Chromatogram Profiles and Cytotoxic Activity of Irradiated *Mahkota Dewa* (*Phaleria Macrocarpa* (Scheff.) Boerl) Leaves

E. Katrin^{1*}, Selvie² and H. Winarno¹

 ¹Center for the Application of Isotopes and Radiation Technology, National Nuclear Energy Agency Jl. Lebak Bulus Raya No. 49, Jakarta 12440, Indonesia
 ²Faculty of Pharmacy - Pancasila University, Srengseng Sawah, Jagakarsa, Jakarta 12640, Indonesia

ARTICLE INFO

Article history: Received 20 November 2010 Received in Revised form 06 April 2011 Accepted 14 April 2011

Keywords: Mahkota dewa leaves Phaleria macrocarpa Gamma irradiation Cytotoxicity Leukemia L1210 HPLC

ABSTRACT

Gamma irradiation has been used by the industries for preservation of herbal medicine, but it has not been studied the effect of gamma irradiation on their efficacy, especially their bioactivity as anticancer substances. The purpose of this research was to study the effect of gamma irradiation on the mahkota dewa leaves which has been claimed to contain potent anticancer substances. Maceration of dried mahkota dewa leaves successively with n-hexane, ethyl acetate, and ethanol gave crude extracts which the ethyl acetate was the most cytotoxic extract against leukemia L1210 cells with an inhibition concentration fifty (IC50) value of 10.3 µg/ml. Further separation of ethyl acetate extract by column chromatograph gave 7 fractions, and fraction 2 showed the most cytotoxic fraction exhibited the most cytotoxic extract against leukemia L1210 cells with an IC₅₀ value of 1.9 µg/ml. Since, the fraction 2 of ethyl acetate extract was the most potent fraction, the irradiated samples were treated with the same procedure as treatment of fraction 2 from control sample. Cytotoxic activity test of fractions 2 from irradiated samples showed that the cytotoxic activity decreased depending on increasing of irradiation dose. Gamma irradiation dose up to 7.5 kGy on mahkota dewa leaves could decreased the cytotoxic activity of fraction 2 as the most cytotoxic-potential fraction against leukemia L1210 cells, but decreasing the cytotoxic activity has not exceeded the limit of the fraction declared inactive. So that the irradiation dose up to 7.5 kGy can be used for decontamination of bacteria and fungus/yeast without eliminating the cytotoxic activity. Gamma irradiation also caused changes in the thin layer chromatograph (TLC) spots and HPLC chromatograms profiles of fraction 2 which was the most cytotoxic fraction in ethyl acetate extract of mahkota dewa leaves against leukemia L1210 cells. One of the main peaks (peak 1) on HPLC chromatograms decreased with increasing the irradiation dose. In agree with the decreasing peak 1 in chromatogram of fraction 2 due to irradiation, cytotoxic activity of fraction 2 also decreased along with increasing doses of irradiation. It suggested that peak 1 is the component that contributes to the cytotoxic activity of fraction 2 in ethyl acetate extract of mahkota dewa leaves.

© 2011 Atom Indonesia. All rights reserved

INTRODUCTION

Mahkota dewa, Phaleria macrocarpa (Scheff.) Boerl (Thymelaeaceae) is one of the Indonesian medicinal plants which is recently popular due to its ability for treating various diseases, namely cancer, tumors, diabetes mellitus, hypertension, hepatitis, rheumatism, gout, skin diseases, kidney disorders, allergies, asthma, cold, haemorrhoid, stroke, and migraine [1,2,3]. Leaves and bark of *mahkota dewa* contained alkaloids,

* Corresponding author.

terpenoids, flavonoids, polyphenols, saponins, resins, lignin, and benzophenones [1,4,5].

The stems of *mahkota dewa* believed to be able to heal bone cancer, meanwhile eggshell of the fruits or seeds more efficacious compared with the bark and fruit flesh. Seeds of *mahkota dewa* is only used for external treatment especially skin diseases [6]. Some several benefits in the plant of *mahkota dewa* has been discovered, moreover the research to explore active compounds from *mahkota dewa* has been started. In 2003, it has been reported that one of the fraction of methanol extract from the leaves of *mahkota dewa* showed the cytotoxic activity against *Artemia salina* with the LC₅₀

E-mail address: erminkk@batan.go.id

(lethal concentration fifty) value of 1.04 μ g/ml. Furthermore, the same fraction also exhibited inhibitory activity against myeloma cells (NS-1) with an IC₅₀ value of 81 μ g/ml [7]. An IC₅₀ is the concentration of the tested samples that can inhibit cell proliferation as much as 50%. The same researcher has isolated 4,5-dihydroxy-4'methoxybenzophenone-3-*O*- β -D-glucoside so called phalerin from the methanol extract of *mahkota dewa* leaves [8]. Recently this structure has been revised by Oshimi [9] as 2,4,6-trihydroxy-4methoxybenzophenone-2-*O*- β -D-glucoside.

