et al.*, 1996).

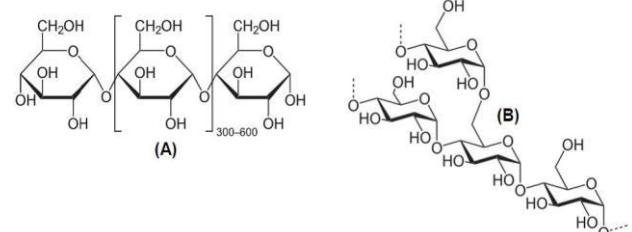


Fig 1: (A) Amylose Molecule. (B) Amylopectin Molecule. Corradini *et al.*, (2005).

Amylases are responsible for the hydrolysis of the starch molecule and are widely distributed in nature. Starch is found mainly in seeds of cereals, such as corn, barley, wheat and rice, and in tubers or roots, such as potatoes and manioc, with the size and shape of the grains being specific for the different cereals (Moraes, 2004). According to Gupta (2003), amylases are divided into two groups, endo-amylases catalyze hydrolyses within the starch molecule in various points of the chain simultaneously. This action causes the formation of dextrans and small polymers composed of glucose units of various lengths, breaking the glycosidic  $\alpha$ -1.4 bonds present on the inside of the amylose or amylopectin chains.  $\alpha$ -Amylase is the most well-known endo-amylase.

The exo-amylases known as amiloglicosidases or glicoamylases, hydrolyze glycosidic  $\alpha$ -1,4 and  $\alpha$ -1,6 bonds (Lorentz, 2005; Onofre *et al.*, 2011). Amylases are applied in the most varied industrial sectors that require the hydrolysis of starch, being mainly used in the food industry for the preparation of beers, jellies and to obtain free glucose for the most varied applications (Michelin, 2005).

In addition to the food industry, these enzymes can be used in the formulation of detergents and in the paper, pharmaceutical, fermentation and textiles industries (Oliveira *et al.*, 2007).

There are two processes for the production of microbial enzymes: solid state fermentation (FES) and submerged fermentation (SmF). SmF is traditionally used for the production of enzymes because it provides a better control of some important process parameters, such as pH and cell growth, in addition to facilitating the recovery of extracellular enzymes and the determination of biomass (Fernandes, 2006).

One of the main characteristics of solid-state fermentation is the use of substrates with low water activity, in which the conditions for growth are similar to the fungi's natural habitat. This facilitates their growth on the solid substrate and the production of large quantities of enzymes (Paris *et al.*, 2008; Rocha, 2010).

The vast majority of microorganisms used in the solid-state fermentation are filamentous fungi (Silva, 2002). The reduced amount of water in the substrate greatly restricts the number of micro-organisms that are capable of adapting to this process, but fungi prove to be quite tolerant to this environment (Pandey *et al.*, 2005).

The objective of this study was to compare the production of  $\alpha$ -amylase produced by the endophytic fungus, *Penicillium digitatum*, strain D1-FB, isolated from *Baccharis dracunculifolia* D.C. (Asteraceae), through the solid state fermentation (SSM) and submerged fermentation (SmF) processes, in addition to characterizing the produced enzyme.

## II. MATERIALS AND METHODS

### 2.1 MICRO-ORGANISM STUDIED

For the realization of this work the endophytic fungus *Penicillium digitatum*, strain D1-FB, was used, isolated from *Baccharis dracunculifolia* D.C. (Asteraceae) maintained in the mycology collection of the Microbiology Laboratory of the Universidade Paranaense - UNIPAR - Campus Francisco Beltrão - Paraná - Brasil.

### 2.2 FERMENTATION PROCESS FOR THE PRODUCTION OF $\alpha$ -AMYLASES

#### 2.2.1 Preparation of the Inoculum

After growing the fungus on PDA for 7 days, the cells were suspended in a phosphate 100mM buffer, pH 7.0, and subjected to stirring in order to obtain a homogeneous solution. The cell concentration was determined by counting in a Neubauer chamber (Germano, 2000).

#### 2.2.2 Preparation of the Fermentation Media

Two different fermentation processes for the production of amylase were tested: solid state fermentation (SSF) and submerged fermentation (SmF). The substrate of choice for SSF was rice bran. Each Erlenmeyer flask of 125 mL received 10 g of rice bran (Fa), with the production of the enzyme in pure bran (Fa) being compared with bran supplemented with 20% of manioc starch (Fm).

