HIGH INTERLEUKIN-6, LOW CD4⁺ AND CD8⁺ T-LYMPHOCYTES EXPRESSIONS AS RISK FACTORS OF CERVICAL CARSINOMA INFECTED BY HUMAN PAPILLOMA VIRUS TYPE-52

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ABSTRACT

In Indonesia cervical carcinoma is the most common cancer in women and one of the leading cause of mortality. High risk human papillomavirus (HPV) is the major risk factor of cervical cancer. This study aims to know the role of IL-6, CD4⁺ and CD8⁺ T-lymphocyte for the risk of cervical carcinoma infected by HPV52. This study was a case control study, specimens of cervical carcinoma patients infected by HPV type-52 as the case group and HPV type-16 or 18 as the control group. HPV genotyping used SPF10 primer and type specific E7 primer by LiPA. Immunohistochemistry method was used to know expression of IL-6, CD4⁺ and CD8⁺ T lymphocyte. Pearson's χ^2 test was applied with statistical significance was set at the 2-sided 0.05 level. The odds ratios (OR) were calculated for the risk, with 95% confidence intervals on SPSS 16.0 for windows. PCR examination was performed in 185 paraffin-embedded tissue. The risk of high IL-6 expression in cervical carcinoma infected by HPV type-52 was statistically significant 6-fold higher compare with cervical carcinoma infected by HPV type 16 (OR = 6.00; Cl 95% = 1.13-31.99; p = 0.03; p < 0.05) and HPV type 18 (OR = 6.00 ; Cl 95% = 1.13-31.99; p = 0.03; p < 0.05). The risk of low CD4⁺ T lymphocyte expression in cervical carcinoma infected by HPV type 52 was statistically significant 6-fold higher and 7.43fold higher respectively compare with cervical carcinoma infected by HPV type 16 (OR = 6.00; CI 95% = 1.003-35.91; p = 0.04; p < 0.05) and HPV type 18 (OR = 7.43; Cl 95% = 1.23-45.01; p = 0.02; p < 0.05). The risk of low CD8⁺ T lymphocyte expression in cervical carcinoma infected by HPV type 52 was statistically significant 13.5-fold higher and 11-fold higher respectively compare with cervical carcinoma infected by HPV type 16 (OR = 13.50; CI 95% = 1.42-128.26; p = 0.01; p < 0.05) and HPV type 18 (OR = 11.00; CI 95% = 1.16-103.94; p = 0.02; p < 0.05). No significance different between cases and controls group in mean-age, parity and sexual activity (p > 0.05). In conclusion, this study found that high IL-6 expression, low CD4⁺ and CD8⁺ T lymphocyte expression were the risk factors of cervical carcinoma infected by HPV type 52.

Keywords: IL-6, CD4⁺T-lymphocyte, CD8⁺T-lymphocyte, cervical carcinoma, HPV type-52

INTRODUCTION

Cervical cancer is the second most common cancer in women in developing countries. It accounts for 80-90% of all cancer cases.^{1,2,3} In Indonesia cervical carcinoma is the most common cancer in women and one of the leading cause of mortality. In Bali cervical cancer is the second most common after breast cancer, it accounts for 27.5%.^{4,5,6} Many attempts have

Corresponding Author: I G. A. S. Mahendra-Dewi Anatomical Pathology Department, Faculty of Medicine Udayana University, Bali-Indonesia E-mail: mahendradewi@rocketmail.com been carried out, however, many obstacle and could not decrease the incidence and mortality rate of cervical cancer were faced. High risk oncogenic human papilloma virus (HPV) is the major risk factor of cervical cancer. It can be detected in about 90% of all cervical cancers.^{5,7,8} The two predominant high-risk HPV types are HPV-16 and 18 associated with more than 90% of cervical cancers worldwide.^{5,8,9} In Bali HPV type-52 is the most common type found in cervical cancer patients, account for 18%.¹⁰ The main pathogenesis of cervical carcinoma infected by HPV type-16 and 18 have been well established and widely accepted worldwide through tumor suppressor gene p53 and pRb. Further study is needed to know the pathogenesis of cervical carcinoma infected by other high risk HPV. ^{8,11,12} The aim of this study was to know the role of immunologic factor, especially IL-6, CD4⁺ Tlymphocyte and CD8⁺ T- lymphocyte for the risk of cervical carcinoma infected by HPV type-52, because HPV type-52 is the most common type in Bali. The results of this study were expected to be used for new input in pathogenesis of cervical cancer infected by HPV type-52, for early diagnosis and as predictor factors.