Chloroform extract of *mahkota dewa* leaf showed an antiproliferation activity against cervical cancer cells HeLa (IC_{50} 40.2 µg/ml), melanoma skin cancer cells HM3KO (IC_{50} 62.9 µg/ml) and breast cancer cell MCF-7 (IC_{50} 70.8 µg/ml) [10]. In the other hand, ethyl acetate extract of *mahkota dewa* bark showed the highest cytotoxic activity against L1210 leukemia cells compared with *n*-hexane extract and ethanol with an IC_{50} value of 10.2 µg/ml. Further fractionation of this ethyl acetate extract gave fraction 2 was the most active fraction with the IC_{50} value of 8.3 µg/ml [11]. The phalerin has also been isolated from the *mahkota dewa* bark [12].

In the recent years, the use of herbal medicines for complementary treatments of some diseases has been popular. While, dry herbs easily damaged during storage due to microbial contamination, cause reducing the efficacy and quality of herbal medicines. To overcome this problem, gamma irradiation can be used. Codex Alimentarius Commission stating that ionizing radiation has been used to decontaminate the pathogenic microbes and to extend the shelf life of herbs, foods, and spices with the maximum absorbed dose should not exceed 10 kGy, except when necessary to achieve legitimate technological purposes [13]. In Indonesia the regulation of the ionizing radiation for sterilization of products has been regulated by the Minister of Health No. 701/MENKES/PER/VIII/2009 [14]. Irradiation dose up to 10 kGy can reduce the number of bacteria on mahkota dewa dried powder from 10¹⁰ cfu/g to 10^5 cfu/g [15], this value is qualified according to The National Agency of Drug and Food Control (NADFC = BPOM) [16]. Gamma irradiation technique has several advantages including high penetrating energy, is processed at room temperature, leaving no residue, environmentally friendly, and can be packaged in the end products [13,16].

In spite of gamma irradiation has been used by some of the herbal medicine industries, but the effect of gamma irradiation on the efficacies has not been studied yet. In previous study, gamma irradiation with doses up to 7.5 kGy on the mahkota dewa bark could be used for decontamination of pathogenic microbes and to extend the self life without affecting the cytotoxic activity on L1210 leukemia cancer cells [15]. In this report, the samples which had been irradiated at the dose of 7.5 kGy were examined the bioactivity in against human cancer cell vitro lines. namely: cervical cancer cells HeLa, leukemia THP-1, lymphoma HUT-78, and lung carcinoma A-549. The aim of this study was to examine whether the irradiation dose of 7.5 kGy does not eliminate there efficacies as anticancer based on their bioactivity against human cancer cell lines.

EXPERIMENTAL METHODS

General information

Flush column chromatography was carried out on silica gel 60 (70 to 230 mesh ASTM, Merck). Thin-layer chromatography on silica gel 60 F_{254} plate (Merck) was used for monitoring and checking the fractions collected from column chromatography. The spots on TLC were detected by short (254 nm) and long wavelength (366 nm) ultraviolet light and visualized by spraving the plates with 1% Ce $(SO_4)_2$ in 10% aqueous sulfuric acid followed by heating. Concentrating of extracts and fractions were done by using a rotary evaporator in vacuo in temperature of 35°C followed by drying in oven desiccator in vacuo at 40°C until fix weight of the fractions were obtained. Analysis components of fractions by HPLC was done on a Shimadzu LC-6A equipped with UV-detector at 210 nm, C-18 reverse phase coloumn of varian microsorb MV 100-5 with dimension of 250 mm length and 4.6 mm inner diameter, and the comparison of mobile phase methanol to water was 7:3.

Materials and apparatus

The leaves of *mahkota dewa Phaleria macrocarpa* (Scheff.) Boerl were collected from Cibeuteung village, Parung, West Java, Indonesia in May 2006 as described in previous report, and determined by Herbarium Bogoriensis, Research Center for Biology, Indonesian Institute of Sciences, Bogor, Indonesia [11]. The leaves of *mahkota dewa* were then dried, sliced and blended.