The moisture content of the media was guaranteed by a phosphate buffer 100 mM, pH 6.5, in the proportion of 65% for the medium containing pure bran, and of around 90% for the medium supplemented with manioc starch. In SmF, the medium was composed of 50 g.L<sup>-1</sup> of manioc starch, 0.2 g.L<sup>-1</sup> of MgSO<sub>4</sub> and 0.2 g.L<sup>-1</sup> of ZnSO<sub>4</sub>.7H<sub>2</sub>O, solubilized in a phosphate buffer 100 mM, pH 6.5. Two different sources of nitrogen were tested, one organic (urea) and the other inorganic (NaNO<sub>2</sub>) at 2.5g.L<sup>-1</sup>.

The fermentation was conducted in Erlenmeyer flasks of 125 mL containing the medium at 20% of its maximum capacity. 10<sup>8</sup> spores were inoculated in all media. The flasks were incubated in a Shaker at 28°C and 150 rpm. Samples were taken every 24 hours for 4 days for the evaluation of enzyme yields (Onofre *et al.*, 2012).

#### 2.2.3 Recovery of the Enzyme from the Fermented Media

For the recovery of the enzyme from the solid fermented medium, 5 mL of NaCl 1% solution was added for each gram of solid substrate and maintained under stirring for 1 hour at 100 rpm. For the liquid medium, a solution of NaCl 1% was added until the volume reached 50 mL. After this process, the suspension was filtered to obtain the crude extract, and then centrifuged at 2000 rpm for 8 minutes, discarding the precipitate.

### 2.3 ENZYMATIC ASSAY

The activity of  $\alpha$ -amylase was determined by measuring the levels of reducing sugars in solution as a result of the action of the  $\alpha$ -amylase on the starch. The activity was determined in samples in triplicate by quantifying the reducing sugars (glucose) with the Miller (1959) and Fernandes *et al.*, (2007) method. A mixture containing 0.5 mL of enzymatic extract; 0.5 mL of a starch 0.5% solution in a Tris-HCl buffer 0.05M pH 8.5, and 0.2 mL of the same buffer was incubated at 90°C for 10 minutes. After this period, 1.0 mL of the Miller reagent (3,5-Dinitrosalicylic acid) was added to the reaction.

The mixture was placed in boiling water for 10 minutes, then cooled in an ice bath for 5 minutes and 4.8 mL of distilled water was added. The developed color was measured with a SHIMADZU UV-mini 1240 spectrophotometer, using a wavelength of 540nm. The same procedure was performed with the control, except that the miller reagent (3,5-Dinitrosalicylic acid) was added together with the enzyme to the starch 0.5% solution, and this mixture was placed in boiling water as described above.

The content of reducing sugars was determined through a glucose curve. One unit of enzyme activity (U) was defined as the amount of enzyme required to produce 1  $\mu$ mol of reducing sugar per minute per mL from the soluble starch under the test conditions (Onofre *et al.*, 2011; Onofre *et al.*, 2012).

## 2.4 CHARACTERIZATION OF THE $\alpha$ -AMYLASE

### 2.4.1 Optimal pH for Enzymatic Activity

For the assessment of the influence of pH on enzyme activity, the enzymatic assay was conducted according to item 2.3, while varying the addition of buffer solution: pH 5.0 - 6.0 (acetate buffer), 6.5 - 7.5 (phosphate buffer), 8.0 - 8.5 (Tris-HCl buffer) (Thys, 2004).

### 2.4.2 Optimal Temperature for Enzymatic Activity

For the assessment of the influence of temperature on enzyme activity, the enzymatic assay was conducted according to item 2.3, while only varying the incubation temperature: 10, 20, 30, 37, 45 and 65°C (Silva, 2002).

### 2.4.3 Influence of Ions on Enzymatic Activity

For the assessment of the influence of ions on enzyme activity the enzyme was incubated for 10 min at room temperature with the ions CaCl<sub>2</sub>, BaCl<sub>2</sub>, HgCl<sub>2</sub>, FeCl<sub>3</sub> and MgCl<sub>2</sub> at concentrations of 1 and 5 mM, performing the enzymatic assay after this procedure (Giongo, 2006).

### 2.4.4 Thermal Stability of $\alpha$ -Amylase

The thermal stability of the amylase was tested by incubating the enzyme for 30 minutes at the temperatures of: 10, 20, 30, 37, 45 and 65°C, performing the enzymatic assay after this procedure as described by Rasiah and Rehm, (2009).

### 2.4.5 Stability of $\alpha$ -Amylase at Different pH Values

The stability of  $\alpha$ -amylase at different pH values was tested by incubating the enzyme for 30 minutes in the following buffer solutions (100 mM): pH 5.0 - 6.0 (acetate buffer), 6.5 - 7.5 (phosphate buffer), 8.0 - 8.5 (Tris-HCl buffer), performing the enzymatic assay after this procedure as described by Thys, (2004).