MATERIAL AND METHODS

Specimen collection and histopathological diagnosis

This study was a case control study. The sample consisted of biopsy or operation specimens of cervical carcinoma patients infected by HPV type-52 as the case group and biopsy or operation specimens of cervical carcinoma patients infected by HPV type-16 or 18 as the control group. The study was conducted during the years 2011-2012 at Department of Obstetrics and Gynecology Faculty of Medicine Udayana University/Sanglah Hospital Denpasar for the biopsy or cervical operation. And then, these specimens were processed at Department of Anatomical Pathology Faculty of Medicine Udayana University/Sanglah Hospital Denpasar for histopathology diagnosis.

HPV DNA detection and typing

The paraffin-embedded tissue which have been diagnosed as cervical carcinoma were sent for detection and genotyping of HPV DNA by PCR-based methods at Department of Pathology, Leiden University Medical Centre, the Netherland and Molecular Biology Unit, Faculty of Medicine, Udayana University. HPV DNA was detected in 2 steps. In the first step, the SPF10 primer at several dilution were used to amplified the DNA. In the second step, the positive products was performed using specific probes type-specific E7 primer by LiPA which could detected 25 HPV types to know the evidence of HPV type 52, 16 and 18.

Immunohistochemistry examination

Ten specimens with HPV type 52 positive were used as cases. The specimen of HPV type 16 or 18 positive, consist of 20 specimens each as controls. Immunohistochemistry examination was carried out to all of these 50 specimens for examination of IL-6, CD4⁺ T-lymphocyte and CD8⁺ T-lymphocyte expression at Department of Anatomical Pathology Faculty of Medicine Gajah Mada University/ Dr. Sardjito Hospital Yogyakarta.

Statistical analysis

The results of this study was analyzed on SPSS 16.0 for windows, i.e. Shapiro-Wilk test for normality of data, Levene's test for homogeneity, logistic regression for ratio odd (RO), and Pearson's chi-square test for significance. Statistical significance for these tests was set at the 2-sided 0.05 level with 95% confidence intervals (CIs).

RESULTS

The case group in this study consist of 10 specimens of cervical carcinoma infected by HPV type-52, the control group consist of 20 specimens of cervical carcinoma infected by HPV type 16 and 20 specimens of cervical carcinoma infected by HPV type 18. Age, parity and sexual activity were controlled by analysis. The results of this study were displayed in Table 1-3 and Figure 1-4.

Table 1IL-6, CD4⁺ and CD8⁺ T-lymphocyte expression inCervical Carcinoma Infected by HPV Tipe-52 and 16

	HPV Type		DO.		
	52	16	- KU	ρ	CI 95%
high	6	4	6.00	0.03	1.13-31.99
IL-6					
low	4	16			
high	1	12	6.00	0.04	1.00-35.91
$CD4^+$					
low	8	8			
high	1	12	13.50	0.01	1.42-128.26
CD8 [⁺]					
low	9	8			
a					

Significant at p < 0.05

The risk of high IL-6 expression in cervical carcinoma infected by HPV type 52 was statistically significant 6-fold higher compare with cervical carcinoma infected by HPV type 16 (RO = 6.00; Cl 95% = 1.13-31.99; p = 0.03; p < 0.05). The risk of low CD4⁺ T-lymphocyte expression in cervical carcinoma infected by HPV type 52 was statistically significant 6-fold higher compare with cervical carcinoma infected by HPV type 16 (RO = 6.00; Cl 95% = 1.003-35.91; p = 0.04; p < 0.05). The risk of low CD8⁺ T lymphocyte expression in cervical carcinoma infected by HPV type 52 was statistically significant 13.5-fold higher compare with cervical carcinoma infected by HPV type 16 (RO = 13.50; Cl 95% = 1.42-128.26; p = 0.01; p < 0.05).