The leukemia L210 cells used for cytotoxicity test was liquid cancer cells line isolated from DBA strain mouse induced by methylcholanthrene. Chemicals used in this study were *n*-hexane, ethyl acetate, ethanol, tryphan blue 0.4 %, RPMI Medium-1640, fetal bovine serum (FBS), liquid nitrogen, distilled water, aquabidest, chloroform, methanol, cerium sulfate, H_2SO_4 , silica gel 60 (70-230 mesh), celite 545, GF_{254} silica plate.

Apparatus used were Gamma Irradiator from cobalt-60 with an activity of 140 kCi, high performance liquid chromatograph (Shimadzu LC 6-A), column chromatograph (diameter 8 cm and length 90 cm), vacuum rotary evaporator (Büchi), oven (Prolabo), multi-well plate tissue's culture, haemocytometer Neubauer (0.100 mm \times 0.0025 mm²), hot plate, ultrasonicator, oven desiccator, analytical balance (Mettler Toledo), autoclave (S4-ON), incubator, microscope.

Preparation and irradiation of samples

The dried mahkota dewa leaves was powdered, then was weighed each 100 g and put in polyethylene bags and sealed with a sealer machine. Ten samples were preferred. The each two samples were irradiated at a dose of 0 (control), 5, 7.5, 10, and 15 kGy with dose rate of 10 kGy/h. Determination of the dose was done by Fricke solution as a primary standard dosimeter consisting of 0.001 Μ of ferrous ammonium sulfate and 0.001 M of sulfuric acid.

Preparation and separation of extracts by column chromatograph

The control and irradiated samples were macerated as much as four times, each time with 1.4 L of *n*-hexane. The residue was acetate again extracted with ethyl and then with ethanol with the same procedure as maceration with *n*-hexane. The each filtrate obtained was concentrated using a rotary evaporator followed by drying in oven desiccators in vacuo.

The most active extract against leukemia L1210 cell line was fractionated using silica gel column chromatograph. Amount of 1.0 g of extract was homogenized with celite, then subjected into column chromatograph and was gradiently eluted by n-hexane - ethyl acetate methanol (1:0:0, 5:1:0, 3:1:0, 0:1:0, 0:1:1, 0:0:1). Each fraction was collected with a volume of 150 ml. The fraction obtained was concentrated using a rotary evaporator followed by drying in oven desiccator in vacuo. The obtained fractions were analyzed by TLC and the fractions that had a similar spot pattern were merged.

Analysis of fractions by HPLC

The most active fractions from control and irradiated samples which have been examined against leukemia L1210 were then analyzed by high performance liquid chromatograph (HPLC). Amount of 1.6 mg fraction was dissolved in 1 ml methanol, filtered with membrane filter (0.45 μ m), then 20 μ l of solution was injected into the HPLC.

Cytotoxic activity test against mouse leukemia L1210 cell line [12]

Exponentially growing leukemia L1210 cells at a density of 2×10^5 cells/ml suspended in RPMI-1640 medium was placed into serocluster 24 wells plate, the sample solution of extracts or fractions in various concentrations were added then incubated at 37°C for 48 h under 5% CO₂ condition. The varving concentration of extracts were 0 (control), 5, 10, 20, 40, and 80 μ g/ml, meanwhile the varving concentration of fractions were 0(control), 1, 2, 4, 8, and 16 µg/ml. Control consisted of exposed to fresh medium only. Duplicate wells were prepared for each concentration of extracts or fractions and control. Ninety µl of cell suspension in every well was pipetted and put into serocluster plate and 10 µl of tryphan blue was added and the mixture was homogenized. Furthermore, 10 µl of suspension was put in haemocytometer and the amount of viable cells of the samples and control were enumerated by microscope. The inhibition percentage was calculated as the following equation.

% inhibition =
$$\frac{[\Sigma \text{ cells in control}] - [\Sigma \text{ cells in sample}]}{[\Sigma \text{ cells in control}]} \times 100\%$$

By making the graph of sample concentration in logarithm (*X* axes) *versus* probit of % inhibition (*Y* axes), the linear regression equation Y = aX + b is obtained. The inhibition concentration-fifty (IC₅₀) which expresses the ability of the samples to inhibit 50% of cancer cell proliferation is calculated by the substitution of *Y* by 5 (probit value of 50) to the linear regression equation Y = aX + b. Subsequently, IC₅₀ value = antilogarithm of $X = \{antilogarithm of (5-b)/a\}$ can be determined.