## III. RESULTS AND DISCUSSION

Through the analysis of the fermentation processes, one can observe that the endophytic strain of *P. digitatum*, isolated from *Baccharis dracunculifolia* D.C. (Asteraceae) had better results in the solid medium when compared with fermentation in the liquid medium, yielding 1625 U/mL through solid state fermentation in the medium containing only rice bran, while in submerged fermentation these values didn't exceed 712 U/mL in the medium containing inorganic nitrogen (Liq/Ino).

By comparing the data obtained in this study with those found by Spier (2005) working with *Penicillium* sp., one can see that he obtained similar results as those of this study, since he observed a production of 1690 U/mL of fungal amylases in SSF, with yields surpassing double the activity achieved in SmF. This same behavior was observed by Hu *et al.*, (2013), who found that the fungus *Penicillium* sp. in a semi-solid medium had better results than in submerged fermentation and in a medium supplemented with an inorganic fraction (NaNO<sub>2</sub>) of nitrogen. In Figure 2 the enzyme yield data (in total units) in the different media over the course of 120 hours at 24-hour intervals, is presented.

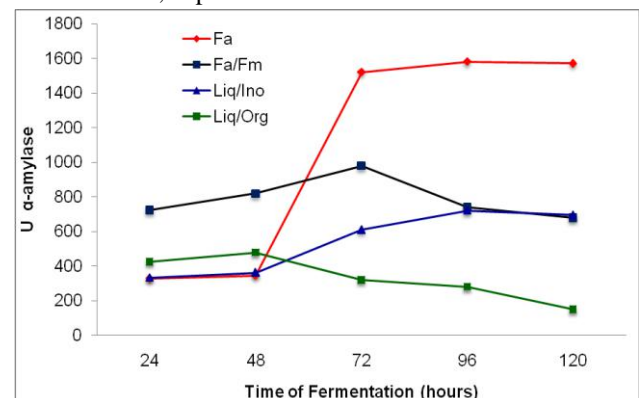


Fig. 2: Yield of  $\alpha$ -amylase as a function of time in solid media containing only rice bran (Fa) and supplemented with manioc starch (Fa/Fm), and in liquid medium with the organic nitrogen source urea (Liq/Org) and the inorganic source NaNO<sub>2</sub> (On/Innovation).

SSF has been described as an excellent option for the growth of the filamentous fungi. In general, fungi have a good ability to grow on solid substrates, which explains their good adaptation to SSF. And for this reason, they have been widely used in this process, mainly to obtain enzymes (Silva, 2002; Lei *et al.*, 2014).

One of the main characteristics of SSF is the use of substrates with low water activity, in which the conditions for growth are similar to the natural habitat of fungi, which facilitates their growth on the solid substrate and the yield of large quantities of enzymes. The organic matter present in this material is used as a source of

energy for growth, and the carbon is used for the synthesis of cellular biomass and of the products of the microbial metabolism (Mitchell and Lonsane, 1992; Hattori *et al.*, 2013; Dris *et al.*, 2014).

It should be noted, however, that the SSF technology should not be seen as a technique that replaces submerged fermentation. In fact, each technique has its own potential and particularities. However, there is a consensus about the need for ongoing research on the factors related to SSF in order to take advantage of the full potential of this technology (Pandey *et al.*, 2001).

According to Del Bianchi *et al.*, (2001) and Pandey (2002), the control of certain variables is necessary to obtain products with constant and uniform characteristics. One can therefore state that the observation of these factors, and the correct handling in relation to each one of them, will certainly bring about better results in the solid state fermentation process. Environmental conditions, such as temperature, pH, water activity, oxygen level and the concentration of nutrients and products, significantly affect cell growth and product formation.

When the liquid fermentation media are compared, the fungus can be observed to have adapted better to the inorganic source of nitrogen, showing a maximum yield of 712 U/mL, while in the medium containing organic nitrogen the peak yield of total amylases was 438 U/mL.

According to Gupta *et al.*, (2003), organic nitrogen sources are preferred for the production of  $\alpha$ -amylase by bacteria. On the other hand, various inorganic salts, such as ammonium sulfate, sodium nitrate and ammonium nitrate, have been reported in improved yields of  $\alpha$ -amylase by fungi.

Both the solid media had good enzyme yields: 1625 U/mL in the medium containing rice bran, and 932 U/mL in the medium containing rice bran supplemented with manioc starch.

One would expect that the medium supplemented by starch would have higher enzyme yields, since the presence of starch should induce the production of amylolytic enzymes, but this was not the behavior observed. This fact may have occurred due to the presence of starch, which gelatinized after heat treatment (sterilization), making the medium more compact. As such, this made aeration of the solid medium harder. Spier (2005) reported this same limitation with the use of starch in submerged cultivation, noting that enzyme activity decreased with the increase of the concentration of starch in the medium.