As can be seen from Table 2, the risk of high IL-6 expression in cervical carcinoma infected by HPV type-52 was statistically significant 6-fold higher compare with cervical carcinoma infected by HPV type Indonesían Journal of Bí*omedical Sciences Vo*lume 7, Number 2, July-December 2013: 57-62 Prínt-ISSN: 2085-4773, E-ISSN: 2302-2906.

18 (RO = 6.00; CI 95% = 1.13-31.99; p = 0.03; p < 0.05). The risk of low CD4⁺ T lymphocyte expression in cervical carcinoma infected by HPV type 52 was statistically significant 7.43-fold higher compare with cervical carcinoma infected by HPV type 18 (OR = 7.43 ; CI 95% = 1.23-45.01; p = 0.02; p < 0.05).

Table 2 IL-6, CD4⁺ and CD8⁺ T-lymphocyte expression in Cervical Carcinoma Infected by HPV Tipe-52 and 18

	HPV Type		DO				
	52	18	RU	ρ	CI 95%		
high	6	4	6.00	0.03	1.13-31.99		
IL-6							
low	4	16					
high CD4 ⁺	2	13	7.43	0.02	1.23-45.01		
low	8	7					
high CD8⁺	1	11	11.00	0.02	1.16-103.94		
low	9	9					

Significant at p < 0.05

The risk of low CD8⁺ T-lymphocyte expression in cervical carcinoma infected by HPV type-52 was statistically significant 11-fold higher compare with cervical carcinoma infected by HPV type 18 (RO = 11.00; CI 95% = 1.16-103.94; p = 0.02; p < 0.05).

Table 3 Mean-age, Parity and Sexual Activity between Case and Control Group

Variable	Case	Control	p			
valiable	HPV-52	HPV-16				
Age (years)	53.40±13.10	48.20±10.52	0.24			
Parity	3.40±1.35	2.80±1.11	0.23			
Sexual Activiy	2.40±3.18	23.40±2.76	0.54			
Variable	Case	Control				
valiable	HPV-52	HPV-18	μ			
Age (years)	53.40±13.10	50.80±10.24	0.69			
Parity	3.40±1.35	3.70±1.49	0.64			
Sexual Activiy	2.40±3.18	22.05±2.70	0.11			
Significant at $n < 0.05$						

Significant at p < 0.05

No significance different between case and controls group in mean-age, parity and sexual activity (p > 0.05).



Figure 1



bp: base pair, Line 1: Standard DNA, Line 2: Positive control (102 bp), Line 3: Negative control, Line 4 and 5: Positive result (102 bp), Line 6: Negative result.



Figure 2

The product of PCR amplification on HPV DNA type-16 Remarks:

bp: base pair, Line 1: Standard DNA, Line 2: Positive control (102 bp), Line 3: Negative control, Line 6 and 10: Positive result (102 bp), Line 4, 5, 7, 8, 9 and 11: Negative results.



Figure 3

The product of PCR amplification on HPV DNA type-18 Remarks:

Bp: base pair Line 1: Standard DNA, Line 2: Positive control (102 bp), Line 3: Negative control, Line 4, 5, 6 and 7: Positive result (102 bp).