Data analysis

The data of analysis IC_{50} values and HPLC chromatogram for studying the effect of

gamma irradiation toward cytotoxic activity of the samples, was performed by one-way ANOVA using SPSS 17.0 *for windows* in the confidence level of 95%.

RESULTS AND DISCUSSION

100 Maceration from g of control and irradiated samples by *n*-hexane, ethvl acetate, and ethanol was shown in Table 1. It can be seen that the yield of *n*-hexane extracts from control and irradiated samples significantly change, did not but the of ethyl extracts vield acetate as well as ethanol extracts from irradiated doses samples at of 10 and 15 kGy, slightly increased. This is apparently due radiation energy causing the samples to become soft and brittle, making the chemical components was more easily extracted.

 Table 1. The yield of extracts from control and irradiated mahkota dewa leaves.

Dose (kGy)	yield of <i>n</i> -hexane extract (%)	yield of ethyl acetate extract (%)	yield of ethanol extract (%)
0	2.3	5.9	18.8
5	2.4	5.9	19.2
7.5	2.4	5.3	19.2
10	2.3	7.1	19.5
15	2.4	7.1	19.7

Furthermore, the extracts from irradiated sample) control sample (non then were assayed their cytotoxic activity against leukemia L1210 cells line. The obtained inhibition percentage value was converted to probit value and then performed linear curve was in versus log of ethyl acetate extract concentration as seen in Fig. 1. Inhibition concentration fifty (IC_{50}) value was calculated based on equation Y =the linear 5, then it was obtained the concentration as IC₅₀. The results showed all that extracts exhibited cytotoxic activity with an IC₅₀ < 30 µg/ml (Table 2).



Fig. 1. Curve of log of samples concentrations *versus* probit of inhibition percentage.

Table 2. Inhibition percentage of ethanol, *n*-hexane, and ethyl acetate extracts from control sample against leukemia L1210 cells and its IC_{50} value.

Sample	Sample concentration (µg/ml)	Inhibition (%)	IC ₅₀ (μg/ml); regression eq.; linearity value, R
ethanol extract	5	28.2	24.1
	10	59.7	21.1
	20	61.8	Y = 1.771 X + 3.238 R = 0.979
	40	85.4	
	80	95.4	
<i>n</i> -hexane extract	5	27.4	12.4
	10	43.4	12.4
	20	59.7	Y = 1.678 X + 3.175 R = 0.995
	40	83.3	
	80	91.3	
ethyl acetate extract	5	8.2	10.3
	10	18.0	10.5
	20	37.6	Y = 2.432 X + 1.764 R = 0.983
	40	64.5	
	80	90.6	

According to the American National Cancer Institute quoted by Nor Aini *et al.* [17] and Magalhaes *et al.* [18], an extract exhibits a potent anticancer if the IC₅₀ value < 30 µg/ml, while according to Mans *et al.* [19], an extract exhibits a potent anticancer if the IC₅₀ value < 50 µg/ml. Among the extracts, the ethyl acetate extract gave highest cytotoxicity against leukemia L1210 cells (IC₅₀ 10.3 µg/ml) compared to *n*-hexane extract (12.4 µg/ml) and ethanol extract (IC₅₀ 24.1 µg/ml), so in this experiment, ethyl acetate extract has been chosen to be further separation using column chromatograph. Subsequently, to study the effect of irradiation onto the leaves of dewa. acetate mahkota the ethyl extract from irradiated sample were then assaved separation using column and also further chromatograph the as same manner as treatment of the control sample.

Fractionation of ethyl acetate extract from control sample gave 7 fractions (Fr.1-Fr.7). Furthermore the Fr.1-Fr.7 were then assayed against leukemia L1210 cells as the same manner as bioassay of extracts, gave IC_{50} values as presented in Table 3. Since, Fr.2 was showed the lowest IC_{50} value $(1.9 \ \mu g/ml)$ (Table 3), it indicated Fr.2 was the most active fraction the which exhibited cytotoxic activity against L1210 leukemia cells. so assayed the fractions from irradiated samples was done toward only Fr.2.

Table 3. The yield and IC_{50} value Fr.1-Fr.7 of ethyl acetate extract from control (non irradiated) sample against leukemia L1210 cells.