### 3.1 CHARACTERIZATION OF THE ENZYME

#### 3.1.1 Optimal pH for Enzymatic Activity

The  $\alpha$ -amylase produced by SSF in the medium containing rice bran were used in the tests for

characterization of the enzyme. Figures 3 and 4 show the activity of the enzyme as a function of pH and temperature, respectively.

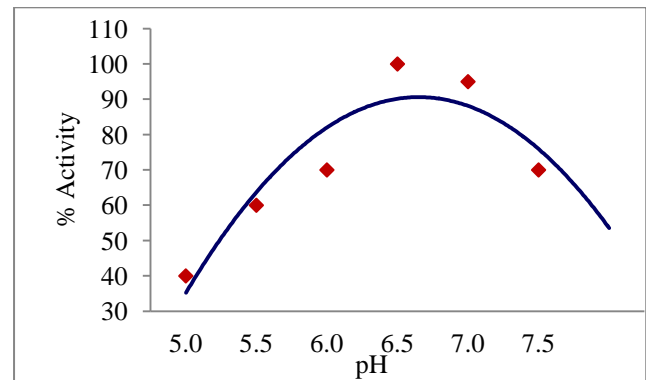


Fig. 3: Curve representing the optimal pH for  $\alpha$ -amylase activity.

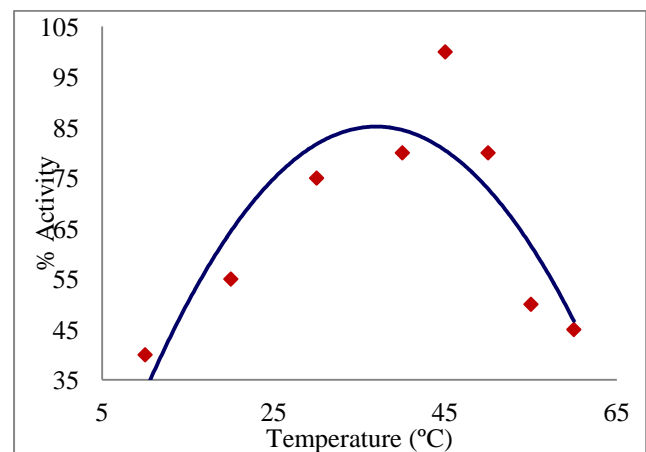


Fig. 4: Curve representing the optimal temperature for  $\alpha$ -amylase activity.

The enzyme showed optimal pH at 6.5, while maintaining 96% of its activity at pH 7.0, characterizing it as a neutral  $\alpha$ -amylase. This data is consistent with Figueira *et al.*, (2000) and Oliveira *et al.*, (2010) who produced amylases with the fungi *Fusarium moniliforme* and *Aspergillus flavus*, obtaining optimal pH values close to 6.9, demonstrating the trend for the production of slightly neutral amylases by filamentous fungi. Spier (2005), on the other hand, obtained a higher yield of amylolytic enzymes with an initial pH equal to 4.0 when working with *Aspergillus niger*, which shows that each species of fungus may have a different behavior in specific pH.

According to Soccol (1992), Onofre *et al.*, (2011); Onofre *et al.*, (2012), the growth capacity of fungi is limited in extreme conditions of acidity and alkalinity. This characteristic is of extreme importance in fermentation processes since they show that under these conditions the vast majority of the bacteria responsible for the contamination of fermentation processes are inhibited.

The optimal temperature was 45°C, with the enzyme being completely inactivated at the temperature of 65°C. Spier (2005) obtained similar results to those in this study for a fungal amylase, reporting optimal pH at 6.0 and optimal temperature between 45-55°C. This behavior is also in agreement with Kundu *et al.*, (1973) and Ueno *et al.*, (1987) cited by Pandey *et al.*, (2005), who reported that optimal yields of  $\alpha$ -amylase were obtained at temperatures between 30 and 37°C.

### 3.1.2 Influence of Ions on Enzymatic Activity

Metal ions have a variety of functions in proteins. These ions may be directly involved in the catalytic process during the enzymatic reaction, or they may participate in electron transfer or redox reactions (Najafi *et al.*, 2005). The stabilization of some enzymes can be induced, mainly, by such divalent ions as  $\text{Ca}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Zn}^{2+}$  e  $\text{Mg}^{2+}$ . If used in low concentrations, these ions can stabilize the tertiary structure of the protein, promoting the formation of cross-links that provide a greater stability to it (Tomazic, 1991). Figure 5 shows the behavior of  $\alpha$ -amylase regarding the incubation with some ions.