Immunohistochemistry Staining

Remarks:

- A. Strong positive expression of IL-6 (450x)
- B. Strong positive expression of CD4⁺ T Lymphocyte (450x)
- C. Strong positive expression of CD8⁺ T Lymphocyte (450x)

DISCUSSION

A total of 185 cervical carcinoma specimens were examined in this study for genotyping of HPV by PCR. The positive HPV prevalence was 108 (58.38%). A total of 12 different high risk HPV were detected in 106 (98.15%) specimens i.e type 16, 18, 33, 35, 39, 45, 51, 52, 53, 58, 59, 66, but 2 specimens (1.85%) of unknown type. The three most common type were 43 (39.8%) HPV type 16, 22 (20.4%) HPV type 18 and 11 (10.2%) HPV type 52. Multiple infections were 12 (11.1%). Study at Dr. Cipto Mangunkusumo Hospital, Jakarta in 74 cervical cancer specimen also found these three most common HPVs prevalence were 44% HPV type-16, 39% HPV type-18 dan 14% HPV type-52.⁴ Population-base study for HPV genotyping in Bali found the three most common HPV type were 18% HPV type 52, 15% HPV type-16 dan 12% HPV type-18.¹⁰

Immunologic factor, genetic, environment and behavior have been implicated in development of HPV infection.¹³ Persistent HPV infection is require for disease progression because it lead to genomic instability and resulting in cervical cancer formation.^{12,14,15} Dysfunctional immune response is likely increase this persistence.¹⁶ Several studies found that immune responses vary by HPV type.¹⁷⁻¹⁹

Cytokines are pleiotropic glycoproteins for cell survival, proliferation and cell activation. IL-6 is one of cytokines that has role in both innate and adaptive immunity.^{16,20} In this study the risk of high IL-6 expression in cervical carcinoma infected by HPV type-52 was statistically significant 6-fold higher compare with cervical carcinoma infected by HPV type 16 (RO = 6.00; CI 95% = 1.13-31.99; p = 0.03; p < 0.05) and HPV type 18 (RO = 6.00; CI 95% = 1.13-31.99; p = 0.03; p < 0.05). IL-6 increases the regulation of *vascular endothelial growth factor-A* (VEGF-A) protein, which plays role in tumor angiogenesis so increase tumor development. IL-6 is a tumor angiogenesis

modulator.^{21,22} IL-6 increases cells tumor invasion by affecting endothelial cell permeability, cause by decrease in endothel gap junction endothel, rearrangement of actin fibers, resulting in changes in cell shape so malignant cells easier to invasion.²³ IL-6 also has role as antiapoptotic through increases the expression of Mcl-1, the family of Bcl-2 via PI3-K/Akt pathway (Wei dkk., 2001a),²⁴ increases tumor cell motility and decreases intercellular adhesion.^{25,26}

Cell-mediated immunity (CMI) especially CD4⁺ and CD8⁺ T lymphocytes play important role in adaptive immunity.^{27,28} Tumor infiltrating lymphocyte (TIL) in cervical carcinoma, the ability of persistence and progression were vary depend on HPV type.^{18,29} HPV immunity is genotype specific. ³⁰ In this study the risk of low CD4⁺ T lymphocyte expression in cervical carcinoma infected by HPV type-52 was statistically significant 6-fold higher and 7.43 fold higher respectively compare to cervical carcinoma infected by HPV type16 (RO = 6.00 ; CI 95% = 1.003-35.91; p = 0.04; p < 0.05) and HPV type 18 (RO = 7.43 ; CI 95% = 1.23-45.01; p = 0.02; p < 0.05). The risk of low CD8⁺ Tlymphocyte expression in cervical carcinoma infected by HPV type-52 was statistically significant 13.5 fold higher and 11 fold higher, respectively compare to cervical carcinoma infected by HPV type 16 (RO = 13.50; CI 95% = 1.42-128.26; p = 0.01; p < 0.05) and HPV type 18 (RO = 11.00 ; CI 95% = 1.16-103.94; p = 0.02; p < 0.05). The role of this immunologic factor may occur in various mechanisms.