Sample _	Yield		IC (ug/ml)	
Sample -	mg	%	$-1C_{50}(\mu g/IIII)$	
Fr.1	6	0.6	4.2	
Fr.2	21	2.1	1.9	
Fr.3	18	1.8	4.3	
Fr.4	174	17.4	4.4	
Fr.5	72	7.2	5.6	
Fr.6	676	67.6	5.0	
Fr.7	30	3.0	5.0	

Table 4 showed the comparison of IC_{50} values of ethyl acetate extracts and Fr.2 control and irradiated samples. from Based on the IC₅₀ values of ethyl acetate extract and Fr.2, it appeared that gamma irradiation tends to cause the IC₅₀ value increased. which the means cytotoxic activity decreased. but the increasing of IC₅₀ value at irradiation dose until 15 kGy has not exceeded the limit of an extract and the fraction was declared inactive [17]. Nevertheless, irradiation at the dose of 7.5 kGy caused the IC_{50} values of Fr.2 significantly increased. So it is recommended

irradiation of mahkota that the dewa leaves for decontamination of bacteria and and fungus/yeast for preserving and extending its self life without eliminating the cytotoxic activity was no more than 7.5 kGy.

Table 4. IC_{50} comparison of ethyl acetate extract and Fr.2 from control and from irradiated samples.

Sampla	IC ₅₀ (µg/ml)				
Sample	control	5 kGy	7.5 kGy	10 kGy	15 kGy
ethyl acetate extract	10.3 ^a	14.7 ^{b,c}	14.2 ^{b,c}	13.2 ^{a,b}	17.1 °
Fr.2	1.9 ^a	2.4 ^{ab}	3.1 ^b	4.0°	5.8 ^d

Note: Similar letters as superscript indicate that they were not significantly difference.

effect of Furthermore, to study the irradiation on the components gamma in the Fr.2, the TLC spot and presence HPLC chromatogram profiles of Fr.2 from control and irradiated samples were compared. Figure 2 shows the TLC spots profile of Fr.2 from control and irradiated samples. It appeared that at least 8 spots included in the Fr.2 from control sample (0 kGy). Irradiation dose of 7.5, 10, and 15 kGy caused spot 3 significantly decreased, meanwhile spots 7 and 8 seem to be increased at irradiation dose of 15 kGy. The decreasing of spot 3 means those compounds degraded to another compounds, probably become compound included in the spots 7 and 8. It suggested that the spot 3 contained the active substances which have cytotoxic activity against leukemia L1210 cells.



Fig. 2. TLC spots profile of Fr.2 from control and irradiated samples.



Fig. 3. HPLC chromatogram profile of Fr.2 from control and irradiated samples.



Fig. 4. The effect of irradiation dose of peak 1 and peak 2 of HPLC chromatogram profile of Fr.2.

Further analysis of Fr.2 by HPLC equipped with UV detector at 210 nm showed that there was major peak at retention time of 8.8 min (peak 1) and other minor peaks, such as peak 2 at retention time of 23.6 min (Fig. 3). Peak 1 significantly decreased with increasing the irradiation dose by follows the quadratic equation $(Y = -0.26 X^2 + 0.03 X + 101)$, meanwhile peak 2 was increased with increasing the irradiation dose by follows the quadratic equation $(Y = 0.19 X^2 + 7.52 X + 3.98)$. The changed of peaks 1 and 2 was shown in Fig. 4.

Reducing peak 1 in HPLC chromatogram of Fr.2 as well as spot 3 in TLC spot profile is likely correlated with reducing cytotoxic activity of Fr.2 due to the increasing radiation dose. It also suggested that the peak 1 was the active substance which has cytotoxic activity against leukemia L1210 cells.

CONCLUSION

Gamma irradiation causes changes in the TLC spots and HPLC chromatograms profiles of fraction 2 (Fr.2) which was the most cytotoxic fraction in ethyl acetate extract of mahkota dewa leaves against leukemia L1210 cells. One of the main peak (peak 1) on HPLC chromatograms decreased, whereas the other peak (peak 2) which was a small peak in HPLC chromatogram of fraction 2 from non irradiated sample increased with increasing the irradiation dose. In agree with the decreasing peak 1 in chromatogram of fraction 2 due to irradiation, cytotoxic activity of fraction 2 also decreased along with increasing doses of irradiation. It suggested that peak 1 is the component that contributes to the cytotoxic activity of fraction 2 in ethyl acetate extract of *mahkota dewa* leaves.