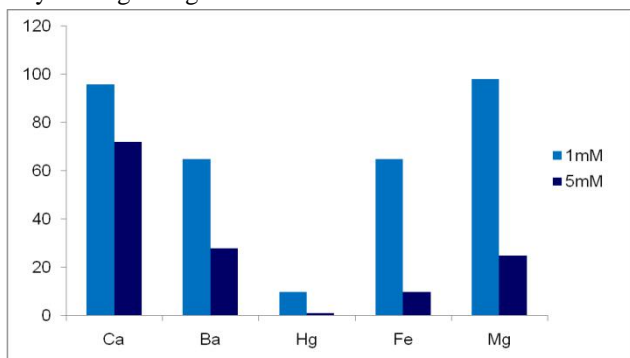


Fig. 5: Influence of ions on enzyme activity at the concentrations of 1 and 5mM.

All the ions tested at concentrations of 1 and 5 mM were associated with a drop in activity of the enzyme of at least 14 %, with the ions  $\text{Hg}^{2+}$ , at both concentrations, and  $\text{Fe}^{3+}$ , at the concentration of 5 mM, leading to the complete loss of enzyme activity.

The change in activity per ion is a very particular characteristic of each enzyme. Some enzymes may require divalent ions for their activation or as a cofactor, commonly  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Zn}^{2+}$  e  $\text{Mn}^{2+}$ , increasing their activity. However, some ions may generate a drastic drop in activity, such as  $\text{Hg}^{2+}$ ,  $\text{Fe}^{3+}$ , and may lead to total inhibition (Giongo, 2006).

It should be noted that high concentrations of ions can have an inhibitory effect. Some studies also show that the stability of  $\alpha$ -amylase is compromised in the presence of small quantities of these ions (Yang and Liu, 2004; Bernhardtsson *et al.*, 2005; Hashim *et al.*, 2005).

### 3.1.3 Thermal Stability of $\alpha$ -Amylase and at Different pH Values

pH is one of the most important factors that affect fermentation processes, since it can change the chemicals of the culture medium, ionize polar molecules, affect enzymatic reactions and the post-translation processes of enzymes.

The pH of the culture medium can influence microbial growth, and induce the expression of genes that result in changes in phenotypes, such as changes in morphology, physiology or the expression of enzymes. The limitation of growth has been associated with a reduction in the production and/or activity of extracellular enzymes (Madigan *et al.*, 2004). Figures 6 and 7 shows the stability of the enzyme regarding the incubation at different pH values and temperatures, respectively.

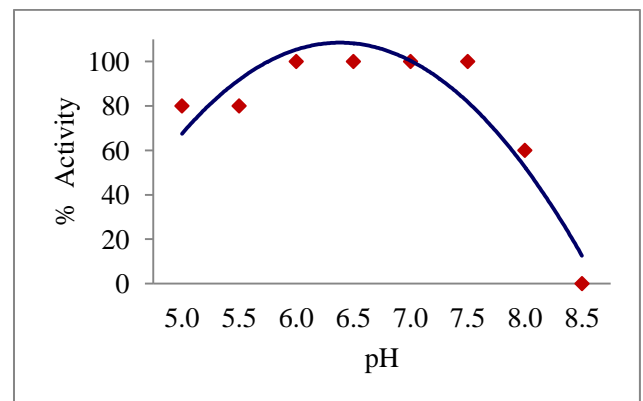


Fig. 6: Stability of  $\alpha$ -Amylase regarding incubation at Different pH Values.

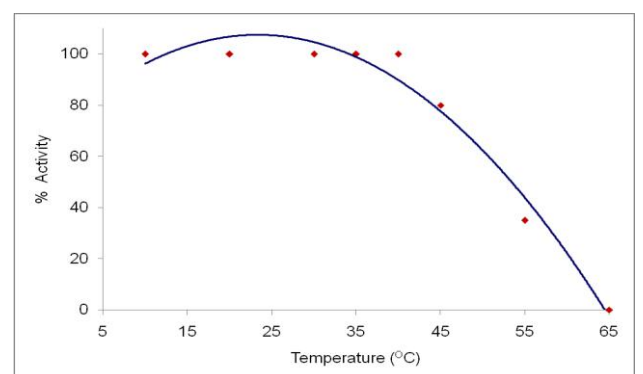


Fig. 7: Stability of  $\alpha$ -amylase regarding different incubation temperatures.