HPV may evade cellular immune responses by decreases expression of MHC I and II.²⁷ MHC class II polymorphisms have positive correlation with cervical cancer. This polymorphisms cause genetic susceptibility and increase HPV infection persistence and evade the tumor cells from host immune recognition.^{13,31} Normal formation and presentation of HLA-I at cell surface for normal response of T cell antiviral need various components of antigen processing machinary (APM).³² Genetic variation in APM components are also significantly correlated with risk of cervical carcinoma.³³ To be full activated, T cells require costimulation with nonspecific molecule. Without this molecule T-cell may be come anergic and tolerant to HPV.¹⁶ Effective immune response also limited cause by late expression of protein viral that occur in terminal differentiation of squamous epithelial cells.²⁷ The role of viral protein E6 and E7 have been shown to be the main role in development of cervical carcinoma infected by HPV type 16 and 18 by inactivation of tumor suppressor gene p53 and pRb.^{8,11,12} Cervical carcinoma infected by HPV type-52 has different E2 disruption profile from HPV type-16. In HPV type 52 the disruption most common occur at N-terminal region, whereas in HPV type 16 at H region. This E2 gene disruption pattern of HPV type 52 could be type specific that occurred early in the course of infection and may be as initiators of oncogenic progression.³⁴

High risk HPV infection is the major risk factor for developing cervical carcinoma but other minor risk factors or co-factors also contributes for disease progression, such as age, parity, sexual activity, race, sexually transmitted infection, multiple sexual partner, alcohol, smoking, contraception, low socio economic, etc.^{10,16,35,36} In this study no significance different between cases and controls group in mean-age, parity and sexual activity (p > 0.05). It means that this control variable not influence the result of IL-6, CD4⁺ and CD8⁺ - lymphocytes expression.

REFERENCES

- 1. World Health Organization. Comprehensive Cervical Cancer Control: A Guide to Essential Practice. Geneva; 2006. p .13-42, 79-101.
- 2. Mu'nozi N, Bosch FX, Castellsagu'e X, Di'az M, de Sanjose S, Hammouda D, *et al*. Against Which Human Papillomavirus Types Shall We Vaccinate and Screen? The International Perspective. Int. J. Cancer 2004; 111: 278–285.
- 3. Tavassoli FA, Devilee P. World Health Organization Classification of Tumours. Pathology and Genetics of Tumours of the Breast and Female Genital Organs. Lyon, IARC Press; 2003. p. 260-271.
- Schellekens MC, Dijkman A, Aziz MF, Siregar B, Cornain S, Kolkman-Uljee S, *et al.* Prevalence of Single and Multiple HPV Types in Cervical Carcinomas in Jakarta, Indonesia. Gynecol Oncol 2004; 93: 49–53.
- Yugawa T, Kiyono T. Molecular Mechanisms of Cervical Carcinogenesis by High-Risk Human papillomavirus : Novel Function of E6 and E7 Oncoprotreins. Rev. Med. Virol 2009; 19 : 97-113.
- 6. Dirjen Yanmed Depkes R.I. 2013. Kanker di Indonesia Tahun 2009. Data Histopatologik.
- 7. Saito MN, Kiyono T. Basic Mechanisms of High-risk Human papillomavirus Induced Carcinogenesis: Roles of E6 and E7 Proteins. Cancer Sci 2007; 98 (10): 1505-1511.
- 8. Wright TC, Ronnett BM, Kurman RJ, Ferenczy A. Precancerous Lesions of the Cervix. In : Kurman RJ, Ellenson LH, Ronnett BM, editors. Blaustein's Pathology of the Female Genital Tract. Sixth edition. New York, Springer; 2011. p.194-252.
- 9. Castellsagué X, Díaz M, de Sanjosé S, Muñoz N, Herrero R, Franceschi S, *et al.* Worldwide Human Papillomavirus Etiology of Cervical Adenocarcinoma and Its Cofactors: Implications for

Screening and Prevention. J Natl Cancer Inst 2006; 98 (5):303-315.