Gamma irradiation dose of 7.5 kGy on *mahkota dewa* leaves could decreased the cytotoxic activity of fraction 2 as the most cytotoxic-potential fraction against leukemia L1210 cells, but decreasing the cytotoxic activity has not exceeded the limit of the fraction declared inactive. So that the irradiation dose up to 7.5 kGy can be use for decontamination of bacteria and fungus/yeast without eliminating the cytotoxic activity.

ACKNOWLEDGEMENTS

The authors would like to thank Mr. Susanto for doing bioassay experiment using leukemia L1210 cell line. Special thanks are also addressed to Mr. Tjahyono, S.P. and all members of Irradiation Facilities-PATIR-BATAN who helped to irradiate the samples.

REFERENCES

 J.R. Hutapea, Inventaris Tanaman Obat Indonesia, Jilid V, Badan Penelitian dan Pengembangan Kesehatan - Departemen Kesehatan Republik Indonesia, Jakarta (1999) 3.

- 2. Kardinan and A. Taryono, Tanaman Obat Penggempur Kanker, Cetakan Pertama, Agromedia Pustaka, Jakarta (2003) 12.
- 3. S. Dalimarta, Atlas Tumbuhan Obat Indonesia, Jilid III, Puspa Swara, Jakarta (2003) 62.
- N. Harmanto, Mahkota Dewa Obat Pusaka Para Dewa, Cetakan Kedua, Agromedia Pustaka, Jakarta (2001) 1.
- W.P. Winarto, Mahkota Dewa, Budi Daya & Pemanfaatan Obat, Cetakan Kelima, Penebar Swadaya, Jakarta (2003) 3.
- 6. N. Harmanto, Sehat dengan Ramuan Tradisional Mahkota Dewa, Edisi Pertama, Agromedika Pustaka, Tangerang (2001) 31.
- M.S.H. Wahyuningsih, S. Mubarika, W.T. Artama and S. Wahyuono, Research Reports of Faculty of Pharmacy, Gadjah Mada University, Yogyakarta (2003).
- M.S.H. Wahyuningsih, S. Mubarika, I.G. Gandjar, M.T. Hamann, K.V. Rao and S. Wahyuono, Indonesian J. Pharmacy. 16 (2005) 51.
- S. Oshimi, K. Zaima, Y. Matsuno, Y. Hirasawa, T. Izuka, H. Studiawan and G. Indrayanto, N.C. Zaini and H. Morita, J. Nat. Med. 62 (2008) 207.
- M.S.H. Wahyuningsih, S. Mubarika, W.T. Artama, S. Wahyuono and I.G. Ganjar, J. Traditional Medicine 10 (2005) 5.
- 11. E. Katrin and H. Winarno, J. Traditional Medicine, **13** (2008) 120. (in Indonesian).

- 12. H. Winarno and E. Katrin, Indo. J. Chem. 9 (2009) 142.
- Codex Alimentarius Commission, Available at http://www.codexalimentarius.net/download/re port/555/a183_12e.pdf. Retrieved August 26 (2008).
- Anonymous, The Minister of Health Regulation, The Republic of Indonesia No. 701/MENKES/PER/VIII/2009. Available at http://www.hukor.depkes.go.id/permenkes/ PMK No. 701ttg Pangan Iradiasi.pdf. Retrieved May 10 (2010). (in Indonesian).
- H. Winarno, Wisnurahadi, S.R. Tamat and E. Katrin, Sci. J. App. Isot. and Radiat. 6 (2010) 1. (in Indonesian).
- 16. Z. Irawati, *Application of Electron Beam Machine in Foods Industries*, Proceeding of Scientific Meeting on Accelerator Technology and its Applications, BATAN, Jakarta (2006) 87. (in Indonesian).
- A.S. Nor Aini, A. Merrina, J. Stanslas and S. Sreeramanan, Int. J. Cancer Res. 4 (2008) 103.
- H.I.F. Magalhaes, P.M.P. Ferreira, E.S. Moura, M.R. Torres, A.P.N.N. Alves, O.D.L. Pessoa, L.V. Costa-Lotufo, M.O. Moraes and C. Pessoa, An. Acad. Bras. Cienc. 82 (2010) 407.
- D.R.A. Mans, A.B. Rocha and G. Schwartsmann, The Oncologist 5 (2000) 185.