In the more extreme pH values tested,  $\alpha$ -amylase underwent a reduction in enzyme activity of at least 24%, with it being completely inhibited at pH 8.5. In the 6.0 - 7.5 pH range, the enzyme proved to be quite stable, maintaining at least 95% of its activity, with this being the recommended pH range for the application of the enzyme.

Thermal resistance is considered one of the most important criteria for the industrial application of the enzyme, given that most processes require the use of high temperatures (Ray and Nanda, 1996). The use of high temperatures for the activity of these enzymes, however, depends on the stability limit of the protein, since it will have a range within which its structure is maintained.

Outside of this range, denaturation would occur, resulting in a loss of activity. An increase in the use of the  $\alpha$ -amylase enzyme in biotechnology has occurred due to its extensive range of working conditions, including high temperatures, extreme pH, and the presence of surfactants and organic solvents (Tanaka and Hoshino, 2002).

When the enzyme was incubated at temperatures of 10 to 37°C, it preserved its full activity. When a temperature of 45°C was employed, the enzyme suffered a decline in residual activity, although this has been its optimal temperature of operation, indicating that the produced  $\alpha$ -amylase should be applied at temperatures below 45°C to maintain its activity for a longer period of time.

Rasiah and Rehm (2009) obtained amylase which proved quite stable at 85°C. Figueira *et al.*, (2000) produced amylases with the fungi *Fusarium moniliforme* and *Aspergillus flavus*, obtaining thermal stability close to 20°C. Fungal Amylases are more sensitive to the elevation of temperature, tending to suffer a drastic drop in activity at temperatures above 50°C (Spier, 2005).

Understanding the thermo-stability of enzymes will promote their addition to products that need to go through some form of heat treatment during their processing, while preventing inactivation or with the intentional inactivation of the enzyme when the expected result of its catalytic activity has been obtained.

#### IV. CONCLUSION

With the results obtained, it is possible to conclude that the production of  $\alpha$ -amylase through solid state fermentation (SSF) was more efficient than the production of the enzyme through submerged fermentation (SmF), obtaining more than twice the activity found in SmF with SSF, with values of 1625 and 712 U/mL for SSF and SmF, respectively.

At 120 hours of solid state fermentation using rice bran as substrate, 1625 U/mL was obtained, demonstrating the excellent  $\alpha$ -amylase secretion ability of the D1-FB strain of the endophytic fungus, *Penicillium digitatum*, isolated from *Baccharis dracunculifolia* D.C. (Asteraceae).

The enzyme characterization tests revealed an optimal pH at 6.5 and an optimal temperature at 45°C. The  $\alpha$ -amylase produced was stable in the neutral pH range, but it showed great drops in activity with temperatures above 45°C and in the presence of ions at a concentration of 5 mM.

These results allow for the conclusion that the produced  $\alpha$ -amylase may be a new enzyme alternative for application in industries that require the saccharification of starch.

#### CONFLICTS OF INTEREST

The authors declare there are no ethical, publishing or financial conflicts of interest regarding the data of this study.

#### REFERENCES

- [1] Bernhardsdotter ECMJ, Nig JD, Garriott OK, Pusey ML (2000). Enzymic properties of an alkaline chelator-resistant  $\alpha$ -amylase from an alkaliphilic *Bacillus* sp. isolate L1711. *Process Biochem.* 40: 2401–2408.
- [2] Brena BM, Pazos C, Franco-Fraguas L, Batista-Vieira F (1999). Chromatographic methods for amylases. *Journal of Chromatography*, 684: 217-237.
- [3] Corradini E, Lotti C, Medeiros ES, Carvalho AJF, Curvelo AAS, Mattoso LHC (2005). Estudo comparativo de amidos termoplásticos derivados do milho com diferentes teores de amilose. *Polímeros*, 15: 268-273.
- [4] Del Bianchi VL, Moraes IO, Capalbo DMF (2001). Fermentação em estado sólido. In: Lima, U.A. et al. (Coord.). *Biotecnologia Industrial: Engenharia Bioquímica*. 1. Ed., v. 3. São Paulo: Edgard Blucher. 13:247-276.
- [5] Driss D, Zouari-Ellouzi S, Chaari F, Kallel F, Ghazala I, Bouaziz F, Ghorbel R, Chaabouni SE. (2014). Production and *in vitro* evaluation of xylooligosaccharides generated from corncobs using immobilized *Penicillium occitanis* xylanase. *Journal of Molecular Catalysis B: Enzymatic*. 102:146-153.
- [6] Fernandes LP, Ulhoa CJ, Asquieri ER, Monteiro VN (2007). Produção de amilases pelo fungo *Macrophomina phaseolina*. *Revista Eletrônica de Farmácia*. 4(1): 43-51.
- [7] Fernandes MLM (2006). Produção de lipases por fermentação no estado sólido e sua utilização em biocatálise. (2006). 130f. Dissertação (Doutorado em Química) - Setor de Ciências Exatas - Universidade Federal do Paraná. Curitiba, Brasil.
- [8] Figueira ELZ, Sá MC, Ida EI, Hirooka EY (2000). Produção e caracterização de amilases de *Fusarium moniliforme* e *Aspergillus flavus*. *B.CEPPA*. 18(1):13-26.
- [9] Germano S (2000). Desenvolvimento de bioprocessos para produção, caracterização e purificação de proteases de *Penicillium* sp. por fermentação no estado sólido. 2000. 142f. Tese