- Vet JNI, de Boer MA, van den Akker BEWM, Lisnawati, Budiningsih S, Tyasmorowati D, et al. Prevalence of Human papillomavirus in Indonesia : A Population-Based Study in Three Regions. Br J Cancer 2008; 99(1) : 214–218.
- Turan T, Kalantari M, Cuschieri, Cubie HA, Skomedal H, Bernard HU. High-Throughput Detection of Human Papillomavirus-18 L1 Gene Methylation, A Candidate Biomarker for The Progression of Cervical Neoplasia. Virol 2007; 361:185–93.
- Senba M, Mori N. Mechanisms of Virus Immune Evasion Lead to Development from Chronic Inflammation to Cancer Formation Associated with Human Papillomavirus Infection. Oncol Rev 2012; 6:e17: 135-44.
- Maciag PC, Schlecht NF, Souza PSA. Major Histocompatibility Complex Class II Polymorphisms and Risk of Cervical Cancer and Human Papillomavirus Infection in Brazilian Women. Cancer Epidemiol Biomarkers Prev 2000; 9:1183-91.
- 14. Aho J, Hankins C, Tremblay C, Forest P, Pourreaux K, Rouah F, *et al.* Genomic Polymorphism of Human Papillomavirus Type 52 Predisposes toward Persistent Infection in Sexually Active Women. JID 2004; 190: 46-52.
- Koshiol J, Linds, L, Pimenta JM, Poole C, Jenkins D, Smith JS. Persistent Human Papillomavirus Infection and Cervical Neoplasia: A Systematic Review and Meta-Analysis. Am J Epidemiol 2008; 168:123–37.
- Koshiol J, Kovacic MB. Cytokines and Markers of Immune Response to HPV Infection. Available from : www.intechopen.com. Diakses tanggal 19 Agustus 2013.
- Fernandes APM, Gonçalves MAG, Duarte G, Cunha FQ, Simo[~]es RT, Donadi EA. HPV16, HPV18, and HIV Infection May Influence Cervical Cytokine Intralesional Levels. Virology 2005; 334(2): 294-8.
- Butsch Kovacic M, Katki HA, Kreimer AR, Sherman ME. Epidemiologic Analysis of Histologic Cervical Inflammation: Relationship to Human Papillomavirus Infections. Human Pathology 2008; 39(7): 1088-95.
- Ovestad IT, Vennestrøm U, Andersen L, Gudlaugsson E, Munk AC, Malpica A, et al. Comparison of Different Commercial Methods for HPV Detection in Follow-Up Cytology After ASCUS/LSIL, Prediction of CIN2-3 in Follow-Up Biopsies and Spontaneous Regression of CIN2-3. Gynecol Oncol. 2011; 123(2):278-83.