- (Doutorado em Processos Biotecnológicos) - Setor de Tecnologia - Universidade Federal do Paraná. Curitiba, Paraná, Brasil.
- [10] Giongo JL 2006. Caracterização e aplicação de proteases produzidas por linhagens de *Bacillus* sp. (2006). 95f. Dissertação (Mestrado em Microbiologia Agrícola e do Ambiente) – Faculdade de Agronomia – Universidade Federal do Rio Grande do Sul. Porto Alegre, Rio Grande do Sul, Brasil.
- [11] Gupta R, Gigras P, Mohapatra H, Goswami VK, Chauhan B (2003). Microbial  $\alpha$ -amylases: a biotechnological perspective. *Process Biochemistry*. 38(11): 1599-1616.
- [12] Hashim SO, Delgado OD, Martínez MA, Kaul RH, Francis JM, Mattiasson B (2005). Alkaline active maltohexaose-forming  $\alpha$ -amylase from *Bacillus halodurans* LBK 34. *Enzyme and Microbial Technology*. 36: 139-146.
- [13] Hattori T, Kato Y, Uno S, Usui T (2001). Mode of action of a  $\beta$ -(1 $\rightarrow$ 6)-glucanase from *Penicillium multicolor*. *Carbohydrate Research*. 366:6-16. 25.
- [14] Hu Y, Liu G, Li Z, Qin Y, Qu Y, Song X (2013) G protein-cAMP signaling pathway mediated by PGA3 plays different roles in regulating the expressions of amylases and cellulases in *Penicillium decumbens*. *Fungal Genetics and Biology*. 58(59): 62–70.
- [15] Lei Y, Liu G, Li Z, Gao L, Qin Y, Qu Y (2014). Functional characterization of protein kinase CK2 regulatory subunits regulating *Penicillium oxalicum* asexual development and hydrolytic enzyme production. *Fungal Genetics and Biology*, 66: 44-53.
- [16] Lorentz RH (2005). Seleção de isolados de *Paenibacillus* spp com atividade enzimática e antimicrobiana. 2005. 103f. Dissertação (Mestrado em Microbiologia Agrícola e do Ambiente) – Faculdade de Agronomia – Universidade Federal do Rio Grande do Sul. Porto Alegre, Rio Grande do Sul, Brasil.
- [17] Madigan MT, Martinko JM, Parker J (2004). *Microbiologia de Brock*. 10. ed. São Paulo: Person Education. 517p.
- [18] Michelin M (2005). Estudo da glucoamilase e da  $\alpha$ -amilase produzidas pelo fungo *Paecilomyces variotii*: purificação, caracterização bioquímica e relações filogenéticas. 2005. 160f. Dissertação (Mestrado em Ciências) – Departamento de Biologia – Universidade de São Paulo, Ribeirão Preto, Brasil.
- [19] Miller GL (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Washington*, 31(3): 426-428.
- [20] Mitchell DA, Lonsane BK (1992). Definition, characterization and economic evaluation. In *Solid Substrate Cultivation*, Elsevier, London.
- [21] Moraes LMP (2004). Amilases. In: Said, S.; Pietro, R. *Enzimas como agentes Biotecnológicos*. Ribeirão Preto: Legis Summa. p.123-128.
- [22] Myers AM, Morell MK, James MG, Ball SG (2000). Recent progress toward understanding biosynthesis of the amylopectin crystal. *Plant Physiology*. 122:989-997.
- [23] Najafi MF, Deobagkar D, Deobagkar D (2005). Purification and characterization of an extracellular  $\alpha$ -amylase from *Bacillus subtilis* AX20. *Protein Expr. and Purif.* 41:349-354.
- [24] Oliveira AN, Flor NS, Oliveira LA (2010). Influência do pH e temperatura sobre a atividade amilolítica de rizóbios isolados de solos da Amazônia. *Acta Amazonica*. 40(2):401-404.
- [25] Oliveira AN, Oliveira LA, Andrade JS, Chagas-Junior AF (2007). Produção de amilase por rizóbios, usando farinha de pupunha como substrato. *Ciência e Tecnologia de Alimentos*. 27(1): 61-66.
- [26] Onofre SB, Groff SA, Sartori A, Bertolini J, Kagimura FY, Rotta D, Mazzali L, Steilmann P (2011). Amylolytic Enzymes Produced by the Fungus *Colletotrichum gloeosporioides* in Rice Semi-Solid Fermentation. *Journal of Yeast and Fungal Research*. 2: 28-32.
- [27] Onofre SB, Groff SA, Sartori A, Bertolini J, Kagimura FY, Rotta D, Mazzali L, Steilmann P (2012). Production of  $\alpha$ -Amylase and Amyloglucosidase by the Fungus *Cylindrocladium* sp. in Semi-Solid State Fermentation. *Journal of Microbiology Research*. 2: 123-126.
- [28] Pandey A (2002). Solid state fermentation. *Biochemical. Engineering Journal*, 3636:1-4.
- [29] Pandey A, Soccol CR, Rodrigues-Leon JA, Nigam P (2001). Solid state fermentation in biotechnology. *Nova Deli, Asiatech*. p.221, 2001.
- [30] Pandey A, Webb C, Soccol CR, Larroche C (2005). *Enzyme Technology*. 1º ed. New Delhi: Asiatech Publishers, Inc, p. 760.
- [31] Paris LD (2008). Produção de enzimas fúngicas por fermentação em estado sólido das sojas orgânica, transgênica e convencional. 115p., (2008). Dissertação de Mestrado. Programa de Pós-graduação em Engenharia Química, Centro de Ciências Exatas, Universidade Estadual do Oeste do Paraná (Unioeste), Toledo, Paraná, Brasil.
- [32] Rasiah IA, Rehm BHA (2009). One-step production of immobilized  $\alpha$ -amylase in recombinant *Escherichia coli*. *Applied and Environmental Microbiology*. 75(7): 2012-2016.

- [33] Ray RR, Nanda G (1996). Microbial  $\beta$ -amylases: Biosynthesis, characteristics, and industrial applications. *Critical Reviews in Microbiology*. 22: 181-199.
- [34] Rocha CP (2010). Otimização da Produção de Enzimas por *Aspergillus niger* em Fermentação em Estado Sólido. 136p. Dissertação de Mestrado. Programa de pós-graduação em Engenharia Química, Universidade Federal de Uberlândia, Faculdade de Engenharia Química, Uberlândia, Brasil.
- [35] Silva ARZ (2002). Desenvolvimento de bioprocessos para a produção de fitase por *Aspergillus niger* em fermentação no estado sólido utilizando subprodutos agrícolas para aplicação como aditivo na alimentação de aves e suínos. 2002. 96f. Dissertação (Mestrado em Tecnologia de Alimentos) – Setor de Tecnologia – Universidade Federal do Paraná. Curitiba, Brasil.
- [36] Soccol CR (1992). Physiologie et Métabolisme de *Rhizopus* em Culture Solide et Submergée em Relation Avec la Dégradation d'Amidon et la Production d'Acide L(+) Lactique. 218p. Thèse de Doctorat. Mention Génie enzymatique, Bioconversion et Microbiologie, Université de Technologie de Compiègne. Compiègne-France.
- [37] Spier MR (2005). Produção de enzimas amilolíticas fúngicas  $\alpha$ -amilase e amiloglicosidase por fermentação no estado sólido. 2005. 178f. Dissertação (Mestrado em Tecnologia de Alimentos) – Setor de Tecnologia – Universidade Federal do Paraná. Curitiba, Brasil.
- [38] Tanaka A, Hoshino E 2002. Thermodynamic and activation parameters for the hydrolysis of amylose with *Bacillus*  $\alpha$ -amylases in a diluted anionic surfactant solution. *Journal of Bioscience and Bioengineering*. 93:485-490.
- [39] Thys RCS 2004. Produção, caracterização, purificação e aplicação de uma protease produzida pelo microrganismo *Microbacterium* sp. *kr10*. 2004. 114f. Dissertação (Mestrado em Microbiologia Agrícola e do Ambiente) – Faculdade de Agronomia – Universidade Federal do Rio Grande do Sul. Porto Alegre, Brasil.
- [40] Tomazic SJ (1991). Protein stabilization. In: *Biocatalysts for industry*. Topic in Applied chemistry. Ed. Plenum Press, New York and London. 330p.
- [41] Yang CH, Liu WH (2014). Purification and properties of a maltotriose producing  $\alpha$ -amylase from *Thermobifida fusca*. *Enzyme and Microbial Technology* 35: 254–260.