- 20. Abbas AK, Lichtman AH, Pillai S. Cellular and Molecular Immunology. Sixth edition. Philadelphia : Saunders. 2010. p. 3-17, 145-146, 189-198, 287-289, 362-365, 406-10.
- 21. Wei LH, Ku, ML, Chen CA, Cheng WF, Cheng SP, Hsieh, FJ. Interleukin-6 in Cervical Cancer : The Relationship with Vascular Endothelial Growth Factor. Gynecol Oncol 2001b; 82 (Issue 1) : 49-56.
- 22. Su JL, Lai KP, Chen CA, Yang CY, Chen PS, Chang CC, et al. A Novel Peptide Specifically Binding to Interleukin-6 Receptor (gp80) Inhibits Angiogenesis and Tumor Growth. Clin Cancer Res 2010.
- D'Anna R, Le Buanec H, Alessandri G, Caruso A, Burny A, Gallo R, *et al.* Selective Activation of Cervical Microvascular Endothelial Cells by Human Papillomavirus 16-E7 Oncoprotein. J Natl Cancer Inst 2001; 93 (24) : 1843-51.
- 24. Wei LH, Kuo ML, Chen CA, Chou CH, Cheng WF, Chang MC, *et al.* The Anti-Apoptotic Role of Interleukin-6 in Human Cervical Cancer is Mediated by Up-Regulation of Mcl-1 Through a PI 3-K/Akt Pathway. Oncogene 2001a; 20 (41) : 5799-809.
- Srivani R, Nagarajan B. A Prognostic Insight on In Vivo Expression of Interleukin-6 in Uterine Cervical Cancer. Int J Gynecol Cancer 2003; 13 (Issue 3) : 331-9.
- 26. Song SH, Lee JK, Seok OS, Saw HS. The Relationship Between Cytokines and HPV-16, HPV-16 E6, E7, and High-Risk HPV Viral Load in The Uterine Cervix. Gynecol Oncol 2007; 104 (3) : 732-8.
- 27. Scott M, Nakagawa M, Moscicki AB. Cell-Mediated Immune Response to Human Papillomavirus Infection. Clin Diagn Lab Immunol 2001; 8 (2) : 209–20.
- 28. Malik ZA, Hailpern SM, Burk RD. Persistent Antibodies to HPV Virus Like Particles Following Natural Infection Are Protective Against Subsequent Cervicovaginal Infection with Related and Unrelated HPV. Viral Immunol 2009; 22 (6): 445-9.
- 29. Lurchachaiwong W, Junyangdikul P, Payungporn S, Sampatanukul P, Chansaenroj J, Tresukosol D, *et al.* Human Papillomavirus Genotypes Among Infected Thai Women with Different Cytological Findings by Analysis of E1 Genes. New Microbiol 2011; 34:147-56.
- 30. Lindemann MLM, Calvo JMS, de Antonio JC, San I, Diaz E, Rubio MD, *et al.* Prevalence and Distribution of High-Risk Genotypes of HPV in Women with Severe Cervical Lesions in Madrid, Spain: Importance of Detecting Genotype 16 and Other High-Risk Genotypes. SAGE-Hindawi Access to Research. Adv Prev Med 2011.

- 31. Zehbe I, H€ohn H, Henryk PH, Neukirc C, Freitag K, Maeurer MJ. Differential MHC Class II Component Expression in HPV-Positive Cervical Cancer Cells: Implication for Immune Surveillance. Int. J. Cancer 2005; 117, 807–15
- 32. Hasim A, Abudula M, Aimiduo R, Ma JQ, Jiao Z, Akula G, et al. Post-Transcriptional and Epigenetic Regulation of Antigen Processing Machinery (APM) Components and HLA-I in Cervical Cancers from Uighur Women. PLoS ON 2011; 7(9): e44952.
- 33. Mehta AM, Jordanova ES, Kenter GG, Ferrone S, Fleuren GJ. Association of Antigen Processing Machinery and HLA Class I Defects with Clinicopathological Outcome in Cervical Carcinoma. Cancer Immunol Immunother 2008; 57(2): 197–206.
- 34. Cheung JLK, Cheung TH, Tang JWT, Chan PKS. Increase of Integration Events and Infection Loads of Human Papillomavirus Type 52 with Lesion Severity from Low-Grade Cervical Lesion to Invasive Cancer. J Clin Microbiol 2008; 46(4) : 1356-62.
- 35. Yang L, Li N, Guo LW, Li Q, Cui H, Dai M. Prevalence of Human Papillomavirus and Analysis of It's Risk Factors in Daqing City, Heilongjiang Province in 2010. Zhonghua Yu Fang Yi Xue Za Zhi. 2013; 47(2):118-23.
- 36. Zhang R, Shi TY, Ren Y, Lu H, Wei ZH, Hou WJ, et al. Risk Factors for Human Papillomavirus Infection in Shanghai Suburbs: A Population-Based Study with 10,000 Women. J Clin Virol. 2013; 13 : 217